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⑤④ **Methods for culturing HCV in eukaryotic cells.**

⑤⑦ The present invention is directed to cells that may be used to propagate HCV in culture. The present invention is also directed to viral antigens produced by such cultures, and the use such antigens, for example, in diagnostics.

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METHODS FOR CULTURING HCV IN EUKARYOTIC CELLS

Technical Field

The invention relates to *in vitro* cell systems for the replication of a virus; more specifically, it relates to cell lines in which an etiologic agent of non-A non-B hepatitis (NANBH), hepatitis C virus (HCV) will replicate.

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Background

Non-A, Non-B hepatitis (NANBH) is a transmissible disease or family of diseases that are believed to be viral-induced, and that are distinguishable from other forms of viral-associated liver diseases, including that caused by the known hepatitis viruses, i.e., hepatitis A virus (HAV), hepatitis B virus (HBV), and delta

hepatitis virus (HDV), as well as the hepatitis induced by cytomegalovirus (CMV) or Epstein-Barr virus (EBV). NANBH was first identified in transfused individuals. Transmission from man to chimpanzee and serial passage in chimpanzees provided evidence that NANBH is due to a transmissible infectious agent or agents. A viral agent, HCV, has been identified as an etiologic agent of NANBH.

5 The demand for sensitive, specific methods for screening and identifying carriers of NANBV and NANBV contaminated blood or blood products is significant. Post-transfusion hepatitis (PTH) occurs in approximately 10% of transfused patients, and NANBH accounts for up to 90% of these cases. The major problem in this disease is the frequent progression to chronic liver damage (25-55%).

10 Patient care as well as the prevention of transmission of NANBH by blood and blood products or by close personal contact require reliable diagnostic and prognostic tools to detect nucleic acids, antigens and antibodies related to NANBV. In addition, there is also a need for effective vaccines and immunotherapeutic therapeutic agents for the prevention and/or treatment of the disease.

15 Tissue culture cells infected with an etiologic agent which causes NANBH would provide a source of viral polypeptides which could be used for immunoassays and/or for vaccines. In addition, they could be useful in drug screening programs in the development of antiviral agents for treating the disease.

Immortalized human or non-human primate cell lines which are derived from liver, and are purportedly infected with an etiologic agent causative of NANBH have been reported in WO/87/05930, filed 1 April 1987, and published 8 October 1987. These cell lines are reportedly non-lymphocytic in nature. In addition, it has not been established that the agent which replicates in these cells is causative of the disease.

20 It has also been reported that a preparation of mononuclear leukocytes, which purportedly were mainly lymphocytes, isolated from a patient with NANBH, caused NANB when infused into a susceptible chimpanzee. Hellings et al. (1985). However, proof of NANBH infection was indirect, in that it rested primarily on morphological and histological changes in the purportedly infected chimpanzee. Moreover, only one of the two chimpanzees receiving the blood cells was purportedly infected.

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Disclosure of the Invention

30 The present invention is directed to cells that may be used to propagate HCV in culture. The present invention is also directed to viral antigens produced by such cultures, and use such antigens, for example, in diagnostics.

Accordingly, one aspect of the invention is a cell line infected with HCV.

35 Another aspect of the invention is a composition for the detection of an anti-HCV antibody comprised of an antigen from an *in vitro* cell infected with HCV, wherein the antigen is affixed to a solid substrate, wherein the antigen is immunologically reactive with said anti-HCV antibody.

Yet another aspect of the invention is an immunoassay for detecting antibodies directed against an HCV antigen comprising:

- (a) incubating a sample suspected of containing anti-HCV antibodies with an HCV antigen produced by an *in vitro* cell line, under conditions which allow the formation of an antibody-antigen complex; and
- 40 (b) detecting the antibody-antigen complex containing the HCV antigen.

Another aspect of the invention is a method for screening for an agent which inhibits HCV

Still another aspect of the invention is a method for proliferating virus comprising growing in culture HCV infected cells.

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Brief Description of the Drawings

Figure 1 shows a composite sequence of the HCV cDNA sense strand derived from HCV strain HCV1.

Figure 2 shows the nucleotide sequence of HCV cDNA clone 35, and the amino acids encoded therein.

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Modes for Carrying Out the Invention

I. Definitions

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The term "hepatitis C virus" has been reserved by workers in the field for an heretofore unknown etiologic agent of NANBH. Accordingly, as used herein, "hepatitis C virus" (HCV) refers to an agent causative of NANBH, which was formerly referred to as NANBV and/or BB-NANBV. The terms HCV,

NANBV, and BB-NANBV are used interchangeably herein. As an extension of this terminology, the disease caused by HCV, formerly called NANB hepatitis (NANBH), is called hepatitis C. The terms NANBH and hepatitis C may be used interchangeably herein.

The term "HCV", as used herein, denotes a viral species of which pathogenic strains cause NANBH, and attenuated strains or defective interfering particles derived therefrom. The HCV genome is comprised of RNA. It is known that RNA containing viruses have relatively high rates of spontaneous mutation, i.e., reportedly on the order of 10^{-3} to 10^{-4} per incorporated nucleotide (Fields & Knipe (1986)). Therefore, there can be multiple strains, which may be virulent or avirulent, within the HCV species. It is believed that HCV is a Flavi-like virus. The morphology and composition of Flavivirus particles are known, and are discussed in Brinton (1986). Generally, with respect to morphology, Flaviviruses contain a central nucleocapsid surrounded by a lipid bilayer. Virions are spherical and have a diameter of about 40-50 nm. Their cores are about 25-30 nm in diameter. Along the outer surface of the virion envelope are projections that are about 5-10 nm long with terminal knobs about 2 nm in diameter.

cDNAs to the genome of a strain of HCV, denoted HCV1, are described in U.S.S.N. 355,002 (Attorney Docket 2300-0063.28), filed May 18, 1989, which is commonly owned by the herein assignee, and which is in its entirety incorporated herein by reference. See also EPO Publication No. 0 318 216 and PCT App. No. US88/04125, which are also incorporated herein by reference. A composite sequence of the HCV cDNA sense strand is shown in Figure 1. Different strains or isolates of HCV are expected to contain variations at the amino acid and nucleic acids levels compared with HCV1. Many isolates are expected to show much (i.e., more than about 40%) homology in the total amino acid sequence compared with HCV1. However, it may also be found that other less homologous HCV isolates would be defined as HCV strains according to various criteria such as an ORF encoding a polypeptide similar in size to that of HCV1, an encoded polypeptide of similar hydrophobic and antigenic character to that of HCV1, and the presence of co-linear peptide sequences that are conserved with HCV1.

HCV encodes at least one epitope which is immunologically identifiable with an epitope encoded in the HCV1 genome. The epitope is unique to HCV when compared to other known Flaviviruses. The uniqueness of the epitope may be determined by its immunological reactivity with anti-HCV antibodies and lack of immunological reactivity with antibodies to other Flavivirus species. Methods for determining immunological reactivity are known in the art, for example, by radioimmunoassay, by ELISA assay, and by hemagglutination. Examples of suitable techniques for determining immunological reactivity of HCV epitopes are described in U.S.S.N. 07/341,334, which is commonly owned by the herein assignee, in Choo et al. (1989), and in Kuo et al. (1989).

In addition to the above, the following parameters of nucleic acid homology and amino acid homology are applicable, either alone or in combination, in identifying a strain or isolate as HCV. Since HCV strains and isolates are evolutionarily related, it is expected that the overall homology of the genomes at the nucleotide level probably will be about 40% or greater, probably about 60% or greater, and even more probably about 80% or greater; and in addition that there will be corresponding contiguous sequences of at least about 13 nucleotides. The correspondence between the putative HCV strain genomic sequence and the HCV1 cDNA sequence can be determined by techniques known in the art. For example, they can be determined by a direct comparison of the sequence information of the polynucleotide from the putative HCV, and the HCV cDNA sequence(s) described herein. For example, also, they can be determined by hybridization of the polynucleotides under conditions which form stable duplexes between homologous regions (for example, those which would be used prior to S_1 digestion), followed by digestion with single stranded specific nuclease(s), followed by size determination of the digested fragments.

Because of the evolutionary relationship of the strains or isolates of HCV, putative HCV strains or isolates are identifiable by their homology at the polypeptide level. Generally, HCV strains or isolates are expected to be more than about 40% homologous, probably more than about 70% homologous, and even more probably more than about 80% homologous, and some may even be more than about 90% homologous at the polypeptide level. The techniques for determining amino acid sequence homology are known in the art. For example, the amino acid sequence may be determined directly and compared to the sequences provided herein. Alternatively the nucleotide sequence of the genomic material of the putative HCV may be determined (usually via a cDNA intermediate), the amino acid sequence encoded therein can be determined, and the corresponding regions compared.

As used herein, a polynucleotide "derived from" a designated polynucleotide sequence refers to a polynucleotide sequence which is comprised of a sequence of approximately at least about 6 nucleotides, preferably at least about 8 nucleotides, more preferably at least about 10-12 nucleotides, and even more preferably at least about 15-20 nucleotides corresponding to a region of the designated nucleotide sequence. "Corresponding" means homologous to or complementary to the designated sequence. Prefer-

ably, the sequence of the region from which the polynucleotide is derived is homologous to or complementary to a sequence which is unique to an HCV genome. Whether or not a sequence is unique to the HCV genome can be determined by techniques known to those of skill in the art. For example, the sequence can be compared to sequences in databanks, e.g., Genbank, to determine whether it is present in the uninfected host or other organisms. The sequence can also be compared to the known sequences of other viral agents, including those which are known to induce hepatitis, e.g., HAV, HBV, and HDV, and to other members of the Flaviviridae. The correspondence or non-correspondence of the derived sequence to other sequences can also be determined by hybridization under the appropriate stringency conditions. Hybridization techniques for determining the complementarity of nucleic acid sequences are known in the art, and are discussed *infra*. See also, for example, Maniatis et al. (1982). In addition, mismatches of duplex polynucleotides formed by hybridization can be determined by known techniques, including for example, digestion with a nuclease such as S1 that specifically digests single-stranded areas in duplex polynucleotides. Regions from which typical DNA sequences may be "derived" include but are not limited to, for example, regions encoding specific epitopes, as well as non-transcribed and/or non-translated regions.

The derived polynucleotide is not necessarily physically derived from the nucleotide sequence shown, but may be generated in any manner, including for example, chemical synthesis or DNA replication or reverse transcription or transcription. In addition, combinations of regions corresponding to that of the designated sequence may be modified in ways known in the art to be consistent with an intended use.

The term "polynucleotide" as used herein refers to a polymeric form of nucleotides of any length, either ribonucleotides or deoxyribonucleotides. This term refers only to the primary structure of the molecule. Thus, this term includes double- and single-stranded DNA, as well as double- and single stranded RNA. It also includes modified, for example, methylation capping and/or by other modifications known in the art, including the substitution of an analog for one or more of the naturally occurring nucleotide bases, as well as unmodified forms of the polynucleotide.

The term "purified viral polynucleotide" refers to a viral genome or fragment thereof which is essentially free, i.e., contains less than about 50%, preferably less than about 70%, and even more preferably less than about 90% of polypeptides with which the viral polynucleotide is naturally associated. Techniques for purifying viral polynucleotides from viral particles are known in the art, and include for example, disruption of the particle with a chaotropic agent, differential extraction and separation of the polynucleotide(s) and polypeptides by ion-exchange chromatography, affinity chromatography, and sedimentation according to density.

An "open reading frame" (ORF) is a region of a polynucleotide sequence which encodes a polypeptide; this region may represent a portion of a coding sequence or a total coding sequence.

A "coding sequence" is a polynucleotide sequence which is transcribed into mRNA and/or translated into a polypeptide when placed under the control of appropriate regulatory sequences. The boundaries of the coding sequence are determined by a translation start codon at the 5'-terminus and a translation stop codon at the 3'-terminus. A coding sequence can include, but is not limited to mRNA, cDNA, and recombinant polynucleotide sequences.

"Immunologically identifiable with/as" refers to the presence of epitope(s) and polypeptides(s) which are also present in the designated polypeptide(s), usually HCV proteins. Immunological identity may be determined by antibody binding and/or competition in binding; these techniques are known to those of average skill in the art, and are also illustrated *infra*.

As used herein, "epitope" refers to an antigenic determinant of a polypeptide; an epitope could comprise 3 or more amino acids in a spatial conformation which is unique to the epitope. Generally an epitope consists of at least 5 such amino acids, and more usually, consists of at least 8-10 such amino acids. Methods of determining the spatial conformation of amino acids are known in the art, and include, for example, X-ray crystallography and 2-dimensional nuclear magnetic resonance.

A polypeptide is "immunologically reactive" with an antibody when it binds to an antibody due to antibody recognition of a specific epitope contained within the polypeptide. Immunological reactivity may be determined by antibody binding, more particularly by the kinetics of antibody binding, and/or by competition in binding using as competitor(s) a known polypeptide(s) containing an epitope against which the antibody is directed. The techniques for determining whether a polypeptide is immunologically reactive with an antibody are known in the art.

As used herein, the term "immunogenic polypeptide" refers to a polypeptide that elicits a cellular and/or humoral response, whether alone or when linked to a carrier in the presence or absence of an adjuvant.

The term "polypeptide" refers to a polymer of amino acids and does not refer to a specific length of the product; thus, peptides, oligopeptides, and proteins are included within the definition of polypeptide. This term also does not refer to or exclude post-expression modifications of the polypeptide, for example,

glycosylations, acetylations, phosphorylations and the like.

An "individual", as used herein, refers to vertebrates, particularly members of the mammalian species, and includes but is not limited to domestic animals, sports animals, and primates, including humans.

As used herein, the "sense strand" or "plus strand" of a nucleic acid contains the sequence that has
 5 sequence homology to that of mRNA. The "antisense strand" or "minus strand" contains a sequence which is complementary to that of the "sense strand" or "plus strand".

As used herein, a "positive stranded genome" of a virus is one in which the genome, whether RNA or DNA, is single-stranded and which encodes a viral polypeptide(s). Examples of positive stranded RNA viruses include Togaviridae, Coronaviridae, Retroviridae, Picornaviridae, and Caliciviridae. Included also, are
 10 the Flaviviridae, which were formerly classified as Togaviridae. See Fields & Knipe (1986).

As used herein, a "replicative intermediate" of an HCV genome refers to an RNA strand or fragment thereof, which is complementary to the viral genome, and which is synthesized during the course of viral replication; the replicative intermediate functions as a template for the synthesis of positive RNA strands.

As used herein, "purified HCV" refers to a preparation of HCV which has been isolated from the cellular
 15 constituents with which the virus is normally associated, and from other types of viruses which may be present in the infected tissue. The techniques for isolating viruses are known to those of skill in the art, and include, for example, centrifugation and affinity chromatography; a method of preparing purified HCV is discussed *infra*.

As used herein, a "biological sample" refers to a sample of tissue or fluid isolated from an individual,
 20 including but not limited to, for example, plasma, serum, spinal fluid, lymph fluid, the external sections of the skin, respiratory, intestinal, and genitourinary tracts, tears, saliva, milk, blood cells, tumors, organs, and also samples of *in vitro* cell culture constituents (including but not limited to conditioned medium resulting from the growth of cells in cell culture medium, putatively virally infected cells, recombinant cells, and cell components).

The term "oligonucleotide", as used herein, refers to primers, probes, oligomer fragments to be
 25 detected, oligomer controls and unlabeled blocking oligomers, and is defined as a molecule comprised of three or more deoxyribonucleotides or ribonucleotides, typically contains more than four, typically a maximum of about 100 nucleotides. Its exact size will depend upon many factors, which in turn depend on the ultimate function or use of the oligonucleotide.

The term "primer", as used herein, refers to an oligonucleotide which is capable of acting as a point of
 30 initiation of polynucleotide synthesis along a complementary strand when placed under conditions in which synthesis of a primer extension product which is complementary to a polynucleotide is catalyzed. Such conditions include the presence of four different nucleotide triphosphates or nucleoside analogs and an agent for polymerization such as DNA polymerase or reverse transcriptase, in an appropriate buffer ("buffer" includes substituents which are cofactors, or which affect pH, ionic strength, etc.), and at a suitable
 35 temperature.

As used herein, the term "target region" refers to a region of the nucleic acid which is to be amplified and/or detected.

As used herein, the term "probe" refers to a polynucleotide or oligonucleotide which forms a hybrid
 40 structure with a sequence in a target region, due to complementarity of at least one sequence in the probe with a sequence in the target region. Preferably the probe, however, does not contain a sequence complementary to sequence(s) used to prime the polymerase chain reaction.

As used herein, the term "thermostable nucleotide polymerase" refers to an enzyme which is relatively
 45 stable to heat when compared to nucleotide polymerases from *E. coli* and which catalyzes the polymerization of nucleotides. Generally, the enzyme will initiate synthesis at the 3'-end of the target sequence utilizing the primer, and will proceed in the 5'-direction along the template until synthesis terminates. A representative thermostable enzyme isolated from *Thermus aquaticus* (Taq), and method for using it is described in Saiki et al. (1988).

As used herein, the term "viral RNA", which includes HCV RNA, refers to RNA from the viral genome,
 50 fragments thereof, transcripts thereof, and mutant sequences derived therefrom.

As used herein, "cell line" refers to a propagated culture of cells after the first subculture, thus there are various embodiments including but not limited to individual cells, harvested cells, and cultures containing cells so long as these are derived from cells of the cell line referred to. By "derived" is meant progeny or issue. It is further known in the art that spontaneous or induced changes can occur in karyotype during
 55 storage or transfer. Therefore, cells derived from the cell line referred to may not be precisely identical to the ancestral cells or cultures, and the cell line referred to includes such variants. The term "cell lines" includes immortalized cells. Preferably, cell lines exclude cells which are derived by the hybridization of three cell types, for example, a trisoma.

As used herein, an "*in vitro*" cell system refers to cells which have replicated *in vitro*, thus the term includes primary cultures and cell lines.

As used herein, the term "blood cell" refers to cells found either circulating in the blood, in tissue, or in bone marrow, and includes cells of the myelocytic series, the monocytic series, the megakaryocytic series, the erythrocytic series, the lymphocytic series, and the plasmocytic series. The morphology by which these cells can be distinguished is discussed in THE MORPHOLOGY OF HUMAN BLOOD CELLS (1985).

As used herein, the term "peripheral blood cell" refers to a cell found circulating in the blood, and refers generally to eosinophils, neutrophils (both band and segmented), basophils, monocytes, thrombocytes, erythrocytes, lymphocytes, and plasmocytes.

As used herein, the term "leukocyte" refers to a heterogeneous group of blood cell types which includes everything except erythrocytes and platelets.

As used herein, the term "lymphocyte" refers to a spherical cell, 6-12 μ m in diameter, involved in immune responses, with a large, round, and deeply staining nucleus and very little cytoplasm. Lymphocytes include B lymphocytes and T lymphocytes.

As used herein, "B lymphocytes" are vertebrate white blood cells which, when mature, are involved in the production of antibody. Mature virgin B lymphocytes (i.e., these that have not encountered antigen) are characterized by the presence of the following surface markers: monomeric IgM receptors for antigen, Fc receptors for IgG; and receptors for the activated form of C3b component of complement.

Precursor B lymphocytes lack immunoglobulin products but may express other characteristic molecules that can be identified by specific monoclonal antibodies. These cells divide and undergo rapid transition to become large lymphoblasts with cytoplasmic μ heavy chains but no light chains. The pre-B cell phenotype (cytoplasmic μ chain $^{+}$, light chain $^{-}$, surface Ig $^{-}$) can first be demonstrated in lymphoblasts actively engaged in DNA synthesis. Human pre-B cells express HLA-DR molecules on their surface and may also express receptors for the C3b complement fragment. However, they lack Fc receptors for IgG and have very few C3d/EBV receptors. Precursor B lymphocytes can also be recognized by their capacity to bind peanut agglutinin.

Mature B lymphocytes may be either activated or resting. Activated B cells express several types of molecules that are not detected on resting B cells. These marker molecules are known by those of skill in the art, and include, for example, interleukin-2 (IL-2) receptors, and transferrin receptors (the latter are expressed by all kinds of dividing cells). Activation of B cells provokes loss of membrane IgD, and both Fc $_{\gamma}$ and C3b receptors are diminished on external membranes, whereas those for C3d/Epstein Barr Virus (EBV) are not significantly affected.

After antigen or mitogen stimulation, B lymphocytes may process along either of 2 branches of a differentiation pathway. They can differentiate into plasma cells, or they can divide and then return to a resting state as memory B cells. Memory B cells have had a loss of IgD during activation; this molecule is not resynthesized by cells entering the memory cell pool.

"Plasma cells" are terminally differentiated B lymphocytes. These cells are of ovoid shape, with an eccentric spoke-wheel nucleus, and intensely basophilic cytoplasm. When stained with fluorochrome-labeled antibodies to immunoglobulin determinants, fixed plasma cells display intense fluorescence throughout their cytoplasm. Membrane-bound immunoglobulin and DR molecules are scant, and receptors for Fc $_{\gamma}$, C3b, C3d, or EBV are generally undetectable.

A "plasmablast" is both morphologically and functionally between the activated lymphocyte (also called B lymphoblast) and plasma cell stages. The cells are large, with a greater nuclear:cytoplasmic ratio than plasma cells. Membrane-bound immunoglobulins and Fc $_{\gamma}$ receptors are present.

As used herein, a "T lymphocyte" is a thymus-derived cell which may participate in a variety of cell-mediated immune responses. There are a variety of types of T lymphocytes, which are distinguishable by their function, and by their cell surface markers. Methods for distinguishing the classes of T lymphocytes, which include those of the T "helper" and T "suppressor" types are known in the art. For example, monoclonal antibodies can be used to detect cell surface antigens, including the CD (cluster of differentiation) antigens. The T cell types marked by the CD antigens, and monoclonal antibodies which detect the specific antigens are shown in Table 1.

Table 1

Cell Type Marked	CD Designation	Antibody Designation	Comments
Cortical thymocytes	CD1	Leu 6 OKT 6	Early T cell antigen also present on Langerhans cells, associated with beta ₂ -microglobulin not present on peripheral T cells.
E rosette-forming cells	CD2	Leu 5 OKT 11 T11	Pan-T cell antigen SRBC receptor on T cells.
Mature T cells	CD3	Leu 4	Also present on T cell ALL and cutaneous T cell lymphoma.
Helper/inducer T cells	CD4	Leu 3 OKT 4 T4	Can be further subdivided into helper and inducer subsets.
Pan-T and -B cell subpopulation	CD5	Leu 1 T1 T101 OKT1	B cell CLL. B cells following marrow transplant.
Mature T and B cell subpopulation	CD6	T12	Malignant T cells.
Pan-T cells, thymocytes	CD7	Leu 9 3A1	T cell leukemias.
Suppressor/cytotoxic T cells	CD8	Leu 2 OHT 8 T8	Can be further subdivided into cytotoxic and suppressor subsets.

As used herein, "large granular lymphocyte" (LGL), also called natural killer (NK) cells, are another type of lymphocyte. These cells comprise a discrete population of large lymphocytes that can be distinguished morphologic features, for example, by characteristic azurophilic granules in their cytoplasm. In addition, LGL can be identified by methods which utilize specific monoclonal antibodies. Several monoclonal antibodies are available that detect either Fc receptors (Leu 11) or specific differentiation antigens (Leu 7, HNK-1, NKH-1, NKH-2) present on these cells. Some LGL cells also express antigens from the CD2 T cell family. Methods for functional testing are also known in the art, and are based on the ability of these nonimmune cells to kill special target cells, for example, erythroleukemia cell line K562. Methods of measuring cytotoxicity are known in the art, and include, for example the ^{51}Cr release assay.

As used herein, "monocytes" denotes large agranulocytic leukocyte blood cells which are phagocytic. In humans, these cells are approximately 10-12 μm in diameter, with oval nuclei. Monocytes which are in tissue are known as macrophages. Monocytes/macrophages may be identified by their morphology. They are larger than granulocytes and most lymphocytes. They have round or kidney-shaped nuclei with fine, lightly stained granules. They may be detected by staining for their nonspecific esterase, or alpha-naphthol esterase. Monoclonal antibodies directed at specific differentiation antigens are also available, and include, for example, Leu M3, and OKT M1. The term "monocytes" includes monoblasts and promonocytes. The cells of the mononuclear phagocyte system, and their localization, are shown in Table 2: methods for distinguishing the cell types are discussed in BASIC & CLINICAL IMMUNOLOGY, 6th edition.

Table 2

Cells of the Mononuclear Phagocyte System *	
Cells	Localization
Stem cells (committed)	Bone marrow
Monoblasts	Bone marrow
Promonocytes	Bone marrow
Monocytes	Bone marrow
Macrophages	Tissues
Normal state, free	
Histocytes	Connective tissues
Alveolar macrophages	Lung
Pleural and peritoneal macrophages	Serous cavities
Normal state, fixed	
Kupffer cells	Liver
Osteoclasts	Bone
Microglial cells	Nervous System
Synovial type A cells	Joints
Fixed tissue macrophages	Spleen, lymph nodes, bonemarrow, and other tissues
Inflammation	
Exudate macrophages	Any tissue
Activated macrophages	Any tissue
Elicited macrophages	Any tissue
Epitheloid cells	Any tissue
Multinucleated giant cells (Langerhans types and foreign body type)	Any tissue

* Adapted from Van Furth R (editor): Mononuclear Phagocytes: Functional Aspects, Martinus Nijhoff, 1980.

As used herein, the term "granulocyte" refers to a polymorphonuclear leukocyte which contains many cytoplasmic granules and a multilobed nucleus. Granulocytes include neutrophils, eosinophils, and neutrophils. Neutrophils are amoeboid in shape, and phagocytic, and stain with both basic and acidic dyes.

5 Eosinophils and basophils contain cytoplasmic granules which stain with acid dyes and with basic dyes, respectively.

"Bone marrow cells" include both primary bone marrow tissue and bone marrow cell lines from mammals, such as primates and humans. Methods of culturing primary bone marrow cells *in vitro* are known, as well as bone marrow cell lines such as IM-9, KG-1 and KG-1a. See, e.g., ATCC CATALOG,
10 *supra*, at 361. Primary bone marrow cells may include, in addition to leukocytes and lymphocytes described elsewhere herein, progenitor cells such as pleuripotent stem cells. See, e.g., Civin U.S. Pat. No. 4,714,680.

15 II. Description of the Invention

The invention described herein allows the replication of HCV in cell lines. In a method of the invention, peripheral blood cells which are infected with replicating virions are detected. Preferably, the detection is accomplished by identifying replicative intermediates of the viral genome in preparations containing HCV
20 infected cells. Alternatively, the virions may be detected using immunoassays to detect a population of HCV infected cells. Cell lines infected with HCV are prepared; these cell lines are usually derived from blood cells or their precursors, more usually are derived from mononuclear cells or their precursors. The preparation of the infected cell line may be accomplished by a variety of methods, including the following. One option is to cultivate susceptible cells with concentrated culture fluids harvested from short term
25 primary cultures of cells which are replicating the virus. Another option is to directly infect susceptible cells with isolated HCV particles. Still another option is to grow susceptible cells in contact with HCV infected cells. The susceptible cells may be from an established cell line, or may be primary cultures. Alternatively, infected cells from an individual with HCV may be fused with established cells from a cell line to produce a permanent cell line. Still alternatively, HCV infected blood cells isolated from an individual may be
30 immortalized. Virions produced by the HCV infected cell lines are useful in immunoassays for the detection of anti-HCV antibodies, and as a substrate for the production of vaccines for treatment of HCV.

The practice of the present invention will employ, unless otherwise indicated, conventional techniques of molecular biology, microbiology, hematology, cell and organ culture, recombinant DNA, and immunology, which are within the skill of the art. Such techniques are explained fully in the literature. See, e.g., ATCC
35 CATALOG OF CELL LINES & HYBRIDOMAS, 6th edition, 1988 (American Type Culture Collection, Rockville, Md.); BASIC & CLINICAL IMMUNOLOGY, 6th edition (Stites, Stobo, and Wells, eds., Appleton and Lange, Norwalk Conn/Los Altos, Ca, 1987); Boggs and Winkelstein, WHITE CELL MANUAL, edition 4 (F.A. Davis Co., Phila., 1983); Diggs, Sturm, and Bell (1985), THE MORPHOLOGY OF HUMAN BLOOD CELLS (Abbott Laboratories, Abbott Park, Il.); Freshney (1987), CULTURE OF ANIMAL CELLS: A MANUAL
40 OF BASIC TECHNIQUE (Alan R. Liss, Inc., N.Y.); Maniatis, Fritsch & Sambrook, MOLECULAR CLONING; A LABORATORY MANUAL (1982); DNA CLONING, VOLUMES I AND II (D.N. Glover ed. 1985); LYMPHOKINES AND INTERFERONS: A PRACTICAL APPROACH (Clemens, Morris, and Gearing, eds., 1987); LYMPHOCYTES: A PRACTICAL APPROACH (Klaus, ed., 1987); VIROLOGY: A PRACTICAL APPROACH (Mahy, ed., 1985); OLIGONUCLEOTIDE SYNTHESIS (M.J. Gait ed, 1984); NUCLEIC ACID HYBRIDIZATION
45 (B.D. Hames & S.J. Higgins eds. 1984); IMMOBILIZED CELLS AND ENZYMES (IRL Press, 1986); B. Perbal, A PRACTICAL GUIDE TO MOLECULAR CLONING (1984); the series, METHODS IN ENZYMOLOGY (Academic Press, Inc.); GENE TRANSFER VECTORS FOR MAMMALIAN CELLS (J.H. Miller and M.P. Calos eds. 1987, Cold Spring Harbor Laboratory), Methods in Enzymology Vol. 154 and Vol. 155 (Wu and Grossman, and Wu, eds., respectively), Mayer and Walker, eds. (1987), IMMUNOCHEMICAL METHODS
50 IN CELL AND MOLECULAR BIOLOGY (Academic Press, London), Scopes, (1987), PROTEIN PURIFICATION: PRINCIPLES AND PRACTICE, Second Edition (Springer-Verlag, N.Y.), and HANDBOOK OF EXPERIMENTAL IMMUNOLOGY, VOLUMES I-IV (D.M. Weir and C. C. Blackwell eds 1986). All patents, patent applications, and publications mentioned herein, both *supra* and *infra*, are hereby incorporated herein by reference.

55

II.A. Detection and Isolation of HCV Infected Cells from an Individual

HCV infected cells in biological samples are screened for the presence of HCV polypeptides or HCV polynucleotides. Usually the biological samples are blood samples, from which the cells infected with HCV and/or replicating HCV are isolated and partially purified utilizing techniques known in the art, for example, by sedimentation through Ficoll-Paque (Pharmacia Corp.) solutions.

5 The screening technique used to detect viral material must be sufficiently sensitive to detect very limited amounts. For example, the presence of virions may be detected by immunoassay, using antibodies directed against a virion polypeptide(s). A suitable antibody is described in U.S.S.N. 355,002 (Attorney Docket No. 2300-0063.28), filed May 18, 1989. Methods for immunoassays, as well as methods for signal enhancement are known in the art.

10 The HCV genomic material, and replicative intermediates thereof, may also be detected by nucleic acid hybridization. Usually the system will involve amplification techniques in the hybridization assay. Such techniques are known in the art. For example, the Enzo Biochemical Corporation "Bio-Bridge" system uses terminal deoxynucleotide transferase to add unmodified 3'-poly-dT-tails to a DNA probe. The poly dT-tailed probe is hybridized to the target nucleotide sequence, and then to a biotin-modified poly-A. PCT application 15 84/03520 and EPA 124221 describe a DNA hybridization assay in which: (1) analyte is annealed to a single-stranded DNA probe that is complementary to an enzyme-labeled oligonucleotide; and (2) the resulting tailed duplex is hybridized to an enzyme-labeled oligonucleotide. EPA 204510 describes a DNA hybridization assay in which analyte DNA is contacted with a probe that has a tail, such as a poly-dT tail, an amplifier strand that has a sequence that hybridizes to the tail of the probe, such as a poly-A sequence, and 20 which is capable of binding a plurality of labeled strands. A particularly desirable technique may first involve amplification of the target HCV sequences in sera until the amplified sequences are at a detectable level. This may be accomplished, for example, by the polymerase chain reactions (PCR) technique described by Saiki et al. (1986), by Mullis, U.S. Patent No. 4,683,195, and by Mullis et al. U.S. Patent No. 4,683,202. The amplified sequence(s) may then be detected using a hybridization assay which is described in co-pending 25 U.S. Serial No. 109,282 (Attorney Docket No. 2300-0171), which was filed 15 October 1987, U.S. Serial No. 185,201 (filed 22 April 1988), and U.S. Serial No. 252,638 (filed 30 September 1988), which are assigned to the herein assignee, and are hereby incorporated herein by reference. These hybridization assays, which should detect sequences at the level of 10^6 /ml, utilize nucleic acid multimers which bind to single-stranded analyte nucleic acid, and which also bind to a multiplicity of single-stranded labeled oligonucleotides. A 30 suitable solution phase sandwich assay which may be used with labeled polynucleotide probes, and the methods for the preparation of probes is described in EPO 225,807, published June 16, 1987, which is assigned to the herein assignee, and which is hereby incorporated herein by reference.

A preferred method for detecting HCV genomic sequences is described in the Examples. This method is based on an HCV/cPCR method described in U.S.S.N. (attorney docket 2300-0063.27), filed May 18, 35 1989, which is commonly owned by the herein assignee, and which is hereby incorporated herein by reference. The method utilizes primers and probes based upon a family of cDNA replicas of portions of the HCV genome; a composite sequence of the cDNA plus strand derived from this family of cDNAs is shown in Figure 1. The first step in the method is the synthesis of a cDNA to either the HCV genome or its replicative intermediate, using reverse transcriptase. If replicative intermediates are to be detected, the 40 primer is complementary to only the negative strand of HCV RNA (i.e., the strand which is the replicative intermediate). After synthesis of the HCV cDNA, and prior to amplification, the RNA in the sample is degraded by techniques known in the art, for example, by alkali denaturation, or by treatment with an RNA specific RNase.

It is anticipated that variant strains of HCV will be present in the infected cells of various individuals. 45 Therefore the detection methods should be directed to constant epitopes or genomic sequences of HCV.

Utilizing the HCV/cPCR method to screen fractions of blood cells separated on Ficoll-Paque, replicative intermediates of the HCV genome were detected in a partially purified composition of cells derived from peripheral blood (See *infra* in the Examples). This composition is comprised of lymphocytes, monocytes/macrophages, platelets, and small amounts of other white blood cell types. The sedimentation 50 through Ficoll-Paque fractionates these cells, which band to the interface, from erythrocytes and granulocytes, which are of greater density.

Further identification of the infected cell types may be accomplished by utilizing techniques known in the art, for example by immunoassays which are directed to distinguish between the various types of blood cells. Monoclonal antibodies which are directed to surface epitopes on various lymphocytes, and 55 monocytes/macrophages are known in the art. Thus, HCV infected cells may be isolated and identified by immunologic binding to specific cell surface antigens.

II.B. Preparation of *In vitro* Infected Cell Systems

A variety of methods may be used to prepare *in vitro* cells, including cell lines, infected with HCV. Candidate cells for infection and propagation *in vitro* can be from any of the above-mentioned cell types, such as lymphocytes or bone marrow cells. The discovery that replicative intermediates of the HCV genome are present in leukocytes in the peripheral blood of infected humans is indicative of the cell type which may support viral replication *in vitro*, i.e., in at least one of the cell types in the population of leukocytes which band at the interface of a leukocyte suspension sedimented through Ficoll-Paque, i.e., mononucleocytes. (This procedure generally removes normal erythrocytes and granulocytes from peripheral blood samples). Further fractionation procedures may be used to separate the different types of mononucleocytes to determine the presence of HCV replicative intermediates and/or HCV antigens associated with these cell types. Procedures for separating different types of monocytes, e.g., various types of lymphocytes, and macrophages, are known in the art. See, e.g., LYMPHOCYTES: A PRACTICAL APPROACH (Klaus, ed.). These procedures may use monoclonal antibodies directed towards specific cell types; the monoclonal antibodies may be affixed to solid substrates, e.g., plates with wells, or magnetic beads. Alternatively, sorting techniques, which rely upon cell sorter instruments may be used. The selection may be either positive or negative to enrich and/or select the desired cells in the population of mononucleocytes. Based upon fractionation studies which isolated lymphocytes and macrophages from the monocyte population, it is suggested that a preferred cell type for replication may be lymphocytes and/or macrophages/monocytes.

Since replication of HCV occurs within mononucleocytes *in vivo*, it is expected that infected cell lines may be derived from known established cell lines of these types, many of which are available from the American Type Culture Collection, and which may include, for example, B cell lines (e.g., EBV transformed B cell derived lines such as B95-8, DAKIKI, LTR228, Mo-B, SC-1, WIL2-NS; B cell lines such as Ia2, BL-3, UC 729-6, Namalwa, U-937, Ramos (RA-1), BCL₁ clone 5B₁b, MC/CAR, Ia2, YB2/O, CA3-F4, SHM-D33, CW13.20-3B3, BC9-E%, WM-115, WM 266-4, WEHI-231, WEHI-279, NFS-1.0 C-1); monocyte lines (e.g., THP-1, WEHI-274.1), mouse lymphoma lines (e.g., 2pk-3), pro-B lymphoblasts (e.g., NFS-70 C-10), pre-B lymphoblasts (e.g., NFS-25 C-3), T-cell derived lines (e.g., CTLL-2, HuT 102, D10.G4.1, LBRM-33 clone 4A2, HuT78, Cl.Ly1⁺ 2⁻/9, UCD-MLA-144, 6T-CEM 20, 6T-CEM, CCRF-CEM, H9/HTLV-III_B, CCRF-SB; CCRF-HSB-2, CEM-CM3, Mo, MOLT-3, MOLT-4, BW5147.3, BW5147.G.1.4, C1498, RAW8.1, CTLL-2, and BW5147.G.1.4.OUA^R.1). Additional human leukemic cell lines of interest include K562 and ARH-77. Conditions for the *in vitro* culture of these cells is described in ATCC CATALOG OF CELL LINES AND HYBRIDOMAS, 6th Edition, 1988.

Another type of cell system in which HCV may replicate *in vitro* is in primary cultures of cell types of cells which are infected with HCV *in vivo*. Primary cultures, as used herein, refers to culture of cells that are directly derived from cells or tissues from an individual, as well as cells derived by passage from these cells, but not to immortalized cells. Methods for preparing primary cultures, particularly of mononucleocytes, and conditions for their growth *in vitro* are known in the art (See, for example, LYMPHOCYTES: A PRACTICAL APPROACH (Klaus, ed.), particularly Hunt (1987); and Freshney (1987). Generally, normal mature macrophages do not proliferate; however, replicating precursor cells of macrophages may be cultured by techniques known in the art. See, for example, Freshney (1987), pp. 284-287.

Still another type of cell system in which HCV may be replicated is in immortal cell lines derived from primary cultures of mononucleocytes. Methods to obtain immortalized cell lines of types of mononucleocytes are known in the art. For example, methods for growing and immortalizing B cells, T cells, and macrophages in culture are described in LYMPHOCYTES: A PRACTICAL APPROACH (Klaus, ed.); LYMPHOKINES AND INTERFERONS (Clemens, Morris, and Gearing, eds), and Freshney (1987). For example, B-cell lines are often immortalized by infection with Epstein-Barr virus (EBV). This may be accomplished by incubating mononucleocytes from infected cells with the supernatant from an EBV infected cell line which is a chronic shedder of EBV, for example, ATCC CRL 1612 cells (B95-8).

The growth of mononucleocytes in culture often requires the presence of stimulatory protein factors (also called cytokines) in the culture medium; these factors are usually produced by a cell of another type. A supply of the cytokines may be provided in conditioned medium, i.e., cell culture supernatant from cells which produce the desired factors; or it may also be provided by feeder cells. The requirement may also be met by the provision of the purified cytokines. Cytokines include, for example, Interleukin 1-alpha and beta (also called lymphocyte activating factor, epidermal cell derived thymocyte activating factor, and haematopoietin H1); these lymphokines are derived from multiple cell types, including monocytes, lymphocytes, and keratinocytes. Interleukin 1-alpha and beta have multiple actions, including stimulating interleukin 2 production and haematopoietic activity. Interleukin 2 (also called T-cell growth factor), derived

from T-lymphocytes, and which stimulates T and B cell proliferation and differentiation, macrophage activation, and natural killer cell activation. Interleukin 3 (also called Multi-CSF), which is derived from T-lymphocytes, and which has a pluripotent effect on growth and differentiation, and which stimulates mast cell growth. Interleukin 4 (also called B-cell growth factor I), which is derived from T-lymphocytes, and which stimulates T and B cell proliferation and differentiation. Interleukin 5 (also called B-cell growth factor II), which is derived from T-lymphocytes, and which stimulates eosinophil differentiation (and which, in the mouse system, acts as a B-cell growth factor). Low molecular weight B-cell growth factor, which is derived from T-lymphocytes, and which stimulates B-cell proliferation. B-cell stimulatory factor 2 (also called interferon beta-2), which is derived from T-lymphocytes, and which stimulates B cell differentiation. Interferon- γ (also called immune interferon), which is produced by T-lymphocytes, and which activates macrophages. Granulocyte colony stimulating factor, which is derived from a number of cell types, and which stimulates the growth and differentiation of granulocytes. Macrophage colony stimulating factor (CSF 1), which is derived from a number of cell types, and which stimulates the growth and differentiation of monocytes/macrophages. Granulocyte macrophage colony stimulating factor, which is derived from a number of cell types, and which stimulates the growth and differentiation of monocytes/granulocytes.

Infection of a susceptible cell line with HCV may be accomplished by any one of several techniques which are used in the art to infect cells with virus, particularly with an RNA virus, and more particularly with a Flavivirus. Generally, virus particles adsorb to cultured cells at physiological temperatures. The virus penetrates, uncoats, replicates its nucleic acid in the nucleus or cytoplasm, and after transcription and translation assembles progeny virions which egress either by cell lysis or budding. The number of progeny virus particles released from one cell is referred to as the burst size. Virus is titrated by adsorbing serial tenfold dilutions to cells and counting the resultant plaques after appropriate incubation at the permissive temperature.

Viral infection of susceptible cells may be accomplished by cocultivation of the susceptible cells and HCV infected cells. The HCV infected cells may be from a primary culture. Usually the HCV infected cells and the primary cells are incubated together in approximately equal numbers in an appropriate cell culture medium. The medium may contain Polybrene, or a functionally similar substance, in an appropriate concentration. The incubation is of sufficient time and at the appropriate temperature to allow the newly replicated virus from the HCV infected cells to allow attachment and uptake by the susceptible cells.

The infection of susceptible cells may also be accomplished using virus containing cell-free supernatants of HCV infected cells. Generally the susceptible cells are pretreated with an appropriate concentration of Polybrene (or a functionally similar substance), and then exposed to concentrated culture fluids harvested from short-term primary cultures of HCV infected cells derived from infected individuals.

Susceptible cells may also be infected with virus isolated from HCV containing biological samples. Suitable methods for the isolation of viruses, particularly RNA containing viruses, and more particularly Togaviruses and Flaviviruses are known in the art, for example, see *VIROLOGY: A PRACTICAL APPROACH* (Mahy, ed.). Also described therein are cells which may be suitable for infection, and growth conditions for these cells.

Another method for preparing HCV infected cells *in vitro* may be by the fusion of HCV infected mononuclear cells with susceptible cells. Methods for fusing somatic cells are known in the art, as are methods for the selection of fused cells. For example, fusion may be catalyzed by the presence of virus, e.g., EBV, Sendai virus, and the like, or by the presence of chemicals, e.g., polyethylene glycol, which induce fusion. The susceptible cell may carry a marker which prevents its growth in the selection media, e.g., the cells may be phenotypically negative for thymidine kinase. Selection is then in media which requires thymidine kinase, e.g., HAT medium. This method is particularly suitable if the HCV infected mononuclear cells isolated from the infected individual do not replicate *in vitro*. A suitable method for the preparation fused cells which may replicate HCV is that described in WO 87/05930, except that the tissue specific cell (reportedly a hepatocyte) which is used to form the trioma is replaced with an HCV infected mononucleocyte.

Yet another method of preparing an HCV infected cell line is by immortalization of infected cells isolated from an HCV infected individual. Generally, mononuclear cells are isolated from peripheral blood. If desired, the mononuclear cell population may be further fractionated. The cells are then grown in primary culture. Immortalization may be accomplished by growing and passaging the cells through the cell crisis period, at which point the majority of cultured cells die; the resulting cells are immortal. Alternative methods for producing immortal T-cells, B-cells, and macrophages are described *supra*.

HCV replication in the HCV infected cell lines prepared as described *supra* may be detected by a variety of assays. For example, viral assays may depend upon the cytopathic or transforming characteristics of the virus. Cytopathic viruses may be assayed by their antimetabolic effects in microtitration plates or by

the formation of characteristic plaques in monolayers of the appropriate susceptible host cell. A viral suspension is serially diluted and added to monolayer culture plates. The number of plaques forming at the limiting dilution is taken as equivalent to the number of infectious particles in the supernatant medium, allowing the concentration of virus in the initial sample to be calculated. Characterization of the virus may be performed with specific antisera, measuring the cytopathic effect of the virus, by immunoassay, or by the cPCR/HCV assay for HCV RNA, described *supra*.

Transforming viruses may be assayed by, for example, the selective growth of transformed clones in suspension, or by looking for transformation foci in monolayer cultures.

In the event that the virus of interest exhibits neither a detectable cytopathic nor a transforming effect, it may be detected in the infected cell preparation or infected cell lysate by a number of techniques, including, for example, immunoassay using a labeled antibody(s) which is specific for the virus, immunoassay of polypeptides labeled during replication using anti-HCV antibodies, and the detection of viral nucleic acids, for example, an enriched population of negative strands complementary to viral positive strands, or of viral nucleic acids labeled during an *in vitro* viral synthesis period.

II.C. Immunoassays Using Antigens from HCV Infected Cells

The antigens from HCV infected cells which react immunologically with serum containing HCV antibodies are useful in immunoassays to detect presence of HCV antibodies in biological samples. The HCV antigens can take the form of, *inter alia*, purified or partially purified virus particles or viral polypeptides, disrupted virus, or infected cells or cell lysate therefrom. The production of the various forms of viral antigen from tissue culture sources is well-known in the art. Design of the immunoassays is subject to a great deal of variation, and a variety of these are known in the art. For example, an immunoassay may use a combination of viral epitopes derived from the same or from different viral polypeptides. It may use, for example, one or more viral antigens isolated from the HCV infected cell or from isolated HCV. Alternatively, fixed cells directly from culture may be used if viral epitopes are expressed internally or externally, or alternatively, cellular lysates may be used. It may also use, for example, whole disrupted virions. General methods for preparing disrupted virions are known in the art.

Immunoassay protocols may vary. Protocols may be based, for example, upon competition, or direct reaction, or sandwich type assays. Protocols may also, for example, use solid supports, or may be by immunoprecipitation. Most assays involve the use of labeled antibody or polypeptide; the labels may be, for example, fluorescent, chemiluminescent, radioactive, or dye molecules. Assays which amplify the signals from the probe are also known; examples of which are assays which utilize biotin and avidin, and enzyme-labeled and mediated immunoassays, such as ELISA assays.

A suitable immunoassay may utilize disrupted virions in an ELISA format. The enzyme-linked immunosorbent assay (ELISA) method depends upon conjugation of an enzyme to either an antigen or an antibody, and uses the bound enzyme activity as a quantitative label. To measure antibody, the known antigen is fixed to a solid phase, (e.g., a microplate or plastic cup), incubated with anti-immunoglobulin labeled with enzyme, and washed again. Enzymes suitable for labeling are known in the art, and include, for example, horseradish peroxidase. Enzyme activity bound to the solid phase is measured by adding the specific substrate, and determining product formation or substrate utilization colorimetrically. The enzyme activity bound is a direct function of the amount of antibody bound.

To measure antigen, a known specific antibody is fixed to the solid phase, the test material containing antigen is added, after an incubation the solid phase is washed, and a second enzyme-labeled antibody is added. After washing, substrate is added, and enzyme activity is estimated colorimetrically, and related to antigen concentration.

Thus, for example, to detect anti-HCV antibodies using an ELISA format, isolated HCV, solubilized, e.g., with detergent, may be used to coat appropriate microtiter plates. After the viral antigens are bound, another protein is also used to coat the plates, to prevent nonspecific binding of antibodies. The biological sample to be tested for anti-HCV antibodies is added to the plates, and the formation of HCV antibody-HCV antigen complexes is detected.

Another suitable immunoassay may utilize gel electrophoresis of HCV lysates, followed by Western blotting with the biological sample to be tested for anti-HCV antibodies. The formation of HCV antibody-HCV antigen complexes is detected using techniques known in the art.

Kits suitable for immunodiagnosis and containing the appropriate labeled reagents are constructed by packaging the appropriate materials, including the antigens derived from the HCV infected cells in suitable containers, along with the remaining reagents and materials required for the conduct of the assay, as well

as a suitable set of assay instructions.

II.D. Use of Infectable Cells in Screening for Antiviral Agents

5 The cells described herein which are capable of replicating HCV, may be used to screen for antiviral agents which inhibit HCV replication, and particularly for those agents which preferentially allow cell growth and multiplication while inhibiting viral replication. The screening methods which can be used are those which examine the inhibition of virus replication in the presence of the putative antiviral agent. Generally, 10 the antiviral agents are tested at a variety of concentrations, for their effect on preventing viral replication in a cell culture system which supports viral replication, and then for an inhibition of infectivity or of viral pathogenicity (and a low level of toxicity) in an animal model system.

Included within antiviral agents are antibodies which block viral infection (neutralizing antibodies). The ability of anti-HCV antibodies to prevent the infection of the cells *in vitro* can be determined using the 15 assays described herein to determine whether viral replication is inhibited by the presence of the putative anti-HCV antibodies. For example also, virus produced in infected cells may also be used as a source of antigens to determine whether antibodies are HCV specific.

20 III. Examples

III.A. Detection of HCV RNA in Mononuclear Cells From Individuals with NANBH

25 The detection of HCV RNA in mononuclear cells from blood of individuals with chronic NANBH was accomplished using essentially the cPCR/HCV assay described in U.S.S.N. 07/355,981.

Blood obtained from 5 individuals with chronic NANBH and which tested positive for anti-HCV antibodies using an HCV-C100 antigen immunoassay, and from 5 control individuals (reportedly free of NANBH, and which was negative in the immunoassay) was examined for the presence of HCV RNA in the 30 mononuclear fraction. A mononucleocyte fraction from the samples was obtained by sedimentation through Ficoll-Paque (Pharmacia), following the manufacturer's directions.

The cPCR/HCV assay used in these studies were performed utilizing the following methods for the preparation of RNA, the reverse transcription of the RNA into cDNA, the amplification of specific segments of the cDNA by PCR, and the analysis of the PCR products.

35 Total RNA was isolated from the mononucleocyte fraction. The cDNA used as a template for the PCR reaction was prepared utilizing the designated samples for preparation of the corresponding cDNAs. Each RNA sample (corresponding to cells in 10 microliters of blood) was incubated in a 25 microliter reaction containing 1 micromolar of each primer, 1 millimolar of each deoxyribonucleotide triphosphate (dNTP), 50 millimolar Tris-HCL, pH 8.3, 5 millimolar MgCl₂, 5 millimolar dithiothreitol (DTT), 73 millimolar KCl, 40 units 40 of RNase inhibitor (RNASIN), and 5 units of AMV reverse transcriptase. The incubation was for 60 minutes at 37° C. Following cDNA synthesis, the reactions were diluted with 50 microliters of deionized water (DIW), boiled for 10 minutes, and cooled on ice.

Amplification of a segment of the HCV cDNA was performed utilizing two synthetic 16-mer oligonucleotide primers whose sequences were derived from HCV cDNA clones 36 (anti-sense) and 37b 45 (sense). The sequence of the primer from clone 36 was:

5' GCA TGT CAT GAT GTA T 3'.

The sequence of the primer from clone 37b was:

5' ACA ATA CGT GTG TCA C 3'.

The primers were used at a final concentration of 1 micromolar each. In order to amplify the segment of 50 HCV cDNA which is flanked by the primers, the cDNA samples were incubated with 0.1 microgram of RNase A and the PCR reactants of the Perkin Elmer Cetus PCR kit (N801-0043 or N801-0055) according to the manufacturer's instructions. The PCR reaction was performed for either 30 cycles or 60 cycles in a Perkin Elmer Cetus DNA thermal cycler. Each cycle consisted of a 1 minute denaturation step at 94° C, an annealing step of 2 minutes at 37° C, and an extension step of 3 minutes at 72° C. However, the extension 55 step in the final cycle (30 or 60) was 7 minutes rather than 3 minutes. After amplification the samples were extracted with an equal volume of phenol: chloroform (1:1), followed by extraction with an equal volume of chloroform, and then the samples were precipitated with ethanol containing 0.2 M sodium acetate.

The cPCR products were analyzed as follows: The products were subjected to electrophoresis on 1.8%

alkaline agarose gels according to Murakawa et al. (1988), and transferred onto Zeta Probe paper (BioRad Corp.) by blotting gels overnight in 0.4 M NaOH. The blots were neutralized in 2 x SSC (1 x SSC contains 0.15 M NaCl, 0.015 M sodium citrate), prehybridized in 0.3 M NaCl, 15 mM sodium phosphate buffer, pH 6.8, 15 mM EDTA, 1.0% SDS, 0.5% nonfat milk (Carnation Co.), and 0.5 mg/ml sonicated denatured salmon sperm DNA. The blots to be analyzed for HCV cDNA fragments were hybridized to a ³²P-labeled probe generated by nick translation of the HCV cDNA insert sequence in clone 35, shown in Figure 2. After hybridization, the blots were washed in 0.1 x SSC (1 x SSC contains 0.15M NaCl, 0.01M Na citrate) at 65 °C, dried, and autoradiographed. The expected product size is 586 nucleotides in length; products which hybridized with the probe and migrated in the gels in this size range were scored as positive for viral RNA.

As a control, cPCR primers designed to amplify α 1-antitrypsin mRNA was performed to verify the presence of RNA in each sample analyzed. The coding region of the α 1-antitrypsin gene is described in Rosenberg et al. (1984). Synthetic 16-mer oligonucleotide primers designed to amplify a 365 nucleotide fragment of the coding region of the α 1-antitrypsin gene were derived from nucleotides 22-37 (sense) and nucleotides 372-387 (anti-sense). The PCR products were detected using a ³²P nick-translated probe which lies between, and not including, the cDNA/PCR primer sequences.

Due to the extreme sensitivity of the PCR reaction, all samples were run a minimum of three times. All false positive signals were eliminated when the following precautions were taken: 1) eliminating aerosols by using screw capped tubes with rubber O-ring seals; 2) pipetting with Rainin Microman positive displacement pipettors with disposable pistons/capillaries; and 3) selecting the oligonucleotide sequences for the cDNA and PCR primers from two non-contiguous cDNA clones.

The results showed that mononucleocytes from 5 of 5 HCV anti-C100 antibody positive samples were positive for HCV RNA in the cPCR HCV assay. All of the controls were negative in the same assay.

III.B. Detection of HCV RNA Minus Strands in Mononucleocytes from HCV Infected Individuals

The detection of minus strands in peripheral blood mononucleocytes was accomplished using the cPCR/HCV assay essentially as described in Section III.A., except that the cDNA was synthesized using either the 16mer primer from clone 36 (i.e., primer 36/16A), described in Section III.A. to prime the plus strand of HCV RNA, or the complement of the 16-mer (i.e., primer 36/16B), to prime the negative strand. In order to carry out the subsequent PCR reaction, the cDNA reaction mixture was boiled and treated with RNase prior to the addition of the second primer.

The results of the cPCR/HCV assay showed that 5 of the HCV anti-C100 antibody samples were positive for HCV negative strands; of these five samples, two showed high levels of HCV RNA negative strands. All 5 of these samples were about equally positive for the plus strands of HCV RNA. Of the five control samples, all were negative for both plus and minus strands of HCV RNA.

As an additional control, plasma samples from three of the HCV positive individuals were also analyzed. The signal obtained from the positive strands were at least 5 to 10 times stronger than was the signal for the negative strands. Thus, the plasma samples were strongly positive for the plus strands, and only weakly positive to negative for the minus strands.

In HCV, which is Flavi-like, the viral genome is positive stranded RNA. The discovery of the great enrichment of negative strands of HCV RNA in monocytes relative to that in plasma, suggests that replicative intermediates of the viral genome are present in these cells.

IV. Commercial Utility

HCV infected cell systems, which are infected and/or replicated *in vitro*, including HCV infected cell lines, serve as a source of viral antigens which are useful in the detection of HCV antibodies, and hence in the diagnosis and the screening of biological materials (including blood) for HCV. These antigens also may have use in the preparation of vaccines for the treatment (prophylactic and/or therapeutic) of viral infections.

In addition, these cell systems are useful in the screening of anti-viral agents directed at preventing HCV replication, including antibodies which can neutralize infection or mediate antibody-dependent cellular cytotoxicity (ADCC).

While the invention has been described above by way of illustration, the present invention is not limited thereto and its scope is to be defined by the appended claims and the equivalents thereof. All of the references cited above are incorporated by reference herein.

Claims

1. A cell line infected with HCV.
2. The cell line of claim 1, wherein the cells in the cell line are mononuclear cells.
3. The cell line of claim 2, wherein the mononuclear cells are lymphocytes.
4. The cell line of claim 3, wherein the lymphocytes are B lymphocytes.
5. The cell line of claim 3, wherein the lymphocytes are T lymphocytes.
6. The cell line of claim 2, wherein the mononuclear cells are macrophage precursor cells.
7. The cell line of claim 1, wherein the cells are mononuclear cells comprised of an antigen selected from the group of antigens CD1, CD2, CD3, CD4, CD5, CD6, CD7, and CD8.
8. The cell line of claim 1, wherein the cells are mononuclear cells selected from the group consisting of stem cells derived from the bone marrow, monoblasts, promonocytes, monocytes, and macrophages.
9. The cell line of claim 8, wherein the macrophages are selected from the group consisting of histocytes, alveolar macrophages, pleural macrophages, peritoneal macrophages, Kupffer cells, osteoclasts, synovial type A cells, fixed tissue macrophages, exudate macrophages, activated macrophages, elicited macrophages, epitheloid cells, and precursor cells from which these cells are derived.
10. The cell line of claim 1, wherein the cells are derived by infection of a cell selected from the group of cell lines B95-8, DAKIKI, LTR228, Mo-B, SC-1, WIL2-NSIa2, BL-3, UC 729-8, Namalwa, U-937, Ramos (RA-1), BCL₁ clone 5B₁b, MC/CAR, IA2, YB2/0, CA3-F4, SHM-D33, CW13.20-3B3, BC9-E%, WM-115, WM 266-4, WEHI-231, WEHI-279, NFS-1.0 C-1, THP-1, WEHI-274, 12pk-3, NFS-70 C-10, NFS-25 C-3, CTLL-2, HuT 102, D10.G4.1, LBRM-33 clone 4A2, HuT78, Cl.Ly1⁺2⁻/9, UCD-MLA-144, 6T-CEM 20, 6T-CEM, CCRF-CEM, H9/HTLV-III_B, CCRF-CEM; CCRF-HSB-2, CEM-CM3, Mo, MOLT-3, MOLT-4, BW5147.3, BW5147.G.1.4, C1498, RAW8.1, CTLL-2, and BW5147.G.1.4.OUA^R.1.
11. A composition for the detection of an anti-HCV antibody comprised of an HCV antigen from an *in vitro* cell infected with HCV, wherein the antigen is affixed to a solid substrate, and wherein the antigen is immunologically reactive with said anti-HCV antibody.
12. A composition according to claim 11, wherein the composition is comprised of a lysate of cells from the cell line of claim 1.
13. A composition according to claim 11, wherein the composition is comprised of a lysate of HCV from the cell line of claim 2.
14. A composition according to claim 11, wherein the composition is comprised of a lysate of HCV from cells selected from the cells of claim 7, claim 8, claim 9, and claim 10.
15. A composition according to claim 11, wherein the composition is comprised of a cell of claim 1 affixed to a solid substrate.
16. A composition according to claim 11, wherein the composition is comprised of a cell of claim 2 affixed to a solid substrate.
17. A method for screening for an agent which inhibits HCV replication comprising:
 - (a) providing a composition comprised of a putative anti-viral agent;
 - (b) providing a cell line capable of replicating HCV;
 - (c) providing a composition comprised of infective HCV;
 - (d) incubating the compositions of (a) and (c) with the cell line of (b) under conditions which would, in the absence of (a) allow infection of and replication of HCV in the cell line; and
 - (e) detecting an inhibition of viral replication after the incubating in (d).

SEQUENCE OF THE HCV cDNA SENSE STRAND

 (AS DEDUCED FROM OVERLAPPING CLONES b114a/18g/
 ag30a/CA205a/CA290a/CA216a/pi14a/CA167b/CA156e/
 CA84a/CA59a/K9-1/26j/13i/12f/14i/11b/7f/7e/8h/
 33c/40b/37b/35/36/81/32/33b/25c/14c/8f/33f/33g/
 39c/35f/19g/26g/15e/b5a/16jh)

CACTCCACCATGAATCACTCCCCTGTGAGGAACTACTGTCTTCACGCAGAA
 AGCGTCTAGCCATGGCGTTAGTATGAGTGTCTGTCAGCCTCCAGGACCCCC
 CCTCCCGGGAGAGCCATAGTGGTCTGCGGAACCGGTGAGTACACCGGAATT
 GCCAGGACGACCGGGTCCTTTCTTGATCAACCCGCTCAATGCCTGGAGAT
 TTGGGCGTGCCCCGCAAGACTGCTAGCCGAGTAGTGTGGGTGCGGAAAG
 GCCTTGTGGTACTGCCTGATAGGTGCTTGCAGAGTGCCCCGGGAG-300

(Putative initiator methionine codon)

GTCTCGTAGACCGTGCACCATGAGCAGCAATCCTAAACCTCAAAAAAAAAA
 CAAACGTAACACCAACCGTTCGCCCACAGGACGTCAAGTTCCCGGGTGGCGG
 TCAGATCGTTGGTGGAGTTTACTTGTTCGCGCAGGGGCCCTAGATTGGG
 TGTGCGCGCGACGAGAAAGACTTCCGAGCGGTGCAACCTCGAGGTAGACC
 TCAGCCTATCCCCAAGGCTCGTTCGCGCCGAGGGCAGGACCTGGGCTCAGCC
 CGGGTACCCCTTGGCCCCCTCTATGGCAATGAGGGCTGCGGGTGGGC-600
 GGGATGGCTCCTGTCTCCCGTGGCTCTCGGCCTAGCTGGGGCCCCACAGA
 CCCCCGGCGTAGGTGCGCAATTTGGGTAAAGGTCATCGATAACCTTACGTG
 CGGCTTCGCGACCTCATGGGGTACATAACGCTCGTTCGGCGCCCCCTTGG
 AGGCGCTGCCAGGGCCCTGGCGCATGGCGTCCGGGTCTGGAAGACGGCGT
 GAACTATGCAACAGGGAACTTCCTGGTTGCTCTTTCTCTATCTTCTTCT
 GGCCCTGCTCTCTTGCTTGACTGTGCCCGCTTCGGCCTACCAAGT-900
 GCGCAACTCCACGGGGCTTTACCACGTCAACCAATGATTGCCCTAACTCGAG
 TATTGTGTACGAGGCGGCCGATGCCATCCTGCACACTCCGGGGTGGCGTCCC
 TTGCGTTTCGTGAGGGCAACGCCTCGAGGTGTTGGGTGGCGATGACCCCTAC
 GGTGGCCACCAGGGATGGCAAACTCCCCGCGACGCAGCTTCGACGTACAT
 CGATCTGCTTGTTCGGGAGCGCCACCCCTCTGTTTCGGCCCTCTACGTGGGGGA
 CCTATGCGGGTCTGTCTTTCTTGTTCGGCCAACTGTTACCTTCTC-1200
 TCCCAGGCGCCACTGGACGACGCAAGGTTGCAATTGCTCTATCTATCCCGG
 CCATATAACGGGTACCGCATGGCATGGGATATGATGATGAACTGGTCCCC
 TACGACGGCGTTGGTAATGGCTCAGCTGCTCCGGATCCACAAAGCCATCTT
 GGACATGATCGCTGGTGTCTACTGGGGAGTCTTGGCGGGCATAGCGTATTT
 CTCCATGGTGGGGAAGTGGGCGAAGGTCCTGGTAGTGCTGCTGCTATTTGC
 CGGCGTCGACGCGGAAACCCACGTCAACGGGGGAAGTGCCGGCCA-1500
 CACTGTGTCTGGATTTGTTAGCCTCCTCGCACCAGGCGCCAAGCAGAACGT
 CCAGCTGATCAACACCAACGGCAGTTGGCACCTCAATAGCACGGCCCTGAA
 CTGCAATGATAGCCTCAACACCGGCTGGTTGGCAGGGCTTTTCTATACCA
 CAAGTTCAACTCTTCAGGCTGTCTGAGAGGCTAGCCAGCTGCCGACCCCT
 TACCGATTTTGACCAGGGCTGGGGCCCTATCAGTTATGCCAACGGAAGCGG
 CCCCCAGCAGCGCCCTACTGCTGGCACTACCCCCCAAACCTTG-1800

FIG. 1-1

CGGTATTGTGCCCCGCGAAGAGTGTGTGTGGTCCGGTATATTGCTTCACTCC
 CAGCCCCGTGGTGGTGGGAACGACCGACAGGTCGGGCGCGCCACCTACAG
 CTGGGGTGAAAATGATACGGACGTCTTCGTCCTTAACAATACCAGGCCACC
 GCTGGGCAATTGGTTTCGGTTGTACCTGGATGAACTCAACTGGATTACACAA
 AGTGTGCGGAGCGCCTCCTTGTGTATCGGAGGGGCGGGCAACAACACCCT
 GCACTGCCCCACTGATTGCTTCCGCAAGCATCCGGACGCCACATA-2100
 CTCTCGGTGCGGCTCCGGTCCCTGGATCACACCCAGGTGCCTGGTCTGACTA
 CCCGTATAGGCTTTGGCATTATCCTTGTACCATCAACTACACCATATTTAA
 AATCAGGATGTACGTGGGAGGGGTGGAACACAGGCTGGAAGCTGCCTGCAA
 CTGGACGCGGGGCGAACGTTGCGATCTGGAAGACAGGGACAGGTCCGAGCT
 CAGCCCGTTACTGCTGACCACTACACAGTGGCAGGTCTCCCGTGTTCCTT
 CACAACCCCTACCAGCCTTGTCCACCGGCCTCATCCACCTCCACCA-2400
 GAAATTGTGGACGTGCAGTACTTGTACGGGGTGGGGTCAAGCATCGCGTC
 CTGGGCCATTAAAGTGGGAGTACGTGCTTCTCCTGTTCTTCTGCTTGCAGA
 CGCGCGCGTCTGCTCCTGCTTGTGGATGATGCTACTCATATCCCAAGCGGA
 GGCGGCTTTGGAGAACCCTCGTAATACTTAATGCAGCATCCCTGGCCGGGAC
 GCACGGTCTTGTATCCTTCTCGTGTTCTTCTGCTTTGCATGGTATTTGAA
 GGGTAAGTGGGTGCCCCGGAGCGGTCTACACCTTCTACGGGATGTG-2700
 GCCTCTCCTCCTGCTCCTGTTGGCGTTGCCCCAGCGGGCGTACGCGCTGGA
 CACGGAGGTGGCCCGTCTGCTGGCGGTGTTGTTCTCGTCGGGTGATGGC
 GCTGACTCTGTACCATATTACAAGCGCTATATCAGCTGGTGTGTTGGTG
 GCTTCAGTATTTTCTGACCAGAGTGGGAAGCGCAACTGCACGTGTGGATTCC
 CCCCCCTCAACGTCCGAGGGGGGCGCGACGCCGTCTTACTCATGTGTGC
 TGTACACCCGACTCTGGTATTTGACATACCAAATTGCTGCTGGC-3000
 CGTCTTCGGACCCCTTGGATTCTTCAAGCCAGTTTGCTTAAAGTACCCTA
 CTTTGTGCGCGTCCAAGGCCTTCTCCGGTTCTGCGCGTTAGCGCGGAAGAT
 GATCGGAGGCCATTACGTGCAAATGGTTCATCATTAAGTTAGGGGCGCTTAC
 TGGCACCTATGTTTATAACCATCTCACTCCTCTTCGGGACTGGGCGCACAA
 CGGCTTTCGAGATCTGGCCGTGGCTGTAGAGCCAGTCGTCTTCTCCCAAAT
 GGAGACCAAGCTCATCACGTGGGGGGCAGATACCGCCGCGTGCGG-3300
 TGACATCATCAACGGCTTGCCTGTTTCCGCCCCGAGGGGCGGGAGATACT
 GCTCGGGCCAGCCGATGGAATGGTCTCCAAGGGGTGGAGGTGCTGGCGCC
 CATCACGGCGTACGCCAGCAGACAAGGGGCCTCCTAGGGTGCATAATCAC
 CAGCCTAACTGGCCGGGACAAAAACCAAGTGGAGGGTGGAGTCCAGATTGT
 GTCAACTGCTGCCCAAACCTTCTTGCAACGTGCATCAATGGGGTGTGCTG
 GACTGTCTACCACGGGGCCGGAACGAGGACCATCGCGTCAACCAA-3600
 GGGTCTGTCTACCATGATGTATACCAATGTAGACCAAGACCTTGTGGGCTG
 GCGCGCTCCGCAAGGTAGCCGCTCATTGACACCCTGCACTTGC GGCTCCTC
 GGACCTTTACCTGGTACGAGGCACGCCGATGTCATTCCCGTGC GGCGGCG
 GGGTGATAGCAGGGGCAGCCTGCTGTGCCCCGGGCCATTTCTTACTTGAA
 AGGCTCCTCGGGGGTCCGCTGTTGTGCCCCGGGGCACGCCGTGGGCAT
 ATTTAGGGCCGCGGTGTGCACCCGTGGAGTGGCTAAGGCGGTGGA-3900
 CTTTATCCCTGTGGAGAACCTAGAGACAACCATGAGGTCCCCGGTGTTCAC
 GGATAACTCCTCTCCACCAGTAGTGCCCCAGAGCTTCCAGGTGGCTCACCT
 CCATGCTCCACAGGCAGCGGCAAAAGCACCAAGGTCCCGGCTGCATATGC
 AGCTCAGGGCTATAAGGTGCTAGTACTCAACCCCTCTGTTGCTGCAACACT
 GGGCTTTGGTGCTTACATGTCCAAGGCTCATGGGATCGATCCTAACATCAG
 GACCGGGGTGAGAACAATTACCACTGGCAGCCCCATCACGTAATC-4200

FIG. 1-2

CACCTACGGCAAGTTCCTTGCCGACGGCGGGTGCTCGGGGGGCGCTTATGA
CATAATAATTTGTGACGAGTGCCACTCCACGGATGCCACATCCATCTTGGG
CATCGGCACTGTCTTGACCAAGCAGAGACTGCGGGGGCGAGACTGGTTGT
GCTCGCCACCGCCACCCCTCCGGGCTCCGTCACTGTGCCCCATCCCAACAT
CGAGGAGGTTGCTCTGTCCACCACGGAGAGATCCCTTTTTACGGCAAGGC
TATCCCCCTCGAAGTAATCAAGGGGGGAGACATCTCATCTTCTG-4500
TCATTCAAAGAAGAAGTGCGACGAACTCGCCGCAAAGCTGGTCGATTGGG
CATCAATGCCGTGGCCTACTACCGCGGTCTTGACGTGTCCGTCACTCCCGAC
CAGCGGCGATGTTGTGTCGTGGCAACCGATGCCCTCATGACCGGCTATAC
CGGCGACTTCGACTCGGTGATAGACTGCAATACGTGTGTCAACCAGACAGT
CGATTTGAGCCTTGACCCTACCTTCACCATTGAGACAATCACGCTCCCCCA
GGATGCTGTCTCCCGCACTCAACGTGCGGGCAGGACTGGCAGGGG-4800
GAAGCCAGGCATCTACAGATTTGTGGCACCGGGGGAGCGCCCTCCGGCAT
GTTGCACTCGTCCGTCTCTGTGAGTGCTATGACGCAGGCTGTGCTTGGTA
TGAGCTCACGCCCCGCGAGACTACAGTTAGGCTACGAGCGTACATGAACAC
CCCGGGGCTTCCCGTGTGCCAGGACCATCTTGAATTTTGGGAGGGCGTCTT
TACAGGCCTCACTCATATAGATGCCCACTTTCTATCCCAGACAAAGCAGAG
TGGGGAGAACCTTCCTTACCTGGTAGCGTACCAAGCCACCGTGTG-5100
CGCTAGGGCTCAAGCCCCCTCCCCCATCGTGGGACCAGATGTGGAAGTGTTT
GATTCGCCTCAAGCCCCACCCTCCATGGGCCAACCCCCCTGCTATACAGACT
GGGCGCTGTTTCAAGATGAAATCACCTGACGCACCCAGTCACCAAATACAT
CATGACATGCATGTGCGCCGACCTGGAGGTCGTCACGAGCACCTGGGTGCT
CGTTGGCGGCGTCTGCTGCTTTGGCCGCGTATTGCCTGTCAACAGGCTG
CGTGGTCATAGTGGGCAGGGTCGTCTTGTCGGGAAGCCGGCAAT-5400
CATACCTGACAGGGAAGTCCTCTACCGAGAGTTCGATGAGATGGAAGAGTG
CTCTCAGCACTTACCGTACATCGAGCAAGGGATGATGCTCGCCGAGCAGTT
CAAGCAGAAGGCCCTCGGCCTCCTGCAGACCGCGTCCCGTCAGGCAGAGGT
TATCGCCCCCTGCTGTCCAGACCAACTGGCAAAACTCGAGACCTTCTGGGC
GAAGCATATGTGGAACCTTCATCAGTGGGATACAATACTTGGCGGGCTTGTC
AACGCTGCCTGGTAACCCCGCCATTGCTTCATTGATGGCTTTTAC-5700
AGCTGCTGTCAACAGCCCACTAACCACTAGCCAAACCCTCCTCTTCAACAT
ATTGGGGGGGTGGGTGGCTGCCAGCTCGCCGCCCCCGGTGCCGCTACTGC
CTTTGTGGGCGCTGGCTTAGCTGGCGCCGCCATCGGCAGTGTTGGACTGGG
GAAGGTCCTCATAGACATCCTTGAGGGTATGGCGCGGGCGTGGCGGGAGC
TCTTGTGGCATTCAGATCATGAGCGGTGAGGTCCCCTCCACGGAGGACCT
GGTCAATCTACTGCCCGCCATCCTCTCGCCCGGAGCCCTCGTAGT-6000
CGGCGTGGTCTGTGCAGCAATACTGCGCCGGCACGTTGGCCCCGGGCGAGGG
GGCAGTGCAGTGGATGAACCGGCTGATAGCCTTCGCCTCCCCGGGGGAACCA
TGTTTCCCCACGCACTACGTGCCGGAGAGCGATGCAGCTGCCCGCGTCAAC
TGCCATACTCAGCAGCCTCACTGTAACCCAGCTCCTGAGGCGACTGCACCA
GTGGATAAGCTCGGAGTGTACCACTCCATGCTCCGGTTCCTGGCTAAGGGA
CATCTGGGACTGGATATGCGAGGTGTTGAGCGACTTTAAGACCTG-6300
GCTAAAAGCTAAGCTCATGCCACAGCTGCCTGGGATCCCCTTTGTGTCTTG
CCAGCGCGGGTATAAGGGGGTCTGGCGAGTGGACGGCATCATGCACACTCG
CTGCCACTGTGGAGCTGAGATCACTGGACATGTCAAAAACGGGACGATGAG
GATCGTCGGTCTTAGGACCTGCAGGAACATGTGGAGTGGGACCTTCCCAT
TAATGCCTACACCACGGGCCCCCTGTACCCCCCTTCTGCGCCGAACCTACAC
GTTGCGCTATGGAGGGTGTCTGCAGAGGAATATGTGGAGATAAG-6600

FIG. 1-3

GCAGGTGGGGGACTTCCACTACGTGACGGGTATGACTACTGACAATCTCAA
 ATGCCCCGTGCCAGGTCCCATCGCCCCGAATTTTTCACAGAATTGGACGGGGT
 GCGCCTACATAGGTTTTCGCCCCCTGCAAGCCCTTGCTGCGGGAGGAGGT
 ATCATTGAGAGTAGGACTCCACGAATACCCGGTAGGGTCGCAATTACCTTG
 CGAGCCCGAACCGGACGTGGCCGTGTTGACGTCCATGCTCACTGATCCCTC
 CCATATAACAGCAGAGGCGGCCGGCGAAGGTTGGCGAGGGGATC-6900
 ACCCCCCCTCTGTGGCCAGCTCCTCGGCTAGCCAGCTATCCGCTCCATCTCT
 CAAGGCAACTTGCACCGCTAACCATGACTCCCCTGATGCTGAGCTCATAGA
 GGCCAACCTCCTATGGAGGCAGGAGATGGGCGGCAACATCACCAGGGTTGA
 GTCAGAAAACAAAGTGGTGATTCTGGACTCCTTCGATCCGCTTGTGGCGGA
 GGAGGACGAGCGGGAGATCTCCGTACCCGCAGAAATCCTGCGGAAGTCTCG
 GAGATTGCGCCAGGCCCTGCCCGTTTGGGCGCGGCCGGACTATAA-7200
 CCCCCCGTAGTGGAGACGTGGAAAAAGCCCGACTACGAACCACCTGTGGT
 CCATGGCTGTCCGCTTCCACCTCCAAAGTCCCCTCCTGTGCCTCCGCTCG
 GAAGAAGCGGACGGTGGTCTCACTGAATCAACCCTATCTACTGCCTTGGC
 CGAGCTCGCCACCAGAAGCTTTGGCAGCTCCTCAACTTCCGGCATTACGGG
 CGACAATACGACAACATCCTCTGAGCCCCGCCCTTCTGGCTGCCCCCCGA
 CTCCGACGCTGAGTCCTATTCTCCATGCCCCCCTGGAGGGGGA-7500
 GCCTGGGGATCCGGATCTTAGCGACGGGTCAATGGTCAACGGTCAGTAGTGA
 GGCCAACGCGGAGGATGTCTGTGTGCTGCTCAATGTCTTACTCTTGGACAGG
 CGCACTCGTCACCCCGTGCGCCGCGGAAGAACAGAACTGCCCATCAATGC
 ACTAAGCAACTCGTTGCTACGTCAACCAATTTGGTGTATTCCACCACCTC
 ACGCAGTGCTTGCCAAAGGCAGAAGAAAGTCACATTTGACAGAGTCAAGT
 TCTGGACAGCCATTACCAGGACGTAATCAAGGAGGTTAAAGCAGC-7800
 GGCGTCAAAAGTGAAGGCTAACTTGCTATCCGTAGAGGAAGCTTGCAGCCT
 GACGCCCCCAGCTCAGCCAAATCCAAGTTTGGTTATGGGGCAAAAGACGT
 CCGTTGCCATGCCAGAAAGGCCGTAAACCCACATCAACTCCGTGTGGAAAGA
 CTTTCTGGAAGACAATGTAACACCAATAGACACTACCATCATGGCTAAGAA
 CGAGGTTTTCTGCGTTCAGCCTGAGAAGGGGGGTGTAAGCCAGCTCGTCT
 CATCGTGTTCCCCGATCTGGGCGTGCCTGTGCGAAAAGATGGC-8100
 TTTGTACGACGTGGTTACAAAGCTCCCCTTGCCCGTGATGGGAAGCTCCTA
 CGGATTCCAATACTCACCAGGACAGCGGGTTGAATTCCTCGTGCAAGCGTG
 GAAGTCCAAGAAAACCCCAATGGGGTTCTCGTATGATACCCGCTGCTTTGA
 CTCCACAGTCACTGAGAGCGACATCCGTACGGAGGCAATCTACCAATG
 TTGTGACCTCGACCCCCAAGCCCGCGTGCCATCAAGTCCCTCACCAGAGG
 GCTTTATGTTGGGGGCCCTCTTACCAATTCAAGGGGGGAGAACTG-8400
 CGGCTATCGCAGGTGCCGCGGAGCGGCGTACTGACAACTAGCTGTGGTAA
 CACCCTCACTTGCTACATCAAGGCCCGGGCAGCCTGTCGAGCCGCAGGGCT
 CCAGGACTGCACCATGCTCGTGTGTGGCGACGACTTAGTCGTTATCTGTGA
 AAGCGCGGGGGTCCAGGAGGACGCGCGAGCCTGAGAGCCTTCACGGAGGC
 TATGACCAGGTAATCCGCCCCCCTGGGGACCCCCACAACCAGAAATACGA
 CTTGGAGCTCATAACATCATGCTCCTCCAACGTGTCAGTCGCCCCA-8700
 CGACGGCGCTGGAAAGAGGGTCTACTACCTCACCCTGACCCTACAACCCC
 CCTCGCGAGAGCTGCGTGGGAGACAGCAAGACACACTCCAGTCAATTCCTG
 GCTAGGCAACATAATCATGTTTGCCCCACACTGTGGGCGAGGATGATACT
 GATGACCCATTTCTTTAGCGTCTTATAGCCAGGGACCAGCTTGAACAGGC
 CCTCGATTGCGAGATCTACGGGGCCTGCTACTCCATAGAACCCTTGATCT
 ACCTCCAATCATTCAAAGACTCCATGGCCTCAGCGCATTTTCACT-9000

FIG. 1-4

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CCACAGTTACTCTCCAGGTGAAATTAATAGGGTGGCCGCATGCCTCAGAAA
ACTTGGGGTACCGCCCTTGCGAGCTTGGAGACACCGGGCCCGGAGCGTCCG
CGCTAGGCTTCTGGCCAGAGGAGGCAGGGCTGCCATATGTGGCAAGTACCT
CTTCAACTGGGCAGTAAGAACAAAGCTCAAAC

FIG. 1-5

FIG. 2 Translation of DNA 35

SerIleGluThrIleThrLeuProGlnAspAlaValSerArgThrGlnArgArgGlyArg
 1 TCCATTGAGACAATCACGCTCCCCAGGATGCTGTCTCCCGCACTCAACGTCGGGGCAGG
 AGGTAACTCTGTTAGTGCAGGGGTCCCTACGACAGAGGGCGTGAGTTGCAGCCCCCGTCC
 ThrGlyArgGlyLysProGlyIleTyrArgPheValAlaProGlyGluArgProSerGly
 61 ACTGGCAGGGGAGCCAGGCATCTACAGATTGTGTGGCACCGGGGAGCGCCCTCCGGC
 TGACCGTCCCCCTTCGGTCCGTAGATGTCTAAACACCGTGGCCCCCTCGCGGGAGGCCG
 MetPheAspSerSerValLeuCysGluCysTyrAspAlaGlyCysAlaTrpTyrGluLeu
 121 ATGTTCCGACTCGTCCGTCCTCTGTGAGTGCTATGACGCAGGCTGTGCTTGGTATGAGCTC
 TACAAGCTGAGCAGGAGGAGACACTCAGATACTGCGTCCGACACGAACCATACTCGAG
 ThrProAlaGluThrThrValArgLeuArgAlaTyrMetAsnThrProGlyLeuProVal
 181 ACGCCCGCCGAGACTACAGTTAGGCTACGAGCGTACATGAACACCCCGGGCTTCCCGTG
 TCGGGCGGCTCTGATGTCAATCCGATGCTCGCATGTACTTGTGGGCCCCGGAAGGCAC
 CysGlnAspHisLeuGluPheTrpGluGlyValPheThrGlyLeuThrHisIleAspAla
 241 TGCCAGGACCATCTTGAATTTGGGAGGGCGTCTTTACAGGCCCTCACTCATATAGATGCC
 ACGGTCCCTGGTAGAACTTAAACCCCTCCCGCAGAAATGTCCGGAGTGAGTATATCTACGG
 HisPheLeuSerGlnThrLysGlnSerGlyGluAsnLeuProTyrLeuValAlaTyrGln
 301 CACTTTCATCCAGACAAGCAGAGTGGGAGAACCTTCTTACCTGGTAGCGTACCAA
 GTGAAAGATAGGGTCTGTTTCGTCTCACCCCTCTTGGAAAGGAATGGACCATCGCATGGTT
 AlaThrValCysAlaArgAlaGlnAlaProProProSerTrpAspGlnMetTrpLysCys
 361 GCCACCGTGTGCGCTAGGGCTCAAGCCCCCTCCCCCATCGTGGGACCAAGATGTGGAAGTGT
 CGGTGGCACACGCGATCCCGAGTTCGGGAGGGGTAGCACCCCTGTCTACACCTTCACA
 LeuIleArgLeuLysProThrLeuHisGlyProThrProLeuLeuTyrArgLeuGlyAla
 421 TTGATTGCGCCTCAAGCCCCACCTCCATGGGCCCAACACCCCTGTCTATACAGACTGGGCGCT
 AACTAAGCGGAGTTCGGGTGGGAGGTACCCGGTTGTGGGACGATATGTCTGACCCCGCGA



European
Patent Office

EUROPEAN SEARCH REPORT

Application Number

EP 90 30 9120

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			TECHNICAL FIELDS SEARCHED (Int. Cl.8)
			A 61 K C 07 K C 12 N
The present search report has been drawn up for all claims			
Place of search		Date of completion of search	Examiner
The Hague		23 November 90	FERNANDEZ Y BRANAS F
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Y: particularly relevant if combined with another document of the same category		D: document cited in the application	
A: technological background		L: document cited for other reasons	
O: non-written disclosure		-----	
P: intermediate document		&: member of the same patent family, corresponding document	
T: theory or principle underlying the invention			

(19)



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(11)

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C12N 7/00, G01N 33/576

(21) Application number: 90309120.5

(22) Date of filing: 21.08.1990

(54) **Methods for culturing HCV in B- or T-lymphocyte cell lines**

Verfahren zur HCV-Züchtung in Zell-Linien aus B- oder T-Lymphozyten

Procédé de culture du HCV dans des lignées cellulaire dérivée des lymphocytes-B ou -T

(84) Designated Contracting States:
AT BE CH DE DK ES FR GB GR IT LI LU NL SE

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Note: Within nine months from the publication of the mention of the grant of the European patent, any person may give notice to the European Patent Office of opposition to the European patent granted. Notice of opposition shall be filed in a written reasoned statement. It shall not be deemed to have been filed until the opposition fee has been paid. (Art. 99(1) European Patent Convention).

DescriptionTechnical Field

- 5 The invention relates to *in vitro* cell systems for the replication of a virus; more specifically, it relates to cell lines in which an etiologic agent of non-A non-B hepatitis (NANBH), hepatitis C virus (HCV) will replicate.

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Background

- 50 Non-A, Non-B hepatitis (NANBH) is a transmissible disease or family of diseases that are believed to be viral-induced, and that are distinguishable from other forms of viral-associated liver diseases, including that caused by the known hepatitis viruses, i.e., hepatitis A virus (HAV), hepatitis B virus (HBV), and delta hepatitis virus (HDV), as well as the hepatitis induced by cytomegalovirus (CMV) or Epstein-Barr virus (EBV). NANBH was first identified in transfused individuals. Transmission from man to chimpanzee and serial passage in chimpanzees provided evidence that NANBH is due to a transmissible infectious agent or agents. A viral agent, HCV, has been identified as an etiologic agent of
- 55 NANBH.

The demand for sensitive, specific methods for screening and identifying carriers of NANBV and NANBV contaminated blood or blood products is significant. Post-transfusion hepatitis (PTH) occurs in approximately 10% of transfused patients, and NANBH accounts for up to 90% of these cases. The major problem in this disease is the frequent pro-

gression to chronic liver damage (25-55%).

Patient care as well as the prevention of transmission of NANBH by blood and blood products or by close personal contact require reliable diagnostic and prognostic tools to detect nucleic acids, antigens and antibodies related to NANBV. In addition, there is also a need for effective vaccines and immunotherapeutic therapeutic agents for the prevention and/or treatment of the disease. EP-A-318 216 provides a family of cDNA sequences derived from Hepatitis C virus.

Tissue culture cells infected with an etiologic agent which causes NANBH would provide a source of viral polypeptides which could be used for immunoassays and/or for vaccines. In addition, they could be useful in drug screening programs in the development of antiviral agents for treating the disease.

Immortalized human or non-human primate cell lines which are derived from liver, and are purportedly infected with an etiologic agent causative of NANBH have been reported in WO/87/05930, filed 1 April 1987, and published 8 October 1987. These cell lines are reportedly non-lymphocytic in nature. In addition, it has not been established that the agent which replicates in these cells is causative of the disease.

It has also been reported that a preparation of mononuclear leukocytes, which purportedly were mainly lymphocytes, isolated from a patient with NANBH, caused NANB when infused into a susceptible chimpanzee. Hellings et al. (1985). However, proof of NANBH infection was indirect, in that it rested primarily on morphological and histological changes in the purportedly infected chimpanzee. Moreover, only one of the two chimpanzees receiving the blood cells was purportedly infected.

Disclosure of the Invention

The present invention is directed to cells that may be used to propagate HCV in culture. The present invention is also directed to viral antigens produced by such cultures, and use such antigens, for example, in diagnostics.

Accordingly, one aspect of the invention is a cell line infected with HCV, wherein the cells in the cell line are B lymphocytes or T lymphocytes and are capable of replicating HCV.

Another aspect of the invention is a method of preparing a composition for the detection of an anti-HCV antibody which comprises: purifying or partially purifying HCV virus particles or polypeptides from a cell line of the invention; and fixing said particles or polypeptides to a solid phase.

The composition prepared by the invention may be used in an immunoassay for detecting antibodies directed against an HCV antigen comprising:

- (a) incubating a sample suspected of containing anti-HCV antibodies with an HCV antigen produced by an *in vitro* cell line, under conditions which allow the formation of an antibody-antigen complex; and
- (b) detecting the antibody-antigen complex containing the HCV antigen.

Another aspect of the invention is a method for screening for an agent which inhibits HCV replication comprising:

- (a) providing a composition comprised of a putative anti-viral agent;
- (b) providing a cell line capable of replicating HCV;
- (c) providing a composition comprised of infective HCV;
- (d) incubating the compositions of (a) and (c) with the cell line of (b) under conditions which would, in the absence of (a) allow infection of and replication of HCV in the cell line; and
- (e) detecting an inhibition of viral replication after the incubating in (d), wherein the cells in the cell line of (b) are B lymphocytes or T lymphocytes.

Still another application of the invention is a method for proliferating virus comprising growing in culture HCV infected cells.

Brief Description of the Drawings

Figure 1 shows a composite sequence of the HCV cDNA sense strand derived from HCV strain HCV1.

Figure 2 shows the nucleotide sequence of HCV cDNA clone 35, and the amino acids encoded therein.

Modes for Carrying Out the Invention

I. Definitions

The term "hepatitis C virus" has been reserved by workers in the field for an heretofore unknown etiologic agent

of NANBH. Accordingly, as used herein, "hepatitis C virus" (HCV) refers to an agent causative of NANBH, which was formerly referred to as NANBV and/or BB-NANBV. The terms HCV, NANBV, and BB-NANBV are used interchangeably herein. As an extension of this terminology, the disease caused by HCV, formerly called NANB hepatitis (NANBH), is called hepatitis C. The terms NANBH and hepatitis C may be used interchangeably herein.

The term "HCV", as used herein, denotes a viral species of which pathogenic strains cause NANBH, and attenuated strains or defective interfering particles derived therefrom. The HCV genome is comprised of RNA. It is known that RNA containing viruses have relatively high rates of spontaneous mutation, i.e., reportedly on the order of 10^{-3} to 10^{-4} per incorporated nucleotide (Fields & Knipe (1986)). Therefore, there can be multiple strains, which may be virulent or avirulent, within the HCV species. It is believed that HCV is a Flavi-like virus. The morphology and composition of Flavivirus particles are known, and are discussed in Brinton (1986). Generally, with respect to morphology, Flaviviruses contain a central nucleocapsid surrounded by a lipid bilayer. Virions are spherical and have a diameter of about 40-50 nm. Their cores are about 25-30 nm in diameter. Along the outer surface of the virion envelope are projections that are about 5-10 nm long with terminal knobs about 2 nm in diameter.

cDNAs to the genome of a strain of HCV, denoted HCV1, are described in EP-A-388 232. See also EPO Publication No. 0 318 216 and PCT App. No. US88/04125. A composite sequence of the HCV cDNA sense strand is shown in Figure 1. Different strains or isolates of HCV are expected to contain variations at the amino acid and nucleic acids levels compared with HCV1. Many isolates are expected to show much (i.e., more than about 40%) homology in the total amino acid sequence compared with HCV1. However, it may also be found that other less homologous HCV isolates would be defined as HCV strains according to various criteria such as an ORF encoding a polypeptide similar in size to that of HCV1, an encoded polypeptide of similar hydrophobic and antigenic character to that of HCV1, and the presence of co-linear peptide sequences that are conserved with HCV1.

HCV encodes at least one epitope which is immunologically identifiable with an epitope encoded in the HCV1 genome. The epitope is unique to HCV when compared to other known Flaviviruses. The uniqueness of the epitope may be determined by its immunological reactivity with anti-HCV antibodies and lack of immunological reactivity with antibodies to other Flavivirus species. Methods for determining immunological reactivity are known in the art, for example, by radioimmunoassay, by ELISA assay, and by hemagglutination. Examples of suitable techniques for determining immunological reactivity of HCV epitopes are described in EP-A-388 232, in Choo et al. (1989), and in Kuo et al. (1989).

In addition to the above, the following parameters of nucleic acid homology and amino acid homology are applicable, either alone or in combination, in identifying a strain or isolate as HCV. Since HCV strains and isolates are evolutionarily related, it is expected that the overall homology of the genomes at the nucleotide level probably will be about 40% or greater, probably about 60% or greater, and even more probably about 80% or greater; and in addition that there will be corresponding contiguous sequences of at least about 13 nucleotides. The correspondence between the putative HCV strain genomic sequence and the HCV1 cDNA sequence can be determined by techniques known in the art. For example, they can be determined by a direct comparison of the sequence information of the polynucleotide from the putative HCV, and the HCV cDNA sequence(s) described herein. For example, also, they can be determined by hybridization of the polynucleotides under conditions which form stable duplexes between homologous regions (for example, those which would be used prior to S_1 digestion), followed by digestion with single stranded specific nuclease (s), followed by size determination of the digested fragments.

Because of the evolutionary relationship of the strains or isolates of HCV, putative HCV strains or isolates are identifiable by their homology at the polypeptide level. Generally, HCV strains or isolates are expected to be more than about 40% homologous, probably more than about 70% homologous, and even more probably more than about 80% homologous, and some may even be more than about 90% homologous at the polypeptide level. The techniques for determining amino acid sequence homology are known in the art. For example, the amino acid sequence may be determined directly and compared to the sequences provided herein. Alternatively the nucleotide sequence of the genomic material of the putative HCV may be determined (usually via a cDNA intermediate), the amino acid sequence encoded therein can be determined, and the corresponding regions compared.

As used herein, a polynucleotide "derived from" a designated polynucleotide sequence refers to a polynucleotide sequence which is comprised of a sequence of approximately at least about 6 nucleotides, preferably at least about 8 nucleotides, more preferably at least about 10-12 nucleotides, and even more preferably at least about 15-20 nucleotides corresponding to a region of the designated nucleotide sequence. "Corresponding" means homologous to or complementary to the designated sequence. Preferably, the sequence of the region from which the polynucleotide is derived is homologous to or complementary to a sequence which is unique to an HCV genome. Whether or not a sequence is unique to the HCV genome can be determined by techniques known to those of skill in the art. For example, the sequence can be compared to sequences in databanks, e.g., Genbank, to determine whether it is present in the uninfected host or other organisms. The sequence can also be compared to the known sequences of other viral agents, including those which are known to induce hepatitis, e.g., HAV, HBV, and HDV, and to other members of the Flaviviridae. The correspondence or non-correspondence of the derived sequence to other sequences can also be determined by

hybridization under the appropriate stringency conditions. Hybridization techniques for determining the complementarity of nucleic acid sequences are known in the art, and are discussed *infra*. See also, for example, Maniatis et al. (1982). In addition, mismatches of duplex polynucleotides formed by hybridization can be determined by known techniques, including for example, digestion with a nuclease such as S1 that specifically digests single-stranded areas in duplex polynucleotides. Regions from which typical DNA sequences may be "derived" include but are not limited to, for example, regions encoding specific epitopes, as well as non-transcribed and/or non-translated regions.

The derived polynucleotide is not necessarily physically derived from the nucleotide sequence shown, but may be generated in any manner, including for example, chemical synthesis or DNA replication or reverse transcription or transcription. In addition, combinations of regions corresponding to that of the designated sequence may be modified in ways known in the art to be consistent with an intended use.

The term "polynucleotide" as used herein refers to a polymeric form of nucleotides of any length, either ribonucleotides or deoxyribonucleotides. This term refers only to the primary structure of the molecule. Thus, this term includes double- and single-stranded DNA, as well as double- and single stranded RNA. It also includes modified, for example, methylation capping and/or by other modifications known in the art, including the substitution of an analog for one or more of the naturally occurring nucleotide bases, as well as unmodified forms of the polynucleotide.

The term "purified viral polynucleotide" refers to a viral genome or fragment thereof which is essentially free, i.e., contains less than about 50%, preferably less than about 70%, and even more preferably less than about 90% of polypeptides with which the viral polynucleotide is naturally associated. Techniques for purifying viral polynucleotides from viral particles are known in the art, and include for example, disruption of the particle with a chaotropic agent, differential extraction and separation of the polynucleotide(s) and polypeptides by ion-exchange chromatography, affinity chromatography, and sedimentation according to density.

An "open reading frame" (ORF) is a region of a polynucleotide sequence which encodes a polypeptide; this region may represent a portion of a coding sequence or a total coding sequence.

A "coding sequence" is a polynucleotide sequence which is transcribed into mRNA and/or translated into a polypeptide when placed under the control of appropriate regulatory sequences. The boundaries of the coding sequence are determined by a translation start codon at the 5'-terminus and a translation stop codon at the 3'-terminus. A coding sequence can include, but is not limited to mRNA, cDNA, and recombinant polynucleotide sequences.

"Immunologically identifiable with/as" refers to the presence of epitope(s) and polypeptides(s) which are also present in the designated polypeptide(s), usually HCV proteins. Immunological identity may be determined by antibody binding and/or competition in binding; these techniques are known to those of average skill in the art, and are also illustrated *infra*.

As used herein, "epitope" refers to an antigenic determinant of a polypeptide; an epitope could comprise 3 or more amino acids in a spatial conformation which is unique to the epitope. Generally an epitope consists of at least 5 such amino acids, and more usually, consists of at least 8-10 such amino acids. Methods of determining the spatial conformation of amino acids are known in the art, and include, for example, X-ray crystallography and 2-dimensional nuclear magnetic resonance.

A polypeptide is "immunologically reactive" with an antibody when it binds to an antibody due to antibody recognition of a specific epitope contained within the polypeptide. Immunological reactivity may be determined by antibody binding, more particularly by the kinetics of antibody binding, and/or by competition in binding using as competitor(s) a known polypeptide(s) containing an epitope against which the antibody is directed. The techniques for determining whether a polypeptide is immunologically reactive with an antibody are known in the art.

As used herein, the term "immunogenic polypeptide" refers to a polypeptide that elicits a cellular and/or humoral response, whether alone or when linked to a carrier in the presence or absence of an adjuvant.

The term "polypeptide" refers to a polymer of amino acids and does not refer to a specific length of the product; thus, peptides, oligopeptides, and proteins are included within the definition of polypeptide. This term also does not refer to or exclude post-expression modifications of the polypeptide, for example, glycosylations, acetylations, phosphorylations and the like.

An "individual", as used herein, refers to vertebrates, particularly members of the mammalian species, and includes but is not limited to domestic animals, sports animals, and primates, including humans.

As used herein, the "sense strand" or "plus strand" of a nucleic acid contains the sequence that has sequence homology to that of mRNA. The "antisense strand" or "minus strand" contains a sequence which is complementary to that of the "sense strand" or "plus strand".

As used herein, a "positive stranded genome" of a virus is one in which the genome, whether RNA or DNA, is single-stranded and which encodes a viral polypeptide(s). Examples of positive stranded RNA viruses include Togaviridae, Coronaviridae, Retroviridae, Picomaviridae, and Caliciviridae. Included also, are the Flaviviridae, which were formerly classified as Togaviridae. See Fields & Knipe (1986).

As used herein, a "replicative intermediate" of an HCV genome refers to an RNA strand or fragment thereof, which is complementary to the viral genome, and which is synthesized during the course of viral replication; the replicative

intermediate functions as a template for the synthesis of positive RNA strands.

As used herein, "purified HCV" refers to a preparation of HCV which has been isolated from the cellular constituents with which the virus is normally associated, and from other types of viruses which may be present in the infected tissue. The techniques for isolating viruses are known to those of skill in the art, and include, for example, centrifugation and affinity chromatography; a method of preparing purified HCV is discussed *infra*.

As used herein, a "biological sample" refers to a sample of tissue or fluid isolated from an individual, including but not limited to, for example, plasma, serum, spinal fluid, lymph fluid, the external sections of the skin, respiratory, intestinal, and genitourinary tracts, tears, saliva, milk, blood cells, tumors, organs, and also samples of *in vitro* cell culture constituents (including but not limited to conditioned medium resulting from the growth of cells in cell culture medium, putatively virally infected cells, recombinant cells, and cell components).

The term "oligonucleotide", as used herein, refers to primers, probes, oligomer fragments to be detected, oligomer controls and unlabeled blocking oligomers, and is defined as a molecule comprised of three or more deoxyribonucleotides or ribonucleotides, typically contains more than four, typically a maximum of about 100 nucleotides. Its exact size will depend upon many factors, which in turn depend on the ultimate function or use of the oligonucleotide.

The term "primer", as used herein, refers to an oligonucleotide which is capable of acting as a point of initiation of polynucleotide synthesis along a complementary strand when placed under conditions in which synthesis of a primer extension product which is complementary to a polynucleotide is catalyzed. Such conditions include the presence of four different nucleotide triphosphates or nucleoside analogs and an agent for polymerization such as DNA polymerase or reverse transcriptase, in an appropriate buffer ("buffer" includes substituents which are cofactors, or which affect pH, ionic strength, etc.), and at a suitable temperature.

As used herein, the term "target region" refers to a region of the nucleic acid which is to be amplified and/or detected.

As used herein, the term "probe" refers to a polynucleotide or oligonucleotide which forms a hybrid structure with a sequence in a target region, due to complementarity of at least one sequence in the probe with a sequence in the target region. Preferably the probe, however, does not contain a sequence complementary to sequence(s) used to prime the polymerase chain reaction.

As used herein, the term "thermostable nucleotide polymerase" refers to an enzyme which is relatively stable to heat when compared to nucleotide polymerases from *E. coli* and which catalyzes the polymerization of nucleotides. Generally, the enzyme will initiate synthesis at the 3'-end of the target sequence utilizing the primer, and will proceed in the 5'-direction along the template until synthesis terminates. A representative thermostable enzyme isolated from *Thermus aquaticus* (Taq), and method for using it is described in Saiki et al. (1988).

As used herein, the term "viral RNA", which includes HCV RNA, refers to RNA from the viral genome, fragments thereof, transcripts thereof, and mutant sequences derived therefrom.

As used herein, "cell line" refers to a propagated culture of cells after the first subculture, thus there are various embodiments including but not limited to individual cells, harvested cells, and cultures containing cells so long as these are derived from cells of the cell line referred to. By "derived" is meant progeny or issue. It is further known in the art that spontaneous or induced changes can occur in karyotype during storage or transfer. Therefore, cells derived from the cell line referred to may not be precisely identical to the ancestral cells or cultures, and the cell line referred to includes such variants. The term "cell lines" includes immortalized cells. Preferably, cell lines exclude cells which are derived by the hybridization of three cell types, for example, a trioma.

As used herein, an "*in vitro*" cell system refers to cells which have replicated *in vitro*, thus the term includes primary cultures and cell lines.

As used herein, the term "blood cell" refers to cells found either circulating in the blood, in tissue, or in bone marrow, and includes cells of the myelocytic series, the monocytic series, the megakaryocytic series, the erythrocytic series, the lymphocytic series, and the plasmocytic series. The morphology by which these cells can be distinguished is discussed in THE MORPHOLOGY OF HUMAN BLOOD CELLS (1985).

As used herein, the term "peripheral blood cell" refers to a cell found circulating in the blood, and refers generally to eosinophils, neutrophils (both band and segmented), basophils, monocytes, thrombocytes, erythrocytes, lymphocytes, and plasmocytes.

As used herein, the term "leukocyte" refers to a heterogeneous group of blood cell types which includes everything except erythrocytes and platelets.

As used herein, the term "lymphocyte" refers to a spherical cell, 6-12 μ m in diameter, involved in immune responses, with a large, round, and deeply staining nucleus and very little cytoplasm. Lymphocytes include B lymphocytes and T lymphocytes.

As used herein, "B lymphocytes" are vertebrate white blood cells which, when mature, are involved in the production of antibody. Mature virgin B lymphocytes (i.e., those that have not encountered antigen) are characterized by the presence of the following surface markers: monomeric IgM receptors for antigen, Fc receptors for IgG; and receptors for the activated form of C3b component of complement.

Precursor B lymphocytes lack immunoglobulin products but may express other characteristic molecules that can

be identified by specific monoclonal antibodies. These cells divide and undergo rapid transition to become large lymphoblasts with cytoplasmic μ heavy chains but no light chains. The pre-B cell phenotype (cytoplasmic μ chain $^{+}$, light chain $^{-}$, surface Ig $^{-}$) can first be demonstrated in lymphoblasts actively engaged in DNA synthesis. Human pre-B cells express HLA-DR molecules on their surface and may also express receptors for the C3b complement fragment. However, they lack Fc receptors for IgG and have very few C3d/EBV receptors. Precursor B lymphocytes can also be recognized by their capacity to bind peanut agglutinin.

Mature B lymphocytes may be either activated or resting. Activated B cells express several types of molecules that are not detected on resting B cells. These marker molecules are known by those of skill in the art, and include, for example, interleukin-2 (IL-2) receptors, and transferrin receptors (the latter are expressed by all kinds of dividing cells). Activation of B cells provokes loss of membrane IgD, and both Fc $_{\gamma}$ and C3b receptors are diminished on external membranes, whereas those for C3d/Epstein Barr Virus (EBV) are not significantly affected.

After antigen or mitogen stimulation, B lymphocytes may proceed along either of 2 branches of a differentiation pathway. They can differentiate into plasma cells, or they can divide and then return to a resting state as memory B cells. Memory B cells have had a loss of IgD during activation; this molecule is not resynthesized by cells entering the memory cell pool.

"Plasma cells" are terminally differentiated B lymphocytes. These cells are of ovoid shape, with an eccentric spoke-wheel nucleus, and intensely basophilic cytoplasm. When stained with fluorochrome-labeled antibodies to immunoglobulin determinants, fixed plasma cells display intense fluorescence throughout their cytoplasm. Membrane-bound immunoglobulin and DR molecules are scant, and receptors for Fc $_{\gamma}$, C3b, C3d, or EBV are generally undetectable.

A "plasmablast" is both morphologically and functionally between the activated lymphocyte (also called B lymphoblast) and plasma cell stages. The cells are large, with a greater nuclear:cytoplasmic ratio than plasma cells. Membrane-bound immunoglobulins and Fc $_{\gamma}$ receptors are present.

As used herein, a "T lymphocyte" is a thymus-derived cell which may participate in a variety of cell-mediated immune responses. There are a variety of types of T lymphocytes, which are distinguishable by their function, and by their cell surface markers. Methods for distinguishing the classes of T lymphocytes, which include those of the T "helper" and T "suppressor" types are known in the art. For example, monoclonal antibodies can be used to detect cell surface antigens, including the CD (cluster of differentiation) antigens. The T cell types marked by the CD antigens, and monoclonal antibodies which detect the specific antigens are shown in Table 1.

Table 1

Cell Type Marked	CD Designation	Antibody Designation	Comments
Cortical thymocytes	CD1	Leu 6 OKT 6	Early T cell antigen also present on Langerhans cells, associated with beta ₂ -microglobulin not present on peripheral T cells.
E rosette-forming cells	CD2	Leu 5 OKT 11 T11	Pan-T cell antigen SRBC receptor on T cells.
Mature T cells	CD3	Leu 4	Also present on T cell ALL and cutaneous T cell lymphoma.
Helper/inducer T cells	CD4	Leu 3 OKT 4 T4	Can be further subdivided into helper and inducer subsets.
Pan-T and -B cell subpopulation	CD5	Leu 1 T1 T101 OKT1	B cell CLL. B cells following marrow transplant.
Mature T and B cell subpopulation	CD6	T12	Malignant T cells.
Pan-T cells, thymocytes	CD7	Leu 9 3A1	T cell leukemias.
Suppressor/cytotoxic T cells	CD8	Leu 2 OHT 8 T8	Can be further subdivided into cytotoxic and suppressor subsets.

As used herein, "large granular lymphocyte" (LGL), also called natural killer (NK) cells, are another type of lymphocyte. These cells comprise a discrete population of large lymphocytes that can be distinguished morphologic features, for example, by characteristic azurophilic granules in their cytoplasm. In addition, LGL can be identified by methods which utilize specific monoclonal antibodies. Several monoclonal antibodies are available that detect either Fc receptors (Leu 11) or specific differentiation antigens (Leu 7, HNK-1, NKH-1, NKH-2) present on these cells. Some LGL cells also express antigens from the CD2 T cell family. Methods for functional testing are also known in the art, and are based on the ability of these nonimmune cells to kill special target cells, for example, erythroleukemia cell line K562. Methods of measuring cytotoxicity are known in the art, and include, for example the ^{51}Cr release assay.

As used herein, "monocytes" denotes large agranulocytic leukocyte blood cells which are phagocytic. In humans, these cells are approximately 10-12 μm in diameter, with oval nuclei. Monocytes which are in tissue are known as macrophages. Monocytes/macrophages may be identified by their morphology. They are larger than granulocytes and most lymphocytes. They have round or kidney-shaped nuclei with fine, lightly stained granules. They may be detected by staining for their nonspecific esterase, or alpha-naphthol esterase. Monoclonal antibodies directed at specific differentiation antigens are also available, and include, for example, Leu M3, and OKT M1. The term "monocytes" includes monoblasts and promonocytes. The cells of the mononuclear phagocyte system, and their localization, are shown in Table 2; methods for distinguishing the cell types are discussed in BASIC & CLINICAL IMMUNOLOGY, 6th edition.

Table 2

Cells of the Mononuclear Phagocyte System*	
Cells	Localization
Stem cells (committed)	Bone marrow
Monoblasts	Bone marrow
Promonocytes	Bone marrow
Monocytes	Bone marrow
Macrophages	Tissues
Normal state, free	
Histocytes	Connective tissues
Alveolar macrophages	Lung
Pleural and peritoneal macrophages	Serous cavities
Normal state, fixed	
Kupffer cells	Liver
Osteoclasts	Bone
Microglial cells	Nervous System
Synovial type A cells	Joints
Fixed tissue macrophages	Spleen, lymph nodes, bonemarrow, and other tissues
Inflammation	
Exudate macrophages	Any tissue
Activated macrophages	Any tissue
Elicited macrophages	Any tissue
Epitheloid cells	Any tissue
Multinucleated giant cells (Langerhans types and foreign body type)	Any tissue

* Adapted from Van Furth R (editor): Mononuclear Phagocytes: Functional Aspects, Martinus Nijhoff, 1980.

As used herein, the term "granulocyte" refers to a polymorphonuclear leukocyte which contains many cytoplasmic granules and a multilobed nucleus. Granulocytes include neutrophils, eosinophils, and neutrophils. Neutrophils are amoeboid in shape, and phagocytic, and stain with both basic and acidic dyes. Eosinophils and basophils contain cytoplasmic granules which stain with acid dyes and with basic dyes, respectively.

"Bone marrow cells" include both primary bone marrow tissue and bone marrow cell lines from mammals; such as primates and humans. Methods of culturing primary bone marrow cells *in vitro* are known, as well as bone marrow cell lines such as IM-9, KG-1 and KG-1a. See, e.g., ATCC CATALOG, *supra*, at 361. Primary bone marrow cells may include, in addition to leukocytes and lymphocytes described elsewhere herein, progenitor cells such as pleuripotent

stem cells. See, e.g., Civin U.S. Pat. No. 4,714,680.

II. Description of the Invention

5 The invention described herein allows the replication of HCV in cell lines wherein the cells are B lymphocytes and T lymphocytes. In a method of the invention, peripheral blood cells which are infected with replicating virions are detected. Preferably, the detection is accomplished by identifying replicative intermediates of the viral genome in preparations containing HCV infected cells. Alternatively, the virions may be detected using immunoassays to detect a population of HCV infected cells. Cell lines wherein the cells are B lymphocytes and T lymphocytes infected with HCV are
10 prepared; these cell lines are usually derived from blood cells or their precursors, more usually are derived from mononuclear cells or their precursors. The preparation of the infected cell line may be accomplished by a variety of methods, including the following. One option is to cultivate susceptible cells with concentrated culture fluids harvested from short term primary cultures of cells which are replicating the virus. Another option is to directly infect susceptible cells with isolated HCV particles. Still another option is to grow susceptible cells in contact with HCV infected cells. The susceptible cells may be from an established cell line, or may be primary cultures. Alternatively, infected cells from an individual
15 with HCV may be fused with established cells from a cell line to produce a permanent cell line. Still alternatively, HCV infected blood cells isolated from an individual may be immortalized. Virions produced by the HCV infected cell lines are useful in immunoassays for the detection of anti-HCV antibodies, and as a substrate for the production of vaccines for treatment of HCV.

20 The practice of the present invention will employ, unless otherwise indicated, conventional techniques of molecular biology, microbiology, hematology, cell and organ culture, recombinant DNA, and immunology, which are within the skill of the art. Such techniques are explained fully in the literature. See, e.g., ATCC CATALOG OF CELL LINES & HYBRIDOMAS, 6th edition, 1988 (American Type Culture Collection, Rockville, Md.); BASIC & CLINICAL IMMUNOLOGY, 6th edition (Stites, Stobo, and Wells, eds., Appleton and Lange, Norwalk Conn/Los Altos, Ca, 1987); Boggs and Winkelstein, WHITE CELL MANUAL, edition 4 (F.A. Davis Co., Phila., 1983); Diggs, Sturm, and Bell (1985), THE MORPHOLOGY OF HUMAN BLOOD CELLS (Abbott Laboratories, Abbott Park, Il.); Freshney (1987), CULTURE OF ANIMAL CELLS: A MANUAL OF BASIC TECHNIQUE (Alan R. Liss, Inc., N.Y.); Maniatis, Fitch & Sambrook, MOLECULAR CLONING; A LABORATORY MANUAL (1982); DNA CLONING, VOLUMES I AND II (D.N Glover ed. 1985); LYMPHOKINES AND INTEFERONS: A PRACTICAL APPROACH (Clemens, Morris, and Gearing, eds., 1987); LYMPHOCYTES: A PRACTICAL APPROACH (Klaus, ed., 1987); VIROLOGY: A PRACTICAL APPROACH (Mahy, ed., 1985); OLIGONUCLEOTIDE SYNTHESIS (M.J. Gait ed, 1984); NUCLEIC ACID HYBRIDIZATION (B.D. Hames & S. J. Higgins eds. 1984); IMMOBILIZED CELLS AND ENZYMES (IRL Press, 1986); B. Perbal, A PRACTICAL GUIDE TO MOLECULAR CLONING (1984); the series, METHODS IN ENZYMOLOGY (Academic Press, Inc.); GENE TRANSFER VECTORS FOR MAMMALIAN CELLS (J.H. Miller and M.P. Calos eds. 1987, Cold Spring Harbor Laboratory),
30 Methods in Enzymology Vol. 154 and Vol. 155 (Wu and Grossman, and Wu, eds., respectively), Mayer and Walker, eds. (1987), IMMUNOCHEMICAL METHODS IN CELL AND MOLECULAR BIOLOGY (Academic Press, London), Scopes, (1987), PROTEIN PURIFICATION: PRINCIPLES AND PRACTICE, Second Edition (Springer-Verlag, N.Y.), and HANDBOOK OF EXPERIMENTAL IMMUNOLOGY, VOLUMES I-IV (D.M. Weir and C. C. Blackwell eds 1986).

40 II.A. Detection and Isolation of HCV Infected Cells from an Individual

HCV infected cells in biological samples are screened for the presence of HCV polypeptides or HCV polynucleotides. Usually the biological samples are blood samples, from which the cells infected with HCV and/or replicating HCV are isolated and partially purified utilizing techniques known in the art, for example, by sedimentation through
45 Ficoll-Paque (Pharmacia Corp.) solutions.

The screening technique used to detect viral material must be sufficiently sensitive to detect very limited amounts. For example, the presence of virions may be detected by immunoassay, using antibodies directed against a virion polypeptide(s). A suitable antibody is described in EP-A-388 232. Methods for immunoassays, as well as methods for signal enhancement are known in the art

50 The HCV genomic material, and replicative intermediates thereof, may also be detected by nucleic acid hybridization. Usually the system will involve amplification techniques in the hybridization assay. Such techniques are known in the art. For example, the Enzo Biochemical Corporation "Bio-Bridge" system uses terminal deoxynucleotide transferase to add unmodified 3'-poly-dT-tails to a DNA probe. The poly dT-tailed probe is hybridized to the target nucleotide sequence, and then to a biotin-modified poly-A. PCT application 84/03520 and EP-A-124221 describe a DNA hybridization assay in which: (1) analyte is annealed to a single-stranded DNA probe that is complementary to an enzyme-labeled oligonucleotide; and (2) the resulting tailed duplex is hybridized to an enzyme-labeled oligonucleotide. EP-A-204510 describes a DNA hybridization assay in which analyte DNA is contacted with a probe that has a tail, such as a poly-dT tail, an amplifier strand that has a sequence that hybridizes to the tail of the probe, such as a poly-A sequence,
55

and which is capable of binding a plurality of labeled strands. A particularly desirable technique may first involve amplification of the target HCV sequences in sera until the amplified sequences are at a detectable level. This may be accomplished, for example, by the polymerase chain reactions (PCR) technique described by Saiki et al. (1986), by Mullis, U.S. Patent No. 4,683,195, and by Mullis et al. U.S. Patent No. 4,683,202. The amplified sequence(s) may then be detected using a hybridization assay. Hybridization assays, which should detect sequences at the level of 10^6 /ml, utilize nucleic acid multimers which bind to single-stranded analyte nucleic acid, and which also bind to a multiplicity of single-stranded labeled oligonucleotides. A suitable solution phase sandwich assay which may be used with labeled polynucleotide probes, and the methods for the preparation of probes is described in EP-A-225 807.

A preferred method for detecting HCV genomic sequences is described in the Examples. The method utilizes primers and probes based upon a family of cDNA replicas of portions of the HCV genome; a composite sequence of the cDNA plus strand derived from this family of cDNAs is shown in Figure 1. The first step in the method is the synthesis of a cDNA to either the HCV genome or its replicative intermediate, using reverse transcriptase. If replicative intermediates are to be detected, the primer is complementary to only the negative strand of HCV RNA (i.e., the strand which is the replicative intermediate). After synthesis of the HCV cDNA, and prior to amplification, the RNA in the sample is degraded by techniques known in the art, for example, by alkali denaturation, or by treatment with an RNA specific RNase.

It is anticipated that variant strains of HCV will be present in the infected cells of various individuals. Therefore the detection methods should be directed to constant epitopes or genomic sequences of HCV.

Utilizing the HCV/cPCR method to screen fractions of blood cells separated on Ficoll-Paque, replicative intermediates of the HCV genome were detected in a partially purified composition of cells derived from peripheral blood (See *infra* in the Examples). This composition is comprised of lymphocytes, monocytes/macrophages, platelets, and small amounts of other white blood cell types. The sedimentation through Ficoll-Paque fractionates these cells, which band to the interface, from erythrocytes and granulocytes, which are of greater density.

Further identification of the infected cell types may be accomplished by utilizing techniques known in the art, for example by immunoassays which are directed to distinguish between the various types of blood cells. Monoclonal antibodies which are directed to surface epitopes on various lymphocytes, and monocytes/macrophages are known in the art. Thus, HCV infected cells may be isolated and identified by immunologic binding to specific cell surface antigens.

II.B. Preparation of *In vitro* Infected Cell Systems

A variety of methods may be used to prepare B lymphocyte or T lymphocyte cell lines, infected with HCV. Candidate cells for infection and propagation *in vitro* can be from any of the above-mentioned cell types, such as lymphocytes or bone marrow cells. The discovery that replicative intermediates of the HCV genome are present in leukocytes in the peripheral blood of infected humans is indicative of the cell type which may support viral replication *in vitro*, i.e., in at least one of the cell types in the population of leukocytes which band at the interface of a leukocyte suspension sedimented through Ficoll-Paque, i.e., mononucleocytes. (This procedure generally removes normal erythrocytes and granulocytes from peripheral blood samples). Further fractionation procedures may be used to separate the different types of mononucleocytes to determine the presence of HCV replicative intermediates and/or HCV antigens associated with these cell types. Procedures for separating different types of monocytes, e.g., various types of lymphocytes, and macrophages, are known in the art. See, e.g., LYMPHOCYTES: A PRACTICAL APPROACH (Klaus, ed.). These procedures may use monoclonal antibodies directed towards specific cell types; the monoclonal antibodies may be affixed to solid substrates, e.g., plates with wells, or magnetic beads. Alternatively, sorting techniques, which rely upon cell sorter instruments may be used. The selection may be either positive or negative to enrich and/or select the desired cells in the population of mononucleocytes. Based upon fractionation studies which isolated lymphocytes and macrophages from the monocyte population, it is suggested that a preferred cell type for replication may be lymphocytes and/or macrophages/monocytes.

Since replication of HCV occurs within mononucleocytes *in vivo*, it is expected that infected cell lines may be derived from known established cell lines of these types, many of which are available from the American Type Culture Collection, and which may include, for example, B cell lines (e.g., EBV transformed B cell derived lines such as B95-8, DAKIKI, LTR228, Mo-B, SC-1, WIL2-NS; B cell lines such as Ia2, BL-3, UC 729-6, Namalwa, U-937, Ramos (RA-1), BCL₁ clone 5B₁b, MC/CAR, IA2, YB2/O, CA3-F4, SHM-D33, CW13.20-3B3, BC9-E%, WM-115, WM 266-4, WEHI-231, WEHI-279, NFS-1.0 C-1); monocyte lines (e.g., THP-1, WEHI-274.1), mouse lymphoma lines (e.g., 2pk-3), pro-B lymphoblasts (e.g., NFS-70 C-10), pre-B lymphoblasts (e.g., NFS-25 C-3), T-cell derived lines (e.g., CTLL-2, HuT 102, D10.G4.1, LBRM-33 clone 4A2, HuT78, Cl.Ly1*2/9, UCD-MLA-144, 6T-CEM 20, 6T-CEM, CCRF-CEM, H9/HTLV-IIIB, CCRF-SB; CCRF-HSB-2, CEM-CM3, Mo, MOLT-3, MOLT-4, BW5147.3, BW5147.G.1.4, C1498, RAW8.1, CTLL-2, and BW5147.G.1.4.OUA^R.1). Additional human leukemic cell lines of interest include K562 and ARH-77. Conditions for the *in vitro* culture of these cells is described in ATCC CATALOG OF CELL LINES AND HYBRIDOMAS, 6th Edition,

1988.

Still another type of cell system in which HCV may be replicated is in immortal cell lines derived from primary cultures of mononucleocytes. Methods to obtain immortalized cell lines of types of mononucleocytes are known in the art. For example, methods for growing and immortalizing B cells and T cells in culture are described in LYMPHOCYTES: A PRACTICAL APPROACH (Klaus, ed.); LYMPHOKINES AND INTERFERONS (Clemens, Morris, and Gearing, eds), and Freshney (1987). For example, B-cell lines are often immortalized by infection with Epstein-Barr virus (EBV). This may be accomplished by incubating mononucleocytes from infected cells with the supernatant from an EBV infected cell line which is a chronic shedder of EBV, for example, ATCC CRL 1612 cells (B95-8).

The growth of B cells and T cells in culture often requires the presence of stimulatory protein factors (also called cytokines) in the culture medium; these factors are usually produced by a cell of another type. A supply of the cytokines may be provided in conditioned medium, i.e., cell culture supernatant from cells which produce the desired factors; or it may also be provided by feeder cells. The requirement may also be met by the provision of the purified cytokines. Cytokines include, for example, the following. Interleukin 1-alpha and beta (also called lymphocyte activating factor, epidermal cell derived thymocyte activating factor, and haemopoietin H1); these lymphokines are derived from multiple cell types, including monocytes, lymphocytes, and keratinocytes. Interleukin 1-alpha and beta have multiple actions, including stimulating interleukin 2 production and haematopoietic activity. Interleukin 2 (also called T-cell growth factor), derived from T-lymphocytes, and which stimulates T and B cell proliferation and differentiation, macrophage activation, and natural killer cell activation. Interleukin 3 (also called Multi-CSF), which is derived from T-lymphocytes, and which has a pluripotent effect on growth and differentiation, and which stimulates mast cell growth. Interleukin 4 (also called B-cell growth factor I), which is derived from T-lymphocytes, and which stimulates T and B cell proliferation and differentiation. Interleukin 5 (also called B-cell growth factor II), which is derived from T-lymphocytes, and which stimulates eosinophil differentiation (and which, in the mouse system, acts as a B-cell growth factor). Low molecular weight B-cell growth factor, which is derived from T-lymphocytes, and which stimulates B-cell proliferation. B-cell stimulatory factor 2 (also called interferon beta-2), which is derived from T-lymphocytes, and which stimulates B cell differentiation. Interferon-gamma (also called immune interferon), which is produced by T-lymphocytes, and which activates macrophages.

Infection of a susceptible cell line with HCV may be accomplished by any one of several techniques which are used in the art to infect cells with virus, particularly with an RNA virus, and more particularly with a Flavivirus. Generally, virus particles adsorb to cultured cells at physiological temperatures. The virus penetrates, uncoats, replicates its nucleic acid in the nucleus or cytoplasm, and after transcription and translation assembles progeny virions which egress either by cell lysis or budding. The number of progeny virus particles released from one cell is referred to as the burst size. Virus is titrated by adsorbing serial tenfold dilutions to cells and counting the resultant plaques after appropriate incubation at the permissive temperature.

Viral infection of susceptible cells may be accomplished by cocultivation of the susceptible cells and HCV infected cells. The HCV infected cells may be from a primary culture. Usually the HCV infected cells and the primary cells are incubated together in approximately equal numbers in an appropriate cell culture medium. The medium may contain Polybrene, or a functionally similar substance, in an appropriate concentration. The incubation is of sufficient time and at the appropriate temperature to allow the newly replicated virus from the HCV infected cells to allow attachment and uptake by the susceptible cells.

The infection of susceptible cells may also be accomplished using virus containing cell-free supernatants of HCV infected cells. Generally the susceptible cells are pretreated with an appropriate concentration of Polybrene (or a functionally similar substance), and then exposed to concentrated culture fluids harvested from short-term primary cultures of HCV infected cells derived from infected individuals.

Susceptible cells may also be infected with virus isolated from HCV containing biological samples. Suitable methods for the isolation of viruses, particularly RNA containing viruses, and more particularly Togaviruses and Flaviviruses are known in the art, for example, see VIROLOGY: A PRACTICAL APPROACH (Mahy, ed.). Also described therein are cells which may be suitable for infection, and growth conditions for these cells.

Another method for preparing HCV infected cells *in vitro* may be by the fusion of HCV infected mononuclear cells with susceptible cells. Methods for fusing somatic cells are known in the art, as are methods for the selection of fused cells. For example, fusion may be catalyzed by the presence of virus, e.g., EBV, Sendai virus, and the like, or by the presence of chemicals, e.g., polyethylene glycol, which induce fusion. The susceptible cell may carry a marker which prevents its growth in the selection media, e.g., the cells may be phenotypically negative for thymidine kinase. Selection is then in media which requires thymidine kinase, e.g., HAT medium. This method is particularly suitable if the HCV infected mononuclear cells isolated from the infected individual do not replicate *in vitro*. A suitable method for the preparation fused cells which may replicate HCV is that described in WO 87/05930, except that the tissue specific cell (reportedly a hepatocyte) which is used to form the trioma is replaced with an HCV infected mononucleocyte.

Yet another method of preparing an HCV infected cell line is by immortalization of infected cells isolated from an HCV infected individual. Generally, mononuclear cells are isolated from peripheral blood. If desired, the mononuclear cell population may be further fractionated. The cells are then grown in primary culture. Immortalization may be ac-

complished by growing and passaging the cells through the cell crisis period, at which point the majority of cultured cells die; the resulting cells are immortal. Alternative methods for producing immortal T-cells and B-cells are described *supra*.

HCV replication in the HCV infected cell lines prepared as described *supra* may be detected by a variety of assays. For example, viral assays may depend upon the cytopathic or transforming characteristics of the virus. Cytopathic viruses may be assayed by their antimetabolic effects in microtitration plates or by the formation of characteristic plaques in monolayers of the appropriate susceptible host cell. A viral suspension is serially diluted and added to monolayer culture plates. The number of plaques forming at the limiting dilution is taken as equivalent to the number of infectious particles in the supernatant medium, allowing the concentration of virus in the initial sample to be calculated. Characterization of the virus may be performed with specific antisera, measuring the cytopathic effect of the virus, by immunoassay, or by the cPCR/HCV assay for HCV RNA, described *supra*.

Transforming viruses may be assayed by, for example, the selective growth of transformed clones in suspension, or by looking for transformation foci in monolayer cultures.

In the event that the virus of interest exhibits neither a detectable cytopathic nor a transforming effect, it may be detected in the infected cell preparation or infected cell lysate by a number of techniques, including, for example, immunoassay using a labeled antibody(s) which is specific for the virus, immunoassay of polypeptides labeled during replication using anti-HCV antibodies, and the detection of viral nucleic acids, for example, an enriched population of negative strands complementary to viral positive strands, or of viral nucleic acids labelled during an *in vitro* viral synthesis period.

II.C. Immunoassays Using Antigens from HCV Infected Cells

The antigens from HCV infected cells which react immunologically with serum containing HCV antibodies are useful in immunoassays to detect presence of HCV antibodies in biological samples. The HCV antigens can take the form of, inter alia, purified or partially purified virus particles or viral polypeptides, disrupted virus, or infected cells or cell lysate therefrom. The production of the various forms of viral antigen from tissue culture sources is well-known in the art. Design of the immunoassays is subject to a great deal of variation, and a variety of these are known in the art. For example, an immunoassay may use a combination of viral epitopes derived from the same or from different viral polypeptides. It may use, for example, one or more viral antigens isolated from the HCV infected cell or from isolated HCV. Alternatively, fixed cells directly from culture may be used if viral epitopes are expressed internally or externally, or alternatively, cellular lysates may be used. It may also use, for example, whole disrupted virions. General methods for preparing disrupted virions are known in the art.

Immunoassay protocols may vary. Protocols may be based, for example, upon competition, or direct reaction, or sandwich type assays. Protocols may also, for example, use solid supports, or may be by immunoprecipitation. Most assays involve the use of labeled antibody or polypeptide; the labels may be, for example, fluorescent, chemiluminescent, radioactive, or dye molecules. Assays which amplify the signals from the probe are also known; examples of which are assays which utilize biotin and avidin, and enzyme-labeled and mediated immunoassays, such as ELISA assays.

A suitable immunoassay may utilize disrupted virions in an ELISA format. The enzyme-linked immunosorbent assay (ELISA) method depends upon conjugation of an enzyme to either an antigen or an antibody, and uses the bound enzyme activity as a quantitative label. To measure antibody, the known antigen is fixed to a solid phase, (e.g., a microplate or plastic cup), incubated with anti-immunoglobulin labeled with enzyme, and washed again. Enzymes suitable for labeling are known in the art, and include, for example, horseradish peroxidase. Enzyme activity bound to the solid phase is measured by adding the specific substrate, and determining product formation or substrate utilization colorimetrically. The enzyme activity bound is a direct function of the amount of antibody bound.

To measure antigen, a known specific antibody is fixed to the solid phase, the test material containing antigen is added, after an incubation the solid phase is washed, and a second enzyme-labeled antibody is added. After washing, substrate is added, and enzyme activity is estimated colorimetrically, and related to antigen concentration.

Thus, for example, to detect anti-HCV antibodies using an ELISA format, isolated HCV, solubilized, e.g., with detergent, may be used to coat appropriate microtiter plates. After the viral antigens are bound, another protein is also used to coat the plates, to prevent nonspecific binding of antibodies. The biological sample to be tested for anti-HCV antibodies is added to the plates, and the formation of HCV antibody-HCV antigen complexes is detected.

Another suitable immunoassay may utilize gel electrophoresis of HCV lysates, followed by Western blotting with the biological sample to be tested for anti-HCV antibodies. The formation of HCV antibody-HCV antigen complexes is detected using techniques known in the art.

Kits suitable for immunodiagnosis and containing the appropriate labeled reagents are constructed by packaging the appropriate materials, including the antigens derived from the HCV infected cells in suitable containers, along with the remaining reagents and materials required for the conduct of the assay, as well as a suitable set of assay instruc-

tions.

II.D. Use of Infectable Cells in Screening for Antiviral Agents

5 The cells described herein which are capable of replicating HCV, may be used to screen for antiviral agents which inhibit HCV replication, and particularly for those agents which preferentially allow cell growth and multiplication while inhibiting viral replication. The screening methods which can be used are those which examine the inhibition of virus replication in the presence of the putative antiviral agent. Generally, the antiviral agents are tested at a variety of concentrations, for their effect on preventing viral replication in a cell culture system which supports viral replication;

10 and then for an inhibition of infectivity or of viral pathogenicity (and a low level of toxicity) in an animal model system. Included within antiviral agents are antibodies which block viral infection (neutralizing antibodies). The ability of anti-HCV antibodies to prevent the infection of the cells *in vitro* can be determined using the assays described herein to determine whether viral replication is inhibited by the presence of the putative anti-HCV antibodies. For example also, virus produced in infected cells may also be used as a source of antigens to determine whether antibodies are

15 HCV specific.

III. Examples

III.A. Detection of HCV RNA in Mononuclear Cells From Individuals with NANBH

20 The detection of HCV RNA in mononuclear cells from blood of individuals with chronic NANBH was accomplished using essentially the cPCR/HCV assay described below.

Blood obtained from 5 individuals with chronic NANBH and which tested positive for anti-HCV antibodies using an HCV-C100 antigen immunoassay, and from 5 control individuals (reportedly free of NANBH, and which was negative

25 in the immunoassay) was examined for the presence of HCV RNA in the mononuclear fraction. A mononucleocyte fraction from the samples was obtained by sedimentation through Ficoll-Paque (Pharmacia), following the manufacturer's directions. The cPCR/HCV assay used in these studies were performed utilizing the following methods for the preparation of RNA, the reverse transcription of the RNA into cDNA, the amplification of specific segments of the cDNA by PCR, and

30 the analysis of the PCR products. Total RNA was isolated from the mononucleocyte fraction. The cDNA used as a template for the PCR reaction was prepared utilizing the designated samples for preparation of the corresponding cDNAs. Each RNA sample (corresponding to cells in 10 microliters of blood) was incubated in a 25 microliter reaction containing 1 micromolar of each primer, 1 millimolar of each deoxyribonucleotide triphosphate (dNTP), 50 millimolar Tris-HCL, pH 8.3, 5 millimolar

35 MgCl₂, 5 millimolar dithiothreitol (DTT), 73 millimolar KCl, 40 units of RNase inhibitor (RNASIN), and 5 units of AMV reverse transcriptase. The incubation was for 60 minutes at 37°C. Following cDNA synthesis, the reactions were diluted with 50 microliters of deionized water (DIW), boiled for 10 minutes, and cooled on ice. Amplification of a segment of the HCV cDNA was performed utilizing two synthetic 16-mer oligonucleotide primers whose sequences were derived from HCV cDNA clones 36 (anti-sense) and 37b (sense). The sequence of the primer

40 from clone 36 was:

5' GCA TGT CAT GAT GTA T 3'.

45

The sequence of the primer from clone 37b was:

5' ACA ATA CGT GTG TCA C 3'.

50

The primers were used at a final concentration of 1 micromolar each. In order to amplify the segment of HCV cDNA which is flanked by the primers, the cDNA samples were incubated with 0.1 microgram of RNase A and the PCR reactants of the Perkin Elmer Cetus PCR kit (N801-0043 or N801-0055) according to the manufacturer's instructions.

55 The PCR reaction was performed for either 30 cycles or 60 cycles in a Perkin Elmer Cetus DNA thermal cycler. Each cycle consisted of a 1 minute denaturation step at 94°C, an annealing step of 2 minutes at 37°C, and an extension step of 3 minutes at 72°C. However, the extension step in the final cycle (30 or 60) was 7 minutes rather than 3 minutes. After amplification the samples were extracted with an equal volume of phenol: chloroform (1:1), followed by extraction

with an equal volume of chloroform, and then the samples were precipitated with ethanol containing 0.2 M sodium acetate.

The cPCR products were analyzed as follows: The products were subjected to electrophoresis on 1.8% alkaline agarose gels according to Murakawa et al. (1988), and transferred onto Zeta Probe paper (BioRad Corp.) by blotting gels overnight in 0.4 M NaOH. The blots were neutralized in 2 × SSC (1 × SSC contains 0.15 M NaCl, 0.015 M sodium citrate), prehybridized in 0.3 M NaCl, 15 mM sodium phosphate buffer, pH 6.8, 15 mM EDTA, 1.0% SDS, 0.5% nonfat milk (Camation Co.), and 0.5 mg/ml sonicated denatured salmon sperm DNA. The blots to be analyzed for HCV cDNA fragments were hybridized to a ³²P-labeled probe generated by nick translation of the HCV cDNA insert sequence in clone 35, shown in Figure 2. After hybridization, the blots were washed in 0.1 × SSC (1 × SSC contains 0.15M NaCl, 0.01M Na citrate) at 65°C, dried, and autoradiographed. The expected product size is 586 nucleotides in length; products which hybridized with the probe and migrated in the gels in this size range were scored as positive for viral RNA.

As a control, cPCR primers designed to amplify α1-antitrypsin mRNA was performed to verify the presence of RNA in each sample analyzed. The coding region of the α1-antitrypsin gene is described in Rosenberg et al. (1984). Synthetic 16-mer oligonucleotide primers designed to amplify a 365 nucleotide fragment of the coding region of the α1-antitrypsin gene were derived from nucleotides 22-37 (sense) and nucleotides 372-387 (anti-sense). The PCR products were detected using a ³²P nick-translated probe which lies between, and not including, the cDNA/PCR primer sequences.

Due to the extreme sensitivity of the PCR reaction, all samples were run a minimum of three times. All false positive signals were eliminated when the following precautions were taken: 1) eliminating aerosols by using screw capped tubes with rubber O-ring seals; 2) pipetting with Rainin Microman positive displacement pipetters with disposable pistons/capillaries; and 3) selecting the oligonucleotide sequences for the cDNA and PCR primers from two non-contiguous cDNA clones.

The results showed that mononucleocytes from 5 of 5 HCV anti-C100 antibody positive samples were positive for HCV RNA in the cPCR HCV assay. All of the controls were negative in the same assay.

III.B. Detection of HCV RNA Minus Strands in Mononucleocytes from HCV Infected Individuals

The detection of minus strands in peripheral blood mononucleocytes was accomplished using the cPCR/HCV assay essentially as described in Section III.A., except that the cDNA was synthesized using either the 16mer primer from clone 36 (i.e., primer 36/16A), described in Section III.A. to prime the plus strand of HCV RNA, or the complement of the 16-mer (i.e., primer 36/16B), to prime the negative strand. In order to carry out the subsequent PCR reaction, the cDNA reaction mixture was boiled and treated with RNase prior to the addition of the second primer.

The results of the cPCR/HCV assay showed that 5 of the HCV anti-C100 antibody samples were positive for HCV negative strands; of these five samples, two showed high levels of HCV RNA negative strands. All 5 of these samples were about equally positive for the plus strands of HCV RNA. Of the five control samples, all were negative for both plus and minus strands of HCV RNA.

As an additional control, plasma samples from three of the HCV positive individuals were also analyzed. The signal obtained from the positive strands were at least 5 to 10 times stronger than was the signal for the negative strands. Thus, the plasma samples were strongly positive for the plus strands, and only weakly positive to negative for the minus strands.

In HCV, which is Flavi-like, the viral genome is positive stranded RNA. The discovery of the great enrichment of negative strands of HCV RNA in monocytes relative to that in plasma, suggests that replicative intermediates of the viral genome are present in these cells.

IV. Commercial Utility

HCV infected cell systems, which are infected and/or replicated *in vitro*, including HCV infected cell lines, serve as a source of viral antigens which are useful in the detection of HCV antibodies, and hence in the diagnosis and the screening of biological materials (including blood) for HCV. These antigens also may have use in the preparation of vaccines for the treatment (prophylactic and/or therapeutic) of viral infections.

In addition, these cell systems are useful in the screening of anti-viral agents directed at preventing HCV replication, including antibodies which can neutralize infection or mediate antibody-dependent cellular cytotoxicity (ADCC).

While the invention has been described above by way of illustration, the present invention is not limited thereto and its scope is to be defined by the appended claims and the equivalents thereof.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- 5 (i) APPLICANT:
- (A) NAME: Chiron Corporation
 - (B) STREET: 4560 Horton Street
 - (C) CITY: Emeryville
 - 10 (D) STATE: California
 - (E) COUNTRY: USA
 - (F) POSTAL CODE (ZIP): 94608
- (ii) TITLE OF INVENTION: Methods for Culturing HCV in Eukaryotic Cells
- 15 (iii) NUMBER OF SEQUENCES: 5
- (v) CURRENT APPLICATION DATA:
- APPLICATION NUMBER: EP 90309120.5

(2) INFORMATION FOR SEQ ID NO:1:

- 20 (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 16 base pairs
 - (B) TYPE: nucleic acid
 - 25 (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

30

GCATGTCATG ATGTAT

16

(2) INFORMATION FOR SEQ ID NO:2:

- 35 (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 16 base pairs
 - (B) TYPE: nucleic acid
 - 40 (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

45

ACAATAOGTG TGTCAC

16

50 (2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 9185 base pairs
 - 55 (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Hepatitis C virus

(B) STRAIN: HCV1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

10	CACTOCACCA TGAATCACTC CCCTGTGAGG AACTACTGTC TTCAOGCAGA AAGOGTCTAG	60
	CCATGGOGTT AGTATGAGTG TOGTGCAGCC TCCAGGACCC CCOCTCCOOG GAGAGCCATA	120
15	GTGGTCTGOG GAACCGGTGA GTACACOGGA ATTGCCAGGA OGACOGGGTC CTTTCTTGGA	180
	TCAACCOGCT CAATGCOCTGG AGATTGGGGC GTGCCCCOOG AAGACTGCTA GCOGAGTAGT	240
	GTGGGGTGOC GAAAGGCOCT GTGGTACTGC CTGATAGGGT GCTTGOGAGT GOCOCGGGAG	300
20	GTCTOGTAGA COGTGCACCA TGAGCACGAA TCCTAAACCT CAAAAAAAAA ACAAAOGTAA	360
	CACCAACOGT OGCCACAGG AOGTCAAGTT CCOGGGTGGC GGTCAGATOG TTGGTGGAGT	420
25	TTACTTGTTG COGOGCAGGG GCOCTAGATT GGGTGTGOGC GOGACGAGAA AGACTTCOGA	480
	GOGGTGCGAA OCTOGAGGTA GAOGTCAGCC TATCCCCAAG GCTOGTOGGC COGAGGGCAG	540
	GAOCTGGGCT CAGCCOOGGT ACOCTTGGCC OCTCTATGGC AATGAGGGCT GOGGGTGGGC	600
30	GGGATGGCTC CTGTCTOCC GTGGCTCTOG GCOCTAGCTGG GGCCCCACAG ACCOCCGGOG	660
	TAGGTGOGC AATTGGGTA AGGTCATOGA TACCOCTAOG TGOGGCTTOG COGACCTCAT	720

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	GGGTACATA OOGCTOGTOG GOGOOOCTCT TGGAGGOGCT GOCAGGGOOD TGGOGCATGG	780
5	OGTOGGGTT CTGGAAGAOG GOGTGAACFA TGCAACAGGG AACCTTCCTG GTTGCTCTTT	840
	CTCTATCTTC CTTCTGGGOC TGCTCTCTTG CTTGACTGTG OCOGCTTCGG OCTACCAAGT	900
	GOGCAACTOC AOGGGGCTTT ACCAOGTCAC CAATGATTGC OCTAACTOGA GTATTGTGTA	960
10	OGAGGCGGOC GATGOCATOC TGCACACTOC GGGGTGOGTC OCTTGOGTTC GTGAGGGCAA	1020
	OGCTOGAGG TGTGGGGTGG OGATGAOOOC TAOGGTGGOC ACCAGGGATG GCAAACTOOC	1080
15	OGOGACGAG CTTGAAGTC ACATOGATCT GCITGTGGG AGOGOCACOC TCTGTTGGC	1140
	OCTCTAOGTG GGGGAOCTAT GGGGTCTGT CTTCTTGTG GGCCAACTGT TCACCTTCTC	1200
	TOOCAGGOGC CACTGGACGA OGCAAGGTIG CAATIGCTCT ATCTATCCOG GOCATATAAC	1260
20	GGGTACOGC ATGGCATGGG ATATGATGAT GAACTGGTCC OCTAOGACGG OGTTGGTAAT	1320
	GGCTCAGCTG CTOGGATOC CACAAGOCAT CTTGGACATG ATOGCTGGTG CTCACTGGG	1380
25	AGTCTGGOG GGCATAGOGT ATTTCTOCAT GGTGGGAAC TGGGOGAAGG TOCTGGTAGT	1440
	GCTGCTGCTA TTTGCOGGOG TOGACOGGA AAOCCAGTC ACOGGGGGAA GTGCOGGCCA	1500
	CACTGTGTCT GGATTTGTTA GOCTCCTOGC ACCAGGOGOC AAGCAGAACG TOCAGCTGAT	1560
30	CAACACCAAC GGCAGTTGGC AOCTCAATAG CAOGGOCTG AACTGCAATG ATAGOCTCAA	1620
	CACOGGCTGG TTGGCAGGGC TTTTCTATCA CCACAAGTTC AACTCTTCAG GCTGTCTGA	1680
35	GAGGCTAGOC AGCTGCOGAC COCTTACGA TTTTGACCAG GGCTGGGGOC CTATCAGTTA	1740
	TGOCACOGGA AGOGGGOOG ACCAGOGOC CTACTGCTGG CACTACCCOC CAAAACCTTG	1800
40	OGGTATTGTG OCOGOGAAGA GTGTGTGTGG TCOGGTATAT TGCTTCACTC CCAGOOOOGT	1860
	GGTGGTGGGA AOGACOGACA GGTGGGGOC GOCCACCTAC AGCTGGGGTG AAAATGATAC	1920
	GGAGTCTTC GTOCTTAACA ATAOCAGGOC ACOGCTGGGC AATTGGTTTG GTTGTAOCTG	1980
45	GATGAACFA ACTGGATTCA CCAAAGTGTG OGGAGOGCT OCTTGTGTCA TOGGAGGGGC	2040
	GGGCAACAAC AOCCTGCACT GGOOACTGA TTGCTTCOGC AAGCATCOGG AOGCCACATA	2100
50	CTCTOGGTG GGTGCOGGT OCTGGATCAC AOCAGGTGC CTGGTOGACT ACOGTATAG	2160
	GCITTGGCAT TATCCTGTGA CCATCAACTA CACCATATTT AAAATCAGGA TGTACGTGGG	2220

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	AGGGGTGAA CACAGGCTGG AAGCTGCTG CAACTGGAOG OGGGGOGAAC GTTGOGATCT	2280
	GGAGACAGG GACAGGTOG AGCTCAGOC GTTACTGCTG ACCACTACAC AGTGGCAGGT	2340
5	OCTOCGTGT TCCTTCACAA CCTACOCAGC CTGTGCCACC GGCTCATCC ACCTOCACCA	2400
	GAACATTGTG GACGTGCAGT ACTTGTAOGG GGTGGGGTCA AGCATOGOGT OCTGGGOCAT	2460
10	TAAGTGGGAG TACGTGTFC TCCTGTTCCT TCTGCTTGCA GAGGOGOGG TCTGCTOCTG	2520
	CTGTGGATG ATGCTACTCA TATCCCAAGC GGAGGOGGCT TTGGAGAACC TGTAAATACT	2580
15	TAATGCAGCA TCCGTGGGCG GGAOGCAOGG TCTGTATCC TTCTGTGTGTT TCTTCTGCTT	2640
	TGCATGGTAT TTGAAGGGTA AGTGGGTGCC CGGAGGGTC TACACCTTCT ACGGGATGTG	2700
	GGCTCTOCTC CTGCTOCTGT TGGGTFGGC CCAGGGGGG TAOGGCTGG ACAOGGAGGT	2760
20	GGGOGGTGG TGTGGGGTG TTGTCTOCT CGGGTTGATG GGGCTGACTC TGTCACCATA	2820
	TTACAAGGC TATATCAGCT GGTGCTGTG GTGGCTTCAG TATTTTCTGA CCAGAGTGA	2880
25	AGGCAACTG CAOGTGTGA TTCCCCCT CAACTGGA GGGGGGGG ACGGCTCAT	2940
	CTTACTCATG TGTGCTGTAC ACGGCTCT GGTATTTGAC ATCAACAAAT TGCTGCTGGC	3000
	GGTCTTGGG CCGCTTTGGA TTCTTCAAGC CAGTTTGCTT AAAGTACCT ACTTTGTGG	3060
30	GGTCAAGGC CTTCTOOGT TCTGOGGTT AGGOGGAAG ATGATOGGAG GOCATTAGT	3120
	GCAAATGGT ATCATTAAGT TAGGGGGCT TACTGGCACC TATGTTTATA ACCATCTCAC	3180
35	TCCTCTTGG GACTGGGGC ACAAGGCTT GCGAGATCTG GCGTGGCTG TAGAGOCAGT	3240
	GGTCTTCTC CAAATGGAGA CCAAGCTCAT CAGTGGGGG GCAGATACG CCGGTGGG	3300
	TGACATCATC AAGGCTTGC CTGTTTGGC CGCAGGGGC OGGGAGATC TGCTOGGGC	3360
40	AGGOGATGA ATGGTCTCA AGGGGTGGG GTTGTGGG OCCATCAOG GTAGGOGCA	3420
	GCAGACAAG GGCTCTTAG GGTGCATAAT CACAGGCTA ACTGGGGG AAAAAACCA	3480
45	AGTGGAGGT GAGGTCCAGA TTGTGTCAAC TGCTGCCAA ACCTTCTGG CAAGTGCAT	3540
	CAATGGGGT TGCTGGACTG TCTACCAOG GCGGGAAG AGGACCATG GTTACCCAA	3600
50	GGGTCTGTC ATCAGATGT ATACCAATGT AGACCAAGC CTGTGGGGT GCGGCTOC	3660
	GCAAGGTAGC GGCTCATGA CACCTGCAC TTGGGGCTC TGGACCTTT ACCTGGTCAC	3720

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GAGGCAAGCC GATGTCAATC OCGTGOGGCG GGGGGGTGAT AGCAGGGGCA GCGTGCTGTC 3780
 GCGCGGGGCG ATTTCTACT TGAAAGGCTC CTGGGGGGT CCGCTGTTGT GCGCGGGG 3840
 5 GCAAGCGGTG GGCATATTTA GGGCGGGGT GTGCAACCGT GGAGTGGCTA AGGCGGTGGA 3900
 CTTTATCOCT GTGGAGAACC TAGAGACAAC CATGAGGTCC CCGGTGTTCA CGGATAACTC 3960
 10 CTCTCCACCA GTAGTGGGCG AGAGCTTCCA GGTGGCTCAC CTGCATGCTC CCACAGGCAG 4020
 CGGCAAAAGC ACCAAGGTCC CGGCTGCATA TGCAGCTCAG GGCTATAAGG TGCTAGTACT 4080
 CAACCCCTCT GTTGCTGCAA CACTGGGCTT TGGTGCTTAC ATGTCCAAGG CTCATGGGAT 4140
 15 CGATCCTAAC ATCAGGACCG GGGTGAGAAC AATTACCACT GGCAGCCCCA TCACGTACTC 4200
 CAOCTAAGGC AAGTTCTTG CGAAGGGG GTGCTGGGG GGGCTTATG ACATAATAAT 4260
 20 TTGTGAAGAG TGCCACTCCA CGGATGCCAC ATCCATCTTG GGCATGGCA CTGTCTTGA 4320
 CCAAGCAGAG ACTGGGGGG CGAGACTGGT TGTGCTGCG ACCGCCACC CTCGGGGCTC 4380
 CGTCACTGTG CCCCATCCA ACATGAGGA GGTGCTCTG TCAACACCG GAGAGATCC 4440
 25 TTTTAAAGC AAGGCTATCC CCTCGAAGT AATCAAGGG GGGAGACATC TCATCTTCTG 4500
 TCATTCAAAG AAGAAGTGG AGGAAGTGC CGCAAAGCTG GTGCATTGG GCATCAATGC 4560
 30 CGTGGCTAC TACGGGGTCT TTGAAGTGT CGTCACTCCG ACCAGGGGG ATGTTGTGCT 4620
 CGTGGCAACC GATGCGCTCA TGACGGGCTA TACGGGGAC TTGACTGG TGATAGACTG 4680
 35 CAATAAGTGT GTCAACCGA CAGTGAATT CAGCTTGAC CTTACCTTCA CCATTGAGAC 4740
 AATCAAGCTC CCGAGGATG CTGTCTCCG CACTCAAAGT CGGGCAGGA CTGGCAGGG 4800
 GAAGCCAGGC ATCTACAGAT TTGTGGCAC GGGGAGGCG CCTCCGGCA TGTTCGACTC 4860
 40 GTCGTCTC TGTGAGTGT ATGAAGCAG CTGTGCTTG TATGAGCTCA CGCGCGCGA 4920
 GACTACAGTT AGGCTAAGAG GTTACATGAA CACCGGGG CTTCCGTTG GCGAGGACA 4980
 45 TCTTGAATTT TGGGAGGGG TCTTTACAGG CTTCACTCAT ATAGATGCC ACTTTCTATC 5040
 CCAGACAAAG CAGAGTGGG AGAACCTTC TTACCTGGTA GGTACCAAG CCACGTTGT 5100
 CGCTAGGGCT CAAGCCCTC CCGATGCTG GGACAGATG TGAAGTGT TGATTGCGCT 5160
 50 CAAGCCACC CTGCATGGG CAACCCCT GCTATACAGA CTGGGGCTG TTCAGAATGA 5220

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	AATCAOCTG ACGCAOCCAG TCACCAAATA CATCATGACA TGCATGTGG COGAOCTGGA	5280
	GGTGTCAAG AGCAOCTGGG TGCTGTGTGG OGGGTGCTG GCTGCTTTGG COGGTATTG	5340
5	OCTGTCAACA GGCTGGGTGG TCATAGTGGG CAGGGTGTG TTGTGCGGA AGCOGGCAAT	5400
	CATACCTGAC AGGGAAGTCC TCTACGAGA GTTGTATGAG ATGGAAGAGT GCTCTCAGCA	5460
10	CTTACCGTAC ATCGAGCAAG GGATGATGCT GCGGAGCAG TTCAAGCAGA AGGCCCTGGG	5520
	OCTOCTGAG ACOGGTCCC GTCAGGCAGA GGTATOGOC OCTGCTGTCC AGACCAACTG	5580
15	GCAAAACTC GAGACCTTCT GGGGAAGCA TATGTGGAAC TTCATCAGTG GGATACAATA	5640
	CTTGGGGGC TTGTCAACGC TGCTGTGTAA CCGGCGCATT GCTTCATTGA TGGCTTTTAC	5700
	AGCTGCTGTC ACCAGCCCAC TAACCACTAG CCAAAOCTC CTCTTCAACA TATTGGGGGG	5760
20	GTGGGTGGCT GCGCAGCTGG CCGCCCCGGG TGCGCTTACT GCGTTTGTGG GCGCTGGCTT	5820
	AGCTGGGGCC GGCATGGCA GTGTGGACT GGGGAAGGTC CTCATAGACA TCCTTGCAGG	5880
25	GTATGGGGG GGGTGGGGG GAGCTCTTGT GGCATTCAAG ATCATGAGGG GTGAGGTCCC	5940
	CTCCAGGAG GACCTGGTCA ATCTACTGCC CGCATCTC TOGCGGGAG CCGCTGTAGT	6000
	CGGGTGGTC TGTGAGCAA TACTGCGCG GCACTTGGC CCGGGGAGG GGGCAGTGA	6060
30	GTGGATGAAC CGGCTGATAG CCTTGGCTC CCGGGGAAC CATGTTTCC CCAOCCACTA	6120
	CGTGOOGGAG AGGATGCAG CTGCGCGGT CACTGCCATA CTCAGCAGCC TCACTGTAA	6180
35	CCAGCTOCTG AGGCGACTGC ACCAGTGGAT AAGCTGGAG TGTACCACTC CATGCTCGG	6240
	TTCTGGCTA AGGGACATCT GGGACTGGAT ATGCGAGGTG TTGAGGACT TTAAGAOCTG	6300
	GCTAAAAGCT AAGCTCATGC CACAGCTGCC TGGGATCCC TTTGTGTCT GCGAGCGGG	6360
40	GTATAAGGGG GTCTGGGAG TGGAGGCGAT CATGCACACT CGCTGCCACT GTGGAGCTGA	6420
	GATCACTGGA CATGTCAAAA ACGGAGCGAT GAGGATGTC GGTCTAGGA CCTGCAGGAA	6480
45	CATGTGGAGT GGGACCTTCC CCATTAAATGC CTACACCAAG GCGOCTGTA CCGGCTTCC	6540
	TGCGCGAAC TACAGTTGG OGCTATGGAG GGTGTCTGCA GAGGAATATG TGGAGATAAG	6600
50	GCAGGTGGGG GACTTCCACT ACGTGAOGGG TATGACTACT GACAATCTCA AATGCCCGTG	6660
	CCAGGTCCA TOGCGGAAT TTTTCACAGA ATTGGAOGGG GTGCGCTAC ATAGGTTTGC	6720
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	GCCCCCTGC AAGCCCTTGC TGOGGGAGGA GGTATCATTG AGAGTAGGAC TOCACGAATA	6780
	CCCGGTAGGG TOGCAATTAC CTTGOGAGCC OGAACGGAC GTGGCOGTGT TGACGTCCAT	6840
5	GCTCACTGAT CCTCCCATTA TAACAGCAGA GGOGGCOGGG OGAAGGTITG OGAGGGGATC	6900
	ACCCCCCTCT GTGGCCAGCT CTOGGGCTAG CCAGCTATCC GCTCCATCTC TCAAGGCAAC	6960
10	TTGCACCGCT AACCATGACT CCCCTGATGC TGAGCTCATA GAGGCCAACC TOCTATGGAG	7020
	GCAGGAGATG GGCGGCAACA TCACCAGGGT TGAGTCAGAA AACAAAGTGG TGATTCTGGA	7080
	CTCCTTOGAT CCGCTTGTGG OGGAGGAGGA OGAGOGGGAG ATCTCOGTAC CCGCAGAAAT	7140
15	CCTGOGGAAG TCTOGGAGAT TOGCCCAGGC CCTGCCCGTT TGGGOGGGC OGGACTATAA	7200
	CCCCCGCTA GTGGAGAOGT GGAAAAAGCC CGACTACGAA CCACTGTGG TCCATGGCTG	7260
20	TCOGCTTCCA CCTCCAAAGT CCCCTCCTGT GCCTCOGCT OGAAGAAGC GGAOGGTGGT	7320
	CCTCACTGAA TCAACCTAT CTA CTACTGCTT GGCGAGCTC GCCACCAGAA GCTTTGGCAG	7380
	CTCTCAACT TOGGCATT A OGGGOGACAA TACGACAACA TCCTCTGAGC CCGCCCCCTC	7440
25	TGGCTGCCCC CCGACTCCG AGCTGAGTC CTATTCTCC ATGCCCCCCC TGGAGGGGGA	7500
	GCTTGGGGAT CCGATCTTA GCGAOGGTC ATGGTCAAAG GTCAGTAGTG AGGCCAAAGC	7560
30	GGAGGATGTC GTGTGCTGCT CAATGTCTTA CTCTGGACA GGCGCACTOG TCACCCCGTG	7620
	CGCGOGGAA GAACAGAAAC TGCCCATCAA TGCACTAAGC AACTCGTTGC TACGTCACCA	7680
35	CAATTTGGTG TATTCCACCA CCTCAGCAG TGCTTGCCAA AGGCAGAAGA AAGTCACATT	7740
	TGACAGACTG CAAGTTCTGG ACAGOCATTA CCAGGAGTA CTCAAGGAGG TTAAAGCAGC	7800
	GGGTCAAAA GTGAAGGCTA ACTTGCTATC CGTAGAGGAA GCTTGCAGCC TGACGCCCCC	7860
40	AACTCAGCC AAATCCAAGT TTGGTTATGG GGCAAAAGAC GTCOGTTGCC ATGCCAGAAA	7920
	GGOGTAAACC CACATCAACT CCGTGTGGAA AGACCTTCTG GAAGACAATG TAACACCAAT	7980
45	AGACACTACC ATCATGGCTA AGAAGAGGT TTTCTGGTT CAGCCTGAGA AGGGGGGTGG	8040
	TAAGCCAGCT CGTCTCATOG TGTTCOOGA TCTGGGOGTG CGGTGTGOG AAAAGATGGC	8100
50	TTTGTACGAC GTGGTTACAA AGCTCCCTT GCGGTGATG GGAAGCTCCT ACGGATTCCA	8160
	ATACTCACCA GGACAGOGG TTGAATTCT CTTGCAAGOG TGGAAGTCCA AGAAAACCCC	8220

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	AATGGGGTTC TOGTATGATA CCOGCTGCTT TGACTIONACA GTCACITGAGA GOGACATCOG	8280
	TACGGAGGAG GCAATCTACC AATGTTGTGA OCTOGACCCC CAAGCCCGOG TGGCCATCAA	8340
5	GTCCCTCACC GAGAGGCTTT ATGTTGGGGG COCTCTTACC AATTCAAGGG GGGAGAACTG	8400
	OGGCTATOGC AGGTGCOGOG OGAGOGGOGT ACTGACAACT AGCTGTGGTA ACACCOCTCAC	8460
10	TTGCTACATC AAGGCOOGGG CAGCCTGTG AGCOGCAGGG CTCCAGGACT GCACCATGCT	8520
	CGTGTGTGGC GAOGACTTAG TOGTTATCTG TGAAAGOGOG GGGGTCCAGG AGGACOGGGC	8580
	GAGCCTGAGA GCCTTCAOGG AGGCTATGAC CAGGTACTCC GCCCCCCTG GGGACCCCCC	8640
15	ACAACCAGAA TAGGACTTGG AGCTCATAAC ATCATGCTCC TCCAACGTGT CAGTOGCOCA	8700
	OGACGGOGCT GGAAAGAGGG TCTACTAOCT CACCOGTGAC OCTACAAACC COCTOGOGAG	8760
20	AGCTGOGTGG GAGACAGCAA GACACACTCC AGTCAATTCC TGGCTAGGCA ACATAATCAT	8820
	GTTTGGCCCC ACACTGTGGG OGAGGATGAT ACTGATGACC CATTTCTTTA GGTCCCTTAT	8880
25	AGCCAGGGAC CAGCTTGAAC AGGCCCTCGA TTGOGAGATC TAOGGGGCT GCTACTCCAT	8940
	AGAACCACTT GATCTACCTC CAATCATTCA AAGACTCCAT GGCTCAGOG CATTTTCACT	9000
	CCACAGTTAC TCTCCAGGTG AAATTAAATAG GGTGGCOGCA TGCTCAGAA AACTTGGGGT	9060
30	ACCGCCCTTG OGAGCTTGA GACACOGGGC OGGGAGGTC OGOGCTAGGC TTCTGGCCAG	9120
	AGGAGGCAGG GCTGCCATAT GTGGCAAGTA OCTCTTCAAC TGGGCAGTAA GAACAAAGCT	9180
35	CAAAC	9185

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 480 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Hepatitis C Virus
- (B) STRAIN: HCV1

(vii) IMMEDIATE SOURCE:

- (B) CLONE: DNA 35

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..480

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

5	TOC ATT GAG ACA ATC ACG CTC CCC CAG GAT GCT GTC TCC OGC ACT CAA	48
	Ser Ile Glu Thr Ile Thr Leu Pro Gln Asp Ala Val Ser Arg Thr Gln	
	1 5 10 15	
10	CGT CGG GGC AGG ACT GGC AGG GGG AAG CCA GGC ATC TAC AGA TTT GTG	96
	Arg Arg Gly Arg Thr Gly Arg Gly Lys Pro Gly Ile Tyr Arg Phe Val	
	20 25 30	
15	GCA OCG GGG GAG OGC CCC TCC GGC ATG TTC GAC TCG TCC GTC CTC TGT	144
	Ala Pro Gly Glu Arg Pro Ser Gly Met Phe Asp Ser Ser Val Leu Cys	
	35 40 45	
20	GAG TGC TAT GAC GCA GGC TGT GCT TGG TAT GAG CTC ACG CCC GCC GAG	192
	Glu Cys Tyr Asp Ala Gly Cys Ala Trp Tyr Glu Leu Thr Pro Ala Glu	
	50 55 60	
25	ACT ACA GTT AGG CTA CGA GCG TAC ATG AAC ACC OCG GGG CTT CCC GTG	240
	Thr Thr Val Arg Leu Arg Ala Tyr Met Asn Thr Pro Gly Leu Pro Val	
	65 70 75 80	
30	TGC CAG GAC CAT CTT GAA TTT TGG GAG GGC GTC TTT ACA GGC CTC ACT	288
	Cys Gln Asp His Leu Glu Phe Trp Glu Gly Val Phe Thr Gly Leu Thr	
	85 90 95	
35	CAT ATA GAT GCC CAC TTT CTA TCC CAG ACA AAG CAG AGT GGG GAG AAC	336
	His Ile Asp Ala His Phe Leu Ser Gln Thr Lys Gln Ser Gly Glu Asn	
	100 105 110	
40	CTT OCT TAC CTG GTA GCG TAC CAA GCC ACC GTG TGC GCT AGG GCT CAA	384
	Leu Pro Tyr Leu Val Ala Tyr Gln Ala Thr Val Cys Ala Arg Ala Gln	
	115 120 125	
45	GCC OCT CCC CCA TCG TGG GAC CAG ATG TGG AAG TGT TTG ATT OGC CTC	432
	Ala Pro Pro Pro Ser Trp Asp Gln Met Trp Lys Cys Leu Ile Arg Leu	
	130 135 140	
50	AAG CCC ACC CTC CAT GGG CCA ACA CCC CTG CTA TAC AGA CTG GGC GCT	480
	Lys Pro Thr Leu His Gly Pro Thr Pro Leu Leu Tyr Arg Leu Gly Ala	
	145 150 155 160	

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 160 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

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Ser Ile Glu Thr Ile Thr Leu Pro Gln Asp Ala Val Ser Arg Thr Gln
 1 5 10 15
 5 Arg Arg Gly Arg Thr Gly Arg Gly Lys Pro Gly Ile Tyr Arg Phe Val
 20 25 30
 Ala Pro Gly Glu Arg Pro Ser Gly Met Phe Asp Ser Ser Val Leu Cys
 35 40 45
 10 Glu Cys Tyr Asp Ala Gly Cys Ala Trp Tyr Glu Leu Thr Pro Ala Glu
 50 55 60
 15 Thr Thr Val Arg Leu Arg Ala Tyr Met Asn Thr Pro Gly Leu Pro Val
 65 70 75 80
 Cys Gln Asp His Leu Glu Phe Trp Glu Gly Val Phe Thr Gly Leu Thr
 85 90 95
 20 His Ile Asp Ala His Phe Leu Ser Gln Thr Lys Gln Ser Gly Glu Asn
 100 105 110
 Leu Pro Tyr Leu Val Ala Tyr Gln Ala Thr Val Cys Ala Arg Ala Gln
 115 120 125
 25 Ala Pro Pro Pro Ser Trp Asp Gln Met Trp Lys Cys Leu Ile Arg Leu
 130 135 140
 30 Lys Pro Thr Leu His Gly Pro Thr Pro Leu Leu Tyr Arg Leu Gly Ala
 145 150 155 160

35 Claims

Claims for the following Contracting States : AT, BE, CH, LI, DE, DK, FR, GB, IT, LU, NL, SE

- 40 1. A cell line infected with hepatitis C virus (HCV), wherein the cells in the cell line are B lymphocytes or T lymphocytes and are capable of replicating HCV.
2. The cell line of claim 1 wherein the cells in the cell line are B lymphocytes.
- 45 3. The cell line of claim 1 wherein the cells in the cell line are T lymphocytes.
4. The cell line of claim 3, wherein the cells are derived by infection of a cell from the cell line MOLT-4.
- 50 5. A method of preparing a composition for the detection of an anti-HCV antibody which comprises: purifying or partially purifying HCV virus particles or polypeptides from a cell line according to any one of claim 1 to 4; and fixing said particles or polypeptides to a solid phase.
6. A method according to claim 5 wherein HCV particles are solubilized with a detergent.
- 55 7. A method according to claim 5 or 6 wherein the composition for the detection of anti-HCV antibodies is packaged in a suitable container to provide a test kit.
8. A method for screening for an agent which inhibits HCV replication comprising:

- (a) providing a composition comprised of a putative anti-viral agent;
- (b) providing a cell line capable of replicating HCV;
- (c) providing a composition comprised of infective HCV;
- (d) incubating the compositions of (a) and (c) with the cell line of (b) under conditions which would, in the absence of (a) allow infection of and replication of HCV in the cell line; and
- (e) detecting an inhibition of viral replication after the incubating in (d), wherein the cells in the cell line of (b) are B lymphocytes or T lymphocytes.

Claims for the following Contracting States : ES, GR

1. A method of preparing a composition for the detection of an anti-HCV antibody which comprises: purifying or partially purifying HCV virus particles or polypeptides from a cell line infected with hepatitis C virus (HCV), wherein the cells in the cell line are B lymphocytes or T lymphocytes and are capable of replicating HCV; and fixing said particles or polypeptides to a solid phase.
2. The method of claim 1 wherein the cells in the cell line are B lymphocytes.
3. The method of claim 1 wherein the cells in the cell line are T lymphocytes.
4. The method of claim 3, wherein the cells are derived by infection of a cell from the cell line MOLT-4.
5. A method according to any one of claims 1 to 4 wherein HCV particles are solubilized with a detergent.
6. A method according to any one of claims 1 to 5 wherein the composition for the detection of anti-HCV antibodies is packaged in a suitable container to provide a test kit.
7. A method for screening for an agent which inhibits HCV replication comprising:
 - (a) providing a composition comprised of a putative anti-viral agent;
 - (b) providing a cell line capable of replicating HCV;
 - (c) providing a composition comprised of infective HCV;
 - (d) incubating the compositions of (a) and (c) with the cell line of (b) under conditions which would, in the absence of (a) allow infection of and replication of HCV in the cell line; and
 - (e) detecting an inhibition of viral replication after the incubating in (d), wherein the cells in the cell line of (b) are B lymphocytes or T lymphocytes.
8. A cell line infected with hepatitis C virus (HCV), wherein the cells in the cell line are B lymphocytes or T lymphocytes and are capable of replicating HCV.
9. The cell line of claim 8 wherein the cells in the cell line are B lymphocytes.
10. The cell line of claim 8 wherein the cells in the cell line are T lymphocytes.
11. The cell line of claim 10, wherein the cells are derived by infection of a cell from the cell line MOLT-4.

Patentansprüche

Patentansprüche für folgende Vertragsstaaten : AT, BE, CH, LI, DE, DK, FR, GB, IT, LU, NL, SE

1. Zelllinie, infiziert mit Hepatitis C-Virus (HCV), dadurch gekennzeichnet, daß die Zellen der Zelllinie B-Lymphozyten oder T-Lymphozyten sind und in der Lage sind, HCV zu replizieren.
2. Zelllinie nach Anspruch 1, dadurch gekennzeichnet, daß die Zellen in der Zelllinie B-Lymphozyten sind.
3. Zelllinie nach Anspruch 1, dadurch gekennzeichnet, daß die Zellen in der Zelllinie T-Lymphozyten sind.

4. Zelllinie nach Anspruch 3, dadurch **gekennzeichnet**, daß die Zellen durch Infektion einer Zelle von der Zelllinie MOLT-4 abgeleitet sind.
5. Verfahren zur Herstellung einer Zusammensetzung zum Nachweis eines Anti-HCV-Antikörpers, umfassend: Reinigen oder teilweises Reinigen von HCV-Viruspartikeln oder Polypeptiden aus eine Zelllinie nach einem der Ansprüche 1 bis 4; und Fixieren der Partikel oder Polypeptide an eine feste Phase.
6. Verfahren nach Anspruch 5, dadurch **gekennzeichnet**, daß die HCV-Partikel in einem Detergens solubilisiert werden.
7. Verfahren nach Anspruch 5 oder 6, dadurch **gekennzeichnet**, daß die Zusammensetzung zum Nachweis von Anti-HCV-Antikörpern in einem geeigneten Behälter verpackt ist, um einen Testkit bereitzustellen.
8. Verfahren zum Durchmustern eines Agens, welches die HCV-Replikation inhibiert, umfassend:
 - (a) Bereitstellung einer Zusammensetzung, umfassend ein mögliches antivirales Agens;
 - (b) Bereitstellung einer Zelllinie, die in der Lage ist, HCV zu replizieren;
 - (c) Bereitstellung einer Zusammensetzung, welche infektiöses HCV umfaßt;
 - (d) Inkubation der Zusammensetzungen (a) und (c) mit der Zelllinie (b) unter Bedingungen, die in Abwesenheit von (a) die Infektion der Zelllinie und die Replikation von HCV in der Zelllinie ermöglichen würden; und
 - (e) Nachweis einer Inhibition der viralen Replikation nach Inkubation in (d), wobei die Zellen in der Zelllinie (b) B-Lymphozyten oder T-Lymphozyten sind.

Patentansprüche für folgende Vertragsstaaten : ES, GR

1. Verfahren zur Herstellung einer Zusammensetzung zum Nachweis eines Anti-HCV-Antikörpers, umfassend: Reinigen oder teilweises Reinigen von HCV-Viruspartikeln oder Polypeptiden aus einer Zelllinie, die mit Hepatitis C-Virus (HCV) infiziert ist, dadurch **gekennzeichnet**, daß die Zellen in der Zelllinie B-Lymphozyten oder T-Lymphozyten sind und in der Lage sind, HCV zu replizieren; und Fixieren der Partikel oder Polypeptide an einer festen Phase.
2. Verfahren nach Anspruch 1, dadurch **gekennzeichnet**, daß die Zellen in der Zelllinie B-Lymphozyten sind.
3. Verfahren nach Anspruch 1, dadurch **gekennzeichnet**, daß die Zellen in der Zelllinie T-Lymphozyten sind.
4. Verfahren nach Anspruch 3, dadurch **gekennzeichnet**, daß die Zellen durch Infektion einer Zelle aus der Zelllinie MOLT-4 abgeleitet sind.
5. Verfahren nach einem der Ansprüche 1 bis 4, dadurch **gekennzeichnet**, daß die HCV-Partikel mit einem Detergens solubilisiert werden.
6. Verfahren nach einem der Ansprüche 1 bis 5, dadurch **gekennzeichnet**, daß die Zusammensetzung zum Nachweis von Anti-HCV-Antikörpern in einem geeigneten Behältnis verpackt ist, um einen Testkit bereitzustellen.
7. Verfahren zum Durchmustern eines Agens, welches die HCV-Replikation inhibiert, umfassend:
 - (a) Bereitstellung einer Zusammensetzung, umfassend ein mögliches antivirales Agens;
 - (b) Bereitstellung einer Zelllinie, die in der Lage ist, HCV zu replizieren;
 - (c) Bereitstellung einer Zusammensetzung, welche infektiöses HCV umfaßt;
 - (d) Inkubation der Zusammensetzungen (a) und (c) mit der Zelllinie (b) unter Bedingungen, die in Abwesenheit von (a) die Infektion der Zelllinie und die Replikation von HCV in der Zelllinie ermöglichen würden; und
 - (e) Nachweis einer Inhibition der viralen Replikation nach Inkubation in (d), wobei die Zellen in der Zelllinie (b) B-Lymphozyten oder T-Lymphozyten sind.
8. Zelllinie, infiziert mit Hepatitis C-Virus (HCV), dadurch **gekennzeichnet**, daß die Zellen in der Zelllinie B-Lymphozyten oder T-Lymphozyten sind und in der Lage sind, HCV zu replizieren.

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9. Zellinie nach Anspruch 8, dadurch **gekennzeichnet**, daß die Zellen in der Zellinie B-Lymphozyten sind.
10. Zellinie nach Anspruch 8, dadurch **gekennzeichnet**, daß die Zellen in der Zellinie T-Lymphozyten sind.
- 5 11. Zellinie nach Anspruch 10, dadurch **gekennzeichnet**, daß die Zellen durch Infektion einer Zelle aus der Zellinie MOLT-4 abgeleitet sind.

Revendications

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Revendications pour les Etats contractants suivants : AT, BE, CH, LI, DE, DK, FR, GB, IT, LU, NL, SE

- 15 1. Lignée cellulaire infectée avec le virus de l'hépatite C (HCV), dans laquelle les cellules dans la lignée cellulaire sont des lymphocytes B ou des lymphocytes T et sont capables de répliquer HCV.
2. Lignée cellulaire de la revendication 1 dans laquelle les cellules dans la lignée cellulaire sont des lymphocytes B.
- 20 3. Lignée cellulaire de la revendication 1 dans laquelle les cellules dans la lignée cellulaire sont des lymphocytes T.
4. Lignée cellulaire de la revendication 3 dans laquelle les cellules sont dérivées par infection d'une cellule provenant de la lignée cellulaire MOLT-4.
- 25 5. Méthode de préparation d'une composition pour la détection d'un anticorps anti-HCV qui comprend les étapes consistant : à purifier ou à purifier de façon partielle des particules de virus HCV ou des polypeptides provenant d'une lignée cellulaire selon l'une quelconque des revendications 1 à 4; et à fixer lesdites particules ou lesdits polypeptides sur une phase solide.
- 30 6. Méthode selon la revendication 5 dans laquelle les particules d'HCV sont solubilisées avec un détergent.
7. Méthode selon la revendication 5 ou 6 dans laquelle la composition pour la détection d'anticorps anti-HCV est conditionnée dans un récipient approprié pour fournir un kit de test.
- 35 8. Méthode pour cribler un agent qui inhibe la réplication d'HCV comprenant les étapes consistant :
- (a) à fournir une composition comprenant un agent anti-viral putatif ;
- (b) à fournir une lignée cellulaire capable de répliquer HCV ;
- (c) à fournir une composition comprenant du HCV infectieux ;
- 40 (d) à faire incuber les compositions de (a) et (c) avec la lignée cellulaire de (b) dans des conditions qui, en l'absence de (a), permettraient l'infection et la réplication de HCV dans la lignée cellulaire ; et
- (e) à détecter une inhibition de la réplication virale après l'incubation dans (d), dans laquelle les cellules dans la lignée cellulaire de (b) sont des lymphocytes B ou des lymphocytes T.

45 **Revendications pour les Etats contractants suivants : ES, GR**

1. Méthode de préparation d'une composition pour la détection d'un anticorps anti-HCV qui comprend les étapes consistant : à purifier ou à purifier de façon partielle des particules de virus HCV ou des polypeptides provenant d'une lignée cellulaire infectée avec le virus de l'hépatite C (HCV), dans laquelle les cellules dans la lignée cellulaire
50 sont des lymphocytes B ou des lymphocytes T et sont capables de réplication d'HCV; et à fixer lesdites particules ou polypeptides sur une phase solide.
2. Méthode de la revendication 1 dans laquelle les cellules dans la lignée cellulaire sont des lymphocytes B.
- 55 3. Méthode de la revendication 1 dans laquelle les cellules dans la lignée cellulaire sont des lymphocytes T.
4. Méthode de la revendication 3, dans laquelle les cellules sont dérivées par infection d'une cellule provenant de la lignée cellulaire MOLT-4.

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5. Méthode selon l'une quelconque des revendications 1 à 4 dans laquelle les particules d'HCV sont solubilisées avec un détergent.
- 5 6. Méthode selon l'une quelconque des revendications 1 à 5 dans laquelle la composition pour la détection d'anticorps anti-HCV est conditionnée dans un récipient approprié pour fournir un kit de test.
7. Méthode pour l'âge d'un agent qui inhibe la réplication d'HCV comprenant les étapes consistant :
- 10 (a) à fournir une composition contenant un agent anti-viral putatif;
(b) à fournir une lignée cellulaire capable de réplication HCV;
(c) à fournir une composition comprenant HCV infectieux;
(d) à incuber les compositions de (a) et (c) avec la lignée cellulaire (b) dans des conditions qui permettraient, en l'absence de (a) de permettre l'infection de et la réplication de HCV dans la lignée cellulaire; et
15 (e) à détecter une inhibition de la réplication virale après l'incubation dans (d), dans laquelle les cellules dans la lignée cellulaire (b) sont des lymphocytes B ou des lymphocytes T.
8. Lignée cellulaire infectée avec le virus de l'hépatite C (HCV), dans laquelle les cellules dans la lignée cellulaire sont des lymphocytes B ou des lymphocytes T et sont capables de réplication d'HCV.
- 20 9. Lignée cellulaire de la revendication 8 dans laquelle les cellules dans la lignée cellulaire sont des lymphocytes B.
10. Lignée cellulaire de la revendication 8 dans laquelle les cellules dans la lignée cellulaire sont des lymphocytes T.
- 25 11. Lignée cellulaire de la revendication 10 dans laquelle les cellules sont dérivées par infection d'une cellule provenant de la lignée cellulaire MOLT-4.

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SEQUENCE OF THE HCV cDNA SENSE STRAND

(AS DEDUCED FROM OVERLAPPING CLONES b114a/18g/
ag30a/CA205a/CA290a/CA216a/pi14a/CA167b/CA156e/
CA84a/CA59a/K9-1/26j/13i/12f/14i/11b/7f/7e/8h/
33c/40b/37b/35/36/81/32/33b/25c/14c/8f/33f/33g/
39c/35f/19g/26g/15e/b5a/16jh)

CACTCCACCATGAATCACTCCCCTGTGAGGAACTACTGTCTTCACGCAGAA
AGCGTCTAGCCATGGCGTTAGTATGAGTGTCTGTCAGCCTCCAGGACCCCC
CCTCCCGGGAGAGCCATAGTGGTCTGCGGAACCGGTGAGTACACCGGAATT
GCCAGGACGACCGGTCCTTTCTTGATCAACCCGCTCAATGCCTGGAGAT
TTGGGCGTGCCCCCGCAAGACTGCTAGCCGAGTAGTGTGGGTGCGGAAAG
GCCTTGTGGTACTGCCTGATAGGGTGCTTGCAGAGTCCCCGGGAG-300

(Putative initiator methionine codon)

GTCTCGTAGACCGTGACCATGAGCACGAATCCTAAACCTCAAAAAAAAAA
CAAACGTAACACCAACCGTCGCCCACAGGACGTCAAGTTCCCGGGTGGCGG
TCAGATCGTTGGTGGAGTTTACTTGTGCGCGCAGGGGCCCTAGATTGGG
TGTGCGCGGACGAGAAAGACTTCCGAGCGGTGCGAACCTCGAGGTAGACG
TCAGCCTATCCCCAAGGCTCGTCGGCCCCGAGGGCAGGACCTGGGCTCAGCC
CGGGTACCCTTGGCCCCCTCTATGGCAATGAGGGCTGCGGGTGGGC-600
GGGATGGCTCCTGTCTCCCCGTGGCTCTCGGCCTAGCTGGGGCCCCACAGA
CCCCCGGCGTAGGTGCGCGCAATTTGGGTAAAGTTCATCGATACCCTTACGTG
CGGCTTCGCCGACCTCATGGGGTACATACCGCTCGTCGGCGCCCCCTCTTGG
AGGCGCTGCCAGGGCCCTGGCGCATGGCGTCCGGGTTCTGGAAGACGGCGT
GAACTATGCAACAGGGAACCTTCTGCTTGCTCTTTCTCTATCTTCTCTCT
GGCCCTGCTCTCTTGCTTGACTGTGCCCCGCTTCGGCCTACCAAGT-900
GCGCAACTCCACGGGGCTTACCACGTACCAATGATTGCCCTAACTCGAG
TATTGTGTACGAGGCGGCCGATGCCATCCTGCACACTCCGGGGTGGCTCCC
TTGCGTTCTGTGAGGGCAACGCCTCGAGGTGTTGGGTGGCGATGACCCCTAC
GGTGGCCACCAGGGATGGCAAACTCCCCGCGACGCAGCTTCGACGTCACAT
CGATCTGCTTGTGCGGAGCGCCACCCTCTGTTCCGGCCCTCTACGTGGGGGA
CCTATGCGGGTCTGTCTTTCTTGTGCGGCAACTGTTACCTTCTC-1200
TCCCAGGCGCCACTGGACGACGCAAGGTTGCAATTGCTCTATCTATCCCGG
CCATATAACGGGTCAACGCATGGCATGGGATATGATGATGAACTGGTCCCC
TACGACGGCGTTGGTAATGGCTCAGCTGCTCCGGATCCCACAAGCCATCTT
GGACATGATCGCTGGTGTCTACTGGGGAGTCTGGCGGGCATAGCGTATTT
CTCCATGGTGGGGAACCTGGGCGAAGGTCCTGGTAGTGCTGCTGCTATTTGC
CGGCGTCGACGCGGAAACCCACGTCACCGGGGGAAGTGCCGGCCA-1500
CACTGTGTCTGGATTTGTTAGCCTCCTCGCACCAGGCGCCAAGCAGAACGT
CCAGCTGATCAACACCAACGGCAGTTGGCACCTCAATAGCACGGCCCTGAA
CTGCAATGATAGCCTCAACACCGGCTGGTTGGCAGGGCTTTTCTATACCA
CAAGTTCAACTCTCAGGCTGTCTGAGAGGCTAGCCAGCTGCCGACCCCT
TACCGATTTTGACCAGGGCTGGGGCCCTATCAGTTATGCCAACGGAAGCGG
CCCCGACCAGCGCCCTACTGCTGGCACTACCCCCCAAACCTTG-1800

FIG. 1-1

CGGTATTGTGCCCCGGAAGAGTGTGTGTGGTCCGGTATATTGCTTCACTCC
 CAGCCCCGTGGTGGTGGGAACGACCGACAGGTCCGGCGCGCCACCTACAG
 CTGGGGTGAAAATGATACGGACGTCTTCGTCTTAACAATACCAGGCCACC
 GCTGGGCAATTGGTTCGGTTGTACCTGGATGAACTCAACTGGATTACCAA
 AGTGTGCGGAGCGCCTCCTTGTGTTCATCGGAGGGGCGGGCAACAACACCCT
 GCACTGCCCCACTGATTGCTTCCGCAAGCATCCGGACGCCACATA-2100
 CTCTCGGTGCGGCTCCGGTCCCTGGATCACACCCAGGTGCCTGGTGCAGTA
 CCCGTATAGGCTTTGGCATTATCCTTGTACCATCAACTACACCATATTTAA
 AATCAGGATGTACGTGGGAGGGGTCGAACACAGGCTGGAAGCTGCCTGCAA
 CTGGACGCGGGGCGAACGTTGCGATCTGGAAGACAGGGACAGGTCCGAGCT
 CAGCCCGTTACTGCTGACCACTACACAGTGGCAGGTCCCTCCCGTGTTCCTT
 CACAACCCTACCAGCCTTGTCCACCGCCTCATCCACCTCCACCA-2400
 GAACATTGTGGACGTGCAGTACTTGTACGGGGTGGGGTCAAGCATCGCGTC
 CTGGGCCATTAAGTGGGAGTACGTCTTCTCCTGTTCTTCTGCTTGCAGA
 CGCGCGCTCTGCTCCTGCTTGTGGATGATGCTACTCATATCCCAAGCGGA
 GGCGGCTTTGGAGAACCTCGTAATACTTAATGCAGCATCCCTGGCCGGGAC
 GCACGGTCTTGTATCCTTCCTCGTGTCTTCTGCTTTGCATGGTATTTGAA
 GGGTAAGTGGGTGCCCCGAGCGGTCTACACCTTCTACGGGATGTG-2700
 GCCTCTCCTCCTGCTCCTGTTGGCGTTGCCCCAGCGGGCGTACGCGCTGGA
 CACGGAGGTGGCCGCGTCTGTGGCGGTGTTGTTCTCGTCCGGTTGATGGC
 GCTGACTCTGTACCATATTACAAGCGCTATATCAGCTGGTGTGTTGGTG
 GCTTCAGTATTTTCTGACCAGAGTGAAGCGCAACTGCACGTGTGGATTCC
 CCCCCTCAACGTCCGAGGGGGGCGCGACGCCGTCATCTTACTCATGTGTGC
 TGTACCCCGACTCTGGTATTTGACATCACCAAATTGCTGCTGGC-3000
 CGTCTTCGGACCCCTTTGGATTCTTCAAGCCAGTTTGCTTAAAGTACCCTA
 CTTTGTGCGGTCCAAGGCCTTCTCCGGTTCTGCGCGTTAGCGCGGAAGAT
 GATCGGAGGCCATTACGTGCAAATGGTTCATCATTAAGTTAGGGCGCTTAC
 TGGCACCTATGTTTATAACCATCTCACTCCTCTTCGGGACTGGGCGCAAA
 CGGCTTGGAGATCTGGCCGTGGCTGTAGAGCCAGTCGTCTTCTCCCAAAT
 GGAGACCAAGCTCATCACGTGGGGGGCAGATACCGCCGCGTGC GG-3300
 TGACATCATCAACGGCTTGCCTGTTTCCGCCCGCAGGGGCGGGAGATACT
 GCTCGGGCCAGCCGATGGAATGGTCTCCAAGGGGTGGAGGTTGCTGGCGCC
 CATCACGGCGTACGCCCGAGCAGACAAGGGGCCTCCTAGGGTGCATAATCAC
 CAGCCTAACTGGCCGGGACAAAAACCAAGTGGAGGGTGAGGTCCAGATTGT
 GTCAACTGCTGCCCAAACCTTCTGGCAACGTGCATCAATGGGGTGTGCTG
 GACTGTCTACCACGGGGCCGGAACGAGGACCATCGCGTCACCCAA-3600
 GGGTCCTGTTCATCCAGATGTATACCAATGTAGACCAAGACCTTGTGGGCTG
 GCGCGCTCCGCAAGGTAGCCGCTCATTGACACCCTGCACTTGGCGCTCCTC
 GGACCTTTACCTGGTCACGAGGCACGCCGATGTCAATCCCGTGGCGCGGGC
 GGGTGATAGCAGGGGCAGCCTGCTGTGCCCCGGGCCATTTCTACTTGAA
 AGGCTCCTCGGGGGTCCGCTGTTGTGCCCCGGGGGCACGCCGTGGGCAT
 ATTTAGGGCCGCGGTGTGCAACCGTGGAGTGGCTAAGGCGGTGGA-3900
 CTTTATCCCTGTGGAGAACCTAGAGACAACCATGAGGTCCCCGGTGTTCAC
 GGATAACTCCTCTCCACCAGTAGTGCCCCAGAGCTTCAGGTGGCTCACCT
 CCATGCTCCACAGGCAGCGGCAAAAGCACCAAGGTCCCGGTGCATATGC
 AGCTCAGGGCTATAAGGTGCTAGTACTCAACCCCTCTGTTGCTGCAACACT
 GGGCTTTGGTGCTTACATGTCCAAGGCTCATGGGATCGATCCTAACATCAG
 GACCGGGGTGAGAACAATTACCACTGGCAGCCCCATCACGTACTC-4200

FIG. 1-2

CACCTACGGCAAGTTCCTTGCCGACGGCGGGTGCTCGGGGGGCGCTTATGA
CATAATAATTTGTGACGAGTGCCACTCCACGGATGCCACATCCATCTTGGG
CATCGGCACTGTCCTTGACCAAGCAGAGACTGCGGGGGCGAGACTGGTTGT
GCTCGCCACCGCCACCCCTCCGGGCTCCGTCACTGTGCCCCATCCCAACAT
CGAGGAGGTTGCTCTGTCCACCACCGGAGAGATCCCTTTTTACGGCAAGGC
TATCCCCCTCGAAGTAATCAAGGGGGGGAGACATCTCATCTTCTG-4500
TCATTCAAAGAAGAAGTGCGACGAACTCGCCGCAAAGCTGGTCGCATTGGG
CATCAATGCCGTGGCCTACTACCGCGGTCTTGACGTGTCCGTCAATCCCGAC
CAGCGGCATGTTGTGTCGTGGCAACCGATGCCCTCATGACCGGCTATAC
CGGCGACTTCGACTCGGTGATAGACTGCAATACGTGTGTCAACCAGACAGT
CGATTTTCAGCCTTGACCCTACCTTACCATTGAGACAATCACGCTCCCCCA
GGATGCTGTCTCCGCACTCAACGTGCGGGCAGGACTGGCAGGGG-4800
GAAGCCAGGCATCTACAGATTTGTGGCACCAGGGGAGCGCCCTCCGGCAT
GTTTCGACTCGTCCGTCTCTGTGAGTGCTATGACGCAGGCTGTGCTTGGA
TGAGCTCACGCCCCGCGAGACTACAGTTAGGCTACGAGCGTACGTAACAC
CCCGGGGCTTCCCGTGTGCCAGGACCATCTTGAATTTTGGGAGGGCGTCTT
TACAGGCCTCACTCATATAGATGCCCACTTTCTATCCAGACAAAGCAGAG
TGGGGAGAACCCTTCTTACCTGGTAGCGTACCAAGCCACCGTGTG-5100
CGCTAGGGCTCAAGCCCCCTCCCCCATCGTGGGACCAGATGTGGAAGTGT
GATTCGCCTCAAGCCCACCCTCCATGGGCCAACACCCCTGCTATACAGACT
GGGCGCTGTTTCAAGATGAAATCACCTGACGCACCCAGTCACCAAATACAT
CATGACATGCATGTGCGCCGACCTGGAGGTCGTACGAGCACCTGGGTGCT
CGTTGGCGGGCTCCTGGCTGCTTTGGCCGCGTATTGCCTGTCAACAGGCTG
CGTGGTCATAGTGGGCAGGGTCGTCTTGTCCGGGAAGCCGGCAAT-5400
CATACCTGACAGGGAAGTCCTTACCGAGAGTTCGATGAGATGGAAGAGTG
CTCTCAGCACTTACCGTACATCGAGCAAGGGATGATGCTCGCCGAGCAGTT
CAAGCAGAAGGCCCTCGGCCTCCTGCAGACCGCGTCCCGTCAGGCAGAGGT
TATCGCCCCCTGCTGTCCAGACCAACTGGCAAAAACCTCGAGACCTTCTGGGC
GAAGCATATGTGGAACCTTCATCAGTGGGATACAATACTTGGCGGGCTTGT
AACGCTGCCTGGTAACCCCGCCATTGCTTCATTGATGGCTTTTAC-5700
AGCTGCTGTCCAGCCCCACTAACCACTAGCCAAACCCCTCCTTCAACAT
ATTGGGGGGGTGGGTGGCTGCCAGCTCGCCGCCCCCGGTGCCGCTACTGC
CTTTGTGGGCGCTGGCTTAGCTGGCGCGCCATCGGCAGTGTGGACTGGG
GAAGGTCCTCATAGACATCCTTGCAGGGTATGGCGCGGGCGTGGCGGGAGC
TCTTGTGGCATTCAAGATCATGAGCGGTGAGGTCCCCTCCACGGAGGACCT
GGTCAATCTACTGCCCGCCATCCTCTCGCCCGGAGCCCTCGTAGT-6000
CGGCGTGGTCTGTGCAGCAATACTGCGCCGGCACGTTGGCCCGGGCGAGGG
GGCAGTGCAGTGGATGAACCGGCTGATAGCCTTCGCCTCCCGGGGGAACCA
TGTTTTCCCCCAGCACTACGTGCCGGAGAGCGATGCAGCTGCCCGCGTCAC
TGCCATACTCAGCAGCCTCACTGTAACCCAGCTCCTGAGGCGACTGCACCA
GTGGATAAGCTCGGAGTGTACCACTCCATGCTCCGGTTCCTGGCTAAGGGA
CATCTGGGACTGGATATGCGAGGTGTTGAGCGACTTTAAGACCTG-6300
GCTAAAAGCTAAGCTCATGCCACAGCTGCCTGGGATCCCCCTTTGTGTCCTG
CCAGCGCGGGTATAAGGGGGTCTGCCGAGTGGACGGCATCATGCACACTCG
CTGCCACTGTGGAGCTGAGATCACTGGACATGTCAAAAACGGGACGATGAG
GATCGTCCGTCTAGGACCTGCAGGAACATGTGGAGTGGGACCTTCCCCAT
TAATGCCTACACCACGGGGCCCCTGTACCCCCCTTCTGCGCCGAACACAC
GTTCGCGCTATGGAGGGTGTCTGCAGAGGAATATGTGGAGATAAG-6600

FIG. 1-3

GCAGGTGGGGGACTTCCACTACGTGACGGGTATGACTACTGACAATCTCAA
 ATGCCCCGTGCCAGGTCCCATCGCCCCGAATTTTTACAGAATTGGACGGGGT
 GCGCCTACATAGGTTTGCGCCCCCTGCAAGCCCTTGCTGCGGGAGGAGGT
 ATCATTAGAGTAGGACTCCACGAATACCCGGTAGGGTTCGCAATTACCTTG
 CGAGCCCGAACCAGGACGTGGCCGTGTTGACGTCCATGCTCACTGATCCCTC
 CCATATAACAGCAGAGGCGGCCGGGCGAAGGTTGGCGAGGGGATC-6900
 ACCCCCCTCTGTGGCCAGCTCCTCGGCTAGCCAGCTATCCGCTCCATCTCT
 CAAGGCAACTTGCACCGCTAACCATGACTCCCCTGATGCTGAGCTCATAGA
 GGCCAACCTCCTATGGAGGCAGGAGATGGGCGGCAACATCACCAGGGTTGA
 GTCAGAAAACAAAGTGGTGATTCTGGACTCCTTCGATCCGCTTGTGGCGGA
 GGAGGACGAGCGGGAGATCTCCGTACCCGCAGAAATCCTGCGGAAGTCTCG
 GAGATTCGCCCAGGCCCTGCCCCGTTTGGGCGCGGCCGGAATAA-7200
 CCCCCCGTAGTGGAGACGTGGAAAAAGCCCGACTACGAACCACCTGTGGT
 CCATGGCTGTCCGCTTCCACCTCCAAAGTCCCCTCCTGTGCCTCCGCTCG
 GAAGAAGCGGACGGTGGTCTCACTGAATCAACCCTATCTACTGCCTTGGC
 CGAGCTCGCCACCAGAAGCTTTGGCAGCTCCTCAACTTCCGGCATTACGGG
 CGACAATACGACAACATCCTCTGAGCCCGCCCCCTTCTGGCTGCCCCCCCCGA
 CTCGACGCTGAGTCCTATTCTCCATGCCCCCCTGGAGGGGGA-7500
 GCCTGGGGATCCGGATCTTAGCGACGGGTCAATGGTCAACGGTCAGTAGTGA
 GGCCAACGCGGAGGATGTCGTGTGCTGCTCAATGTCTTACTCTTGGACAGG
 CGCACTCGTCACCCCGTGCGCCGCGGAAGAACAGAACTGCCCATCAATGC
 ACTAAGCAACTCGTTGCTACGTCACCACAATTTGGTGTATTCCACCACCTC
 ACGCAGTGCTTGCCAAAGGCAGAAGAAAGTCACATTTGACAGACTGCAAGT
 TCTGGACAGCCATTACCAGGACGTACTCAAGGAGGTTAAAGCAGC-7800
 GCGCTCAAAAGTGAAGGCTAACTTGCTATCCGTAGAGGAAGCTTGCAGCCT
 GACGCCCCCAGCTCAGCCAAATCCAAGTTTGGTTATGGGGGAAAAGACGT
 CCGTTGCCATGCCAGAAAGGCCGTAACCCACATCAACTCCGTGTGGAAAGA
 CCTTCTGGAAGACAATGTAACACCAATAGACACTACCATCATGGCTAAGAA
 CGAGGTTTTCTGCGTTGAGCCTGAGAAGGGGGGTGTAAGCCAGCTCGTCT
 CATCGTGTTCCCGATCTGGGCGTGCGCGTGTGCGAAAAGATGGC-8100
 TTTGTACGACGTGGTTACAAAGCTCCCCTTGGCCGTGATGGGAAGCTCCTA
 CGGATTCCAATACTCACCAGGACAGCGGGTTGAATTCCTCGTGCAAGCGTG
 GAAGTCCAAGAAAACCCCAATGGGGTTCTCGTATGATACCCGCTGCTTTGA
 CTCCACAGTCACTGAGAGCGACATCCGTACGGAGGAGGCAATCTACCAATG
 TTGTGACCTCGACCCCCAAGCCCGCTGGCCATCAAGTCCCTCACCAGAG
 GCTTTATGTTGGGGGCCCTCTTACCAATTCAAGGGGGGAGAAGTG-8400
 CGGCTATCGCAGGTGCCGCGGAGCGGCGTACTGACAACTAGCTGTGGTAA
 CACCCTCACTTGCTACATCAAGGCCCGGGCAGCCTGTCGAGCCGAGGGCT
 CCAGGACTGCACCATGCTCGTGTGTGGCGACGACTTAGTCGTTATCTGTGA
 AAGCGCGGGGGTCCAGGAGGACGCGGCGAGCCTGAGAGCCTTACGGAGGC
 TATGACCAGGTACTCCGCCCCCCTGGGGACCCCCACAACCAGAATACGA
 CTTGGAGCTCATAACATCATGCTCCTCCAACGTGTCAGTCGCCCA-8700
 CGACGGCGCTGGAAAGAGGGTCTACTACCTCACCCTGACCCCTACAACCCC
 CCTCGGAGAGCTGCGTGGGAGACAGCAAGACACACTCCAGTCAATTCCTG
 GCTAGGCAACATAATCATGTTTGCCCCACACTGTGGGCGAGGATGATACT
 GATGACCCATTTCTTTAGCGTCCTTATAGCCAGGGACCAGCTTGAACAGGC
 CCTCGATTGCGAGATCTACGGGGCCTGCTACTCCATAGAACCACCTTGATCT
 ACCTCCAATCATTCAAAGACTCCATGGCCTCAGCGCATTTTCACT-9000

FIG. 1-4

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CCACAGTTACTCTCCAGGTGAAATTAATAGGGTGGCCGCATGCCTCAGAAA
ACTTGGGGTACCGCCCTTGCGAGCTTGGAGACACCGGGCCCGGAGCGTCCG
CGCTAGGCTTCTGGCCAGAGGAGGCAGGGCTGCCATATGTGGCAAGTACCT
CTTCAACTGGGCAGTAAGAACAAGCTCAAAC

FIG. 1-5

FIG. 2 Translation of DNA 35

SerIleGluThrIleThrLeuProGlnAspAlaValSerArgThrGlnArgArgGlyArg
 1 TCCATTGAGACAATCACGCTCCCCAGGATGCTGTCTCCGCACTCAACGTCCGGGCAGG
 AGGTAACTCTGTAGTGGAGGGGTCTCTACGACAGAGGGCGTGAGTTGCAGCCCCCGTCC
 ThrGlyArgGlyLysProGlyIleTyrArgPheValAlaProGlyGluArgProSerGly
 61 ACTGGCAGGGGAAGCCAGGCATCTACAGATTGTGGCACCGGGGAGCGCCCTCCCGC
 TGACCGTCCCCCTTCGGTCCGTAGATGTCTAAACACCCGTGGCCCCCTCGCGGGAGGCCG
 MetPheAspSerSerValLeuCysGluCysTyrAspAlaGlyCysAlaTrpTyrGluLeu
 121 ATGTTGACTCGTCCGTCTGTGAGTGCTATGACGCAGGCTGTGCTTGGTATAGCTC
 TACAAGCTGAGCAGGCAGGACACTCACGATACTCGTCCGACACGAACCATACTCGAG
 ThrProAlaGluThrThrValArgLeuArgAlaTyrMetAsnThrProGlyLeuProVal
 181 ACGCCCGCGAGACTACAGTTAGGCTACGAGCGTACATGAACACCCCGGGCTTCCCCGTG
 TCGGGCGGCTCTGATGTCAATCCGATGCTCGCATGTACTTGTGGGCCCCCGAAGGCAC
 CysGlnAspHisLeuGluPheTrpGluGlyValPheThrGlyLeuThrHisIleAspAla
 241 TGCCAGGACCATCTTGAATTTGGAGGGCGTCTTTACAGGCCCTCACTCATATAGATGCC
 ACGGTCCTGGTAGAACTTAAACCCCTCCCGCAGAAATGTCCGGAGTGAGTATATCTACGG
 HisPheLeuSerGlnThrLysGlnSerGlyGluAsnLeuProTyrLeuValAlaTyrGln
 301 CACTTCTATCCAGACAAGCAGAGTGGGGAGAACCTTCCTTACCTGGTAGCGTACCAA
 GTGAAAGATAGGGTCTGTTTCGTCCTCACCCCTCTTGGAGGAATGGACCATCGCATGGTT
 AlaThrValCysAlaArgAlaGlnAlaProProProSerTrpAspGlnMetTrpLysCys
 361 GCCACCGTGTGCGCTAGGGCTCAAGCCCTTCCCCCATCGTGGGACCAGATGTGGAAGTGT
 CGGTGGCACACGGCATCCCGAGTTCGGGGAGGGGTAGCACCCCTGTCTACACCTTCACA
 LeuIleArgLeuLysProThrLeuHisGlyProThrProLeuLeuTyrArgLeuGlyAla
 421 TTGATTGCGCTCAAGCCCAACCTCCATGGGCCAACACCCCTGCTATACAGACTGGGCGCT
 AACTAAGCGGAGTTCGGGTGGGAGGTACCCGGTTGTGGGACGATATGTCTGACCCCGCA